



Conversion factors for Greenland shelf benthos: Weight-to-weight and body size-to-weight relationships

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10 **Abstract.** Climate change and biodiversity loss are rapidly transforming Arctic marine ecosystems. Benthic ecosystems on Arctic shelves are important for biodiversity and ecosystem functioning. Biomass in form of ash-free dry mass (AFDM) is often used as a proxy of ecosystem health but can be labour intensive and costly to obtain. This study addresses a key data gap by providing robust weight and body size to weight conversion factors for Arctic and boreal benthic fauna. We collected samples of common macro- and mega-benthic organisms in SE Greenland (59–67° N and 27–41° W) and calculated conversion factors for wet mass (WM) to dry mass (DM) (40 families) and to AFDM (39 families) and DM to AFDM (42 families) (<https://doi.org/10.5281/zenodo.17714017>) (Behrisch and Zworschke, 2025). To improve sampling output from non-destructive image-based sampling we also calculated conversion factors between body size (length, diameter) and weight for a subset of families (Behrisch and Zworschke, 2025). Our dataset includes several Vulnerable Marine Ecosystem (VME) indicator taxa for the Arctic region. The conversion factors for Atlantic-Arctic benthos presented here can serve as 15 the foundation for a growing database, helping to unify datasets collected using different methodologies.

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1 Introduction

Climate change and biodiversity loss are rapidly reshaping polar marine ecosystems, particularly in the Arctic, where warming occurs at nearly four times the global average rate (Pörtner et al., 2024; Rantanen et al., 2022). Arctic shelf benthos, including habitat-forming species such as sponges, soft corals, bryozoans, and hydroids, provide essential ecosystem services 25 and can transform habitats into biodiversity and blue carbon hotspots (Buhl-Mortensen et al., 2010; Grebmeier et al., 2015; Souster et al., 2024). However, our understanding of the distribution and functioning of these ecosystems remains limited. Across the Arctic, different methodologies and efforts are employed in the study of deep-sea benthic communities (Bluhm et al., 2012, 2020; Piepenburg, 2005; Piepenburg et al., 2010). This is hampering efforts to understand ecosystem dynamics on a large scale and predict their response to climate change.

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Biomass within a community is often used as an indicator of the health of an ecosystem and changes in biomass linked to changes in the abiotic environment potentially caused by anthropogenic pressure (Hewitt et al., 2005; Rombouts et al., 2013; Sherman, 1994). Commonly, biomass is measured as ash-free dry mass (AFDM), which quantifies the organic material in a sample, excluding water and inorganic components found in the wet mass of organisms (Rumohr et al., 1987). The 35 acquisition of AFDM is time and energy intensive, and, considering more novel and less invasive image-based analysis, often unobtainable.

Weight-to-weight or body size-to-weight conversion factors, derived from existing data, provide a cost-effective and practical alternative. They enable conversion of wet mass or body size measurements to AFDM and reduce the need for 40 additional sampling to inform image-based analyses. For example, biomass of a few habitat forming taxa such as corals or sponges could easily outweigh those of a highly abundant species, such as spiroid worms (Kornder et al., 2021; Marlow et al., 2024). Availability of reliable size to weight conversion factors would, thus, provide another important component for the assessment of ecosystems based on non-destructive sampling methods (Javed and Hamid, 2025; Marlow et al., 2024). Over recent years, weight-to-weight conversion factors have been calculated for various benthic macro-invertebrates across 45 several global regions (Brey et al., 2010; Gogina et al., 2020, 2022; Lappalainen and Kangas, 1975; Petersen and Curtis, 1980; Ricciardi and Bourget, 1998; Rumohr et al., 1987; Stratmann et al., 2020; Tumbiolo and Downing, 1994) and limited literature exists on body size-to-weight conversion factors for benthos (Eklöf et al., 2016). There is still a lack of data on weight-to-weight, as well as body size-to-weight conversion factors for most abundant boreal and arctic benthos, especially in regions where little is known about their distribution and ecosystem roles. Our study addresses this gap by providing 50 reliable WM-to-DM, DM-to-AFDM, and WM-to-AFDM, as well as body size-to-WM, body size-to-DM, and body size-to-AFDM conversion factors for common macro- and mega-benthic families, including VME indicator taxa from the FAO VME indicator list for the Northeast Atlantic (FAO, 2025).

The resulting data will form the basis of a growing database designed to encompass more taxa and regions over time.

55 **2 Materials & Methods**

2.1 Sampling location

Benthic macro- and mega-invertebrate samples were collected as part of a trawl-bycatch programme at the Greenland Institute of Natural Resources (GINR) (Zwerschke et al., 2025) from the 11th of July 2023 to the 2nd of August 2023 on the research vessel *Tarajøq* between SE Greenland and the Tasiilaq region (59–67° N and 27–41° W). Sampling took place at 63 60 stations, including the shelf edge and shelf slope, as well as glacial throughs and banks, spanning depths between 200 m and 580 m (Fig. 1 and Table 1).

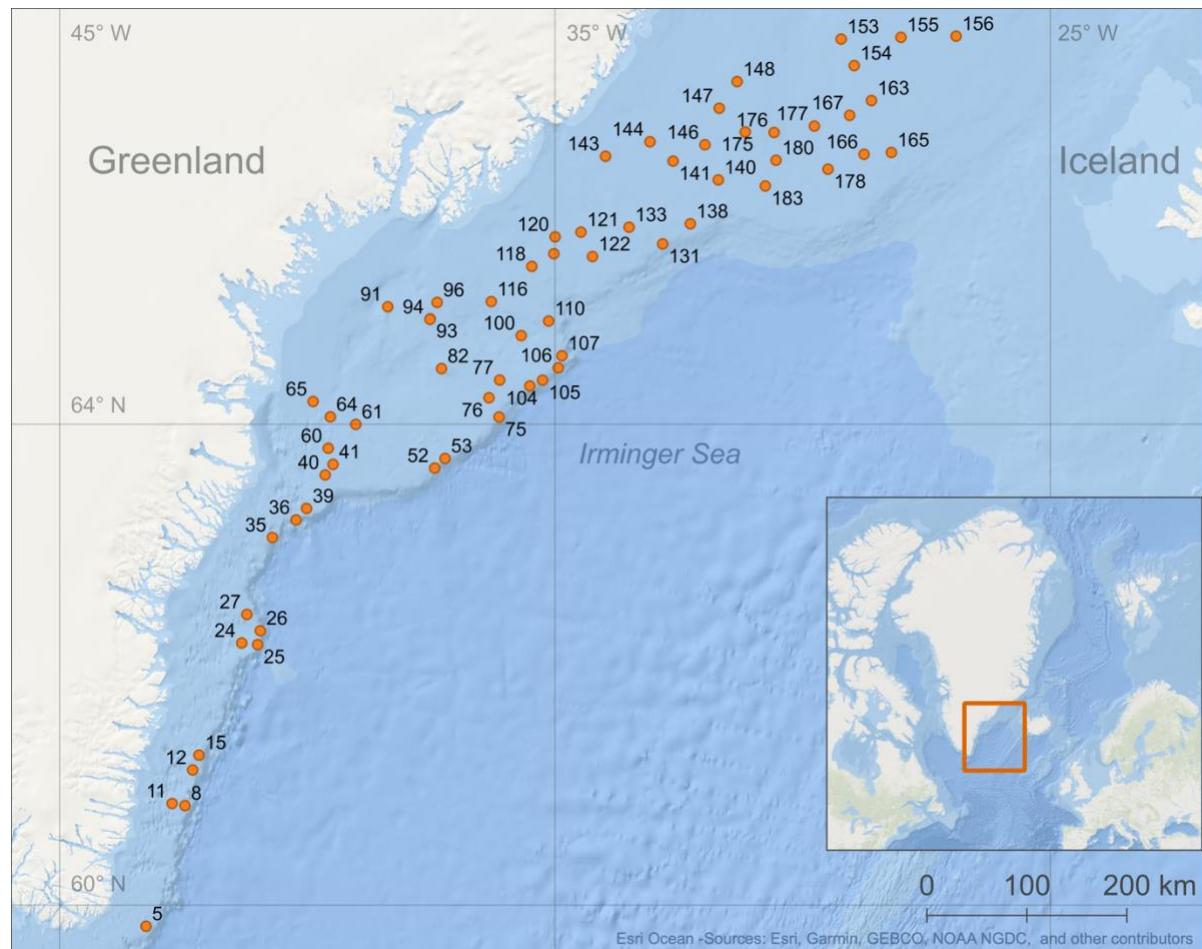


Figure 1: Map of the 63 stations (orange dots) with station numbers sampled during July and August 2023 along the East Greenland Shelf (Basemap: Esri Ocean -Sources: Esri, Garmin, GEBCO, NOAA NGDC, and other contributors).

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Table 1: Sampling station information, including station number, mean depth [m], mean bottom temperature [°C], latitude and longitude per station. Depth and bottom temperature were derived from the Furuno Marport sensor attached to the trawl.

Station	Mean depth [m]	Mean temperature [°C]	Latitude	Longitude
5	300.5	4.65	59°81.184'N	42°46.525'W
8	213.5	4.04	60°87.077'N	41°77.318'W
11	165	3.88	60°88.940'N	42°00.162'W
12	490	5.16	61°17.717'N	41°63.723'W
15	214.5	5.11	61°30.437'N	41°52.427'W
24	580.5	4.93	62°24.513'N	40°76.455'W



Station	Mean depth [m]	Mean temperature [°C]	Latitude	Longitude
25	379	4.92	62°23.245'N	40°48.278'W
26	392.5	4.75	62°34.658'N	40°43.515'W
27	193	3.74	62°48.048'N	40°67.332'W
35	363	4.75	63°10.512'N	40°22.040'W
36	354.5	4.73	63°24.612'N	39°79.898'W
39	442.5	4.70	63°33.703'N	39°61.228'W
40	183	5.41	63°60.283'N	39°28.125'W
41	195	5.58	63°68.608'N	39°14.177'W
52	270.5	4.42	63°65.707'N	37°33.622'W
53	322	4.48	63°73.272'N	37°15.282'W
60	261.5	4.46	63°81.283'N	39°22.742'W
61	289.5	4.45	63°99.913'N	38°73.851'W
64	313	4.48	64°05.777'N	39°18.978'W
65	293	5.57	64°17.658'N	39°49.633'W
75	462	5.42	64°05.635'N	36°19.150'W
76	379.5	4.61	64°20.532'N	36°37.162'W
77	261.5	5.18	64°34.188'N	36°18.130'W
82	161.5	4.99	64°43.042'N	37°21.532'W
91	556	4.18	64°89.973'N	38°17.062'W
93	293	NA	64°80.835'N	37°41.465'W
94	290.5	4.05	64°80.447'N	37°41.927'W
96	268	4.31	64°93.225'N	37°29.207'W
100	532.5	3.71	64°68.143'N	35°79.923'W
104	343	4.66	64°29.672'N	35°64.630'W
105	356	4.27	64°34.083'N	35°42.017'W
106	431	4.70	64°43.552'N	35°14.085'W
107	456	3.74	64°52.795'N	35°07.480'W
110	321	5.11	64°79.160'N	35°31.167'W



Station	Mean depth [m]	Mean temperature [°C]	Latitude	Longitude
116	461	3.68	64°93.838'N	36°33.088'W
118	236.5	4.15	65°20.155'N	35°60.993'W
119	290	3.35	65°29.575'N	35°21.882'W
120	298	NA	65°42.080'N	35°19.630'W
121	342.5	3.68	65°45.550'N	34°73.752'W
122	313	3.76	65°27.505'N	34°53.230'W
131	288	5.49	65°36.842'N	33°28.608'W
138	314.5	3.68	65°51.680'N	32°79.417'W
140	232.5	2.56	65°83.763'N	32°29.770'W
141	332	2.46	65°97.391'N	33°10.083'W
143	261	2.48	66°00.898'N	34°30.135'W
144	309	2.47	66°11.327'N	33°51.083'W
146	272	2.34	66°09.182'N	32°53.517'W
147	334.5	1.48	66°35.278'N	32°28.063'W
148	301	1.61	66°54.197'N	31°96.273'W
153	341.5	1.19	66°84.052'N	30°11.373'W
154	287	1.89	66°65.483'N	29°88.236'W
155	287.5	1.13	66°85.302'N	29°05.215'W
156	356.5	0.96	66°86.088'N	28°07.283'W
163	317	0.82	66°40.780'N	29°57.583'W
165	294	0.30	66°03.616'N	29°22.166'W
166	300	-0.02	66°02.290'N	29°70.723'W
167	354.5	0.87	66°30.260'N	29°96.320'W
175	276.5	2.12	66°18.340'N	31°81.805'W
176	447	1.09	66°17.993'N	31°30.643'W
177	489	0.77	66°22.575'N	30°59.023'W
178	439	0.82	65°91.541'N	30°34.809'W
180	430.5	1.19	65°98.001'N	31°27.038'W



Station	Mean depth [m]	Mean temperature [°C]	Latitude	Longitude
183	369	1.66	65°79.328'N	31°46.166'W

2.2 Sampling procedures and processing

70 2.2.1 Benthic sample collection with cosmos trawl

At each station a cosmos bottom trawl (twin-body trawl) was deployed and towed for 15 min. The trawl consisted of a funnel section, splitting into two extension sections and two codends. The funnel section and extension sections had a mesh size of ~ 4.45 cm. The net was held open by two heavy otter doors and the mouth of the open net had a size of approximately 1.14 m (side) x 3 m (headline). The station start and end location was recorded and temperature [°C], as well as depth [m] 75 recorded every 2 minutes based on the readings of a MARPORT Trawl Explorer Pro ® sensor. Benthic fauna from each trawl were identified to lowest possible resolution based on their morphology and updated towards the most current nomenclature via the World Register of Marine Species (WoRMS Editorial Board, 2025). Most samples were identified to the genus level. However, some could only be identified to family level.

80 For conversion factors, sampling aimed at collecting 10 individuals per benthic family, with priority given to taxa that serve as VME indicators and appeared to be dominant components (e.g. high density of individual in catch) of the habitat. For each taxon, we included a balanced variety of body sizes and morphologies and avoided targeting specific body sizes or inclusion of atypical morphologies, such as disproportionately large shells or spicules. Fragile habitat-dominating species 85 (such as bryozoans, like *Hornera lichenoides* or the glass sponge, *Asconema* sp.), often obtained as fragments, were still included in sample collection, due to their significant presence or ecological significance across stations. The size and biomass of each fragment were measured and processed in the same way as for complete organisms, with conversion factors derived from the fragment measurements rather than from whole organisms. Shells or housings that are produced by taxa such as bivalve shells or echinoderm tests were included in their weight. Individuals that were heavy (> 100 g), or large (e.g., round sponges of the genera *Craniella* sp. and *Geodia* sp.) were sub-sampled. To obtain reliable sub-samples, a cross-section 90 containing a variety of different tissues from an individual was taken. All full samples, fragments and sub-samples were stored in a - 80 °C freezer until further processing in the laboratory.

2.2.2 Assessing the body size, WM, DM and AFDM

95 In the laboratory, the WM [g] of each full individual was estimated. In addition, the body size [mm] of the full individual was measured using species specific morphometric features (Table 2). Afterwards, samples were dried at 70 °C for at least



48 h or until a constant DM [g] was reached and subsequently burned at 450 °C for 24 h. Ash mass (AM) [g] was weighed and subtracted from the DM to calculate the AFDM [g] (weight accuracy of ± 0.0001 g) of a sample. For subsamples AFDM was extrapolated based on the WM of the full sample.

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Table 2: Morphometrics indicating which body part was measured as a proxy for individual size.

Morphometrics	Family
Carapax length	Colossendeidae, Lithodidae, Nymphonidae
Body diameter	Ancorinidae, Coelosphaeridae, Geodiidae, Liponematidae, Polymastiidae, Strongylocentrotidae, Styelidae, Tetillidae, Theneidae
Disk diameter	Gorgonocephalidae, Ophiopholidae, Ophiacanthidae
Outer diameter (across disk & arms)	Benthopectinidae, Echinasteridae, Poraniidae, Pterasteridae, Solasteridae
Body length	Actinostolidae, Aegidae, Aglaopheniidae, Antedonidae, Antholobidae, Aphroditidae, Bubaridae, Buccinidae, Cancellothyrididae, Capnellidae, Chalinidae, Coralliidae, Didemnidae, Euplectellinae, Gofkingiidae, Goniasteridae, Hanleyidae, Hormathiidae, Horneridae, Laetmogonidae, Lafoeidae, Molpadiidae, Mycalidae, Phidoloporidae, Polynoidae, Rossellidae, Sertulariidae

2.3 Statistical Analysis

All statistical analyses were conducted in R version 2024.04.0+735 (R Core Team, 2022). Analyses were performed at the 105 family level to ensure sufficiently large sample sizes. Conversion factors between WM and DM, WM and AFDM, and DM and AFDM were calculated for each taxonomic family as the slope of the regression between each pair of body mass variables. In addition, conversion factors between body size and WM, DM, and AFDM were similarly calculated from the slopes of the regressions between each pair of variables. For body size and mass metrics, homogeneity of variance and normality were tested using Levene's test and the Shapiro-Wilk test, respectively (Fox, 1997, 2011; Fox and Weisberg, 110 2019; Levene, 1960; Shapiro and Wilk, 1965). To evaluate the strength and reliability of the weight-to-weight and body size-to-weight relationships, and thereby to assess the validity of the derived conversion factors, Spearman rank correlation tests were calculated between WM (predictor variable) and DM (response variable), DM (predictor) and AFDM (response), and



WM (predictor) and AFDM (response), and in addition between biomass measures (WM, DM, AFDM; response variables) and body size (predictor variable) (Dodge, 2008; Glasser and Winter, 1961; Spearman, 1904).

115 3 Data availability

All raw measurement data, primary result tables (conversion factors), and R scripts used for data processing and analysis are available in the open repository Zenodo (<https://doi.org/10.5281/zenodo.17714017>) (Behrisch and Zworschke, 2025). All quality-checked measurements are included without selective exclusion. The supplementary tables, which include the Spearman rank correlation test results, are provided in the publication.

120 4 Results

A total of 492 samples were collected across 63 stations. In total, 43 benthic families were sampled for which weight-to-weight, as well as body size-to-weight conversion factors could be calculated (Table 3 & 4) and the relationships among WM, DM, AFDM, and body size measurements could be evaluated (Table S1 & S2) (Behrisch and Zworschke, 2025).

4.1 Weight-to-weight relationships and conversion factors

125 For 38 of the 43 families, including key VME taxa such as demosponges (Theneidae, Ancorinidae, Tetillidae, Geodiidae, Mycalidae, Polymastiidae), feather stars (Antedonidae), glass sponges (Rossellidae), cauliflower soft corals (Capnellidae), gorgonian soft corals (Coralliidae) and bryozoans (Horneridae, Phidoloporidae), Spearman rank correlations show that WM serves as a reliable predictor of DM and AFDM (FAO, 2025) (Table 3 and S1) (p -value > 0.05). A significant relationship between DM and AFDM was present in 42 families (p -value > 0.05) (Table 3 and S1). For several families in which the 130 Spearman rank correlations were not significant, the conversion factors derived from regression still accounted for a substantial proportion of the variance ($R^2 > 0.5$). This indicates that, even when the Spearman rank tests show weaker monotonic relationships, linear allometric models can still provide reliable weight-to-weight conversion factors for these families.

135 Reliable conversion factors were calculated for key VME indicator taxa such as families of glass sponges, feather stars, demosponges, cauliflower soft corals, gorgonian soft corals and bryozoans (FAO, 2025). Reliable conversion factors refer to factors with an R^2 value greater than 0.5. Reliable WM-to-DM conversion factors were calculated for 40 of the 43 families, demonstrating that WM is a good predictor of DM in these cases (Table 3). Only regressions for the families Bubaridae (demosponges, $R^2 = 0.128$), Colossendeidae (sea spiders, $R^2 = 0.490$), and Strongylocentrotidae (sea urchins, $R^2 = 0.230$) 140 were found to have R^2 values below 0.5. Except for the family Bubaridae ($R^2 = 0.454$), reliable DM-to-AFDM conversion factors were determined for the remaining 42 families (Table 3). Most families ($n = 39$) had a good relationship between



WM-to-AFDM (Table 3). However, the families Colossendeidae ($R^2 = 0.459$), Golfingiidae (peanut worms, $R^2 = 0.294$), Polynoidae (scale worms, $R^2 = 0.409$), and Strongylocentrotidae ($R^2 = 0.213$) did not have a linear relationship between WM and AFDM.



145 **Table 3: Weight-to-weight [g] conversion factors (CF) between wet mass (WM), dry mass (DM), and ash-free dry mass (AFDM) were calculated for 43 benthic families, including 95 % confidence intervals (CI) and R² values. For each family, the column “N” represents the number of total included individuals. Conversion factors of families with significant Spearman rank correlation tests (p-value < 0.05) are shown in bold text. The full table of Spearman rank correlation results is provided in the Supplements (Table S1).**

Phylum	Family	Genus	WM-to- AFD M CF	WM-to- AFD M R ²	WM-to- AFD M CI	DM-to- AFD M CF	DM-to- AFD M R ²	DM-to- AFD M CI	WM-to- DM CF	WM-to- DM R ²	WM-to- DM CI	N
Annelida	Aphroditidae	<i>Laetmonice</i> sp., one more not further identified	0.126	0.962	0.103 - 0.148	0.655	0.990	0.596 - 0.715	0.192	0.968	- 0.223	9
Annelida	Golfingiidae	<i>Golfingia</i> sp.	0.020	0.294	-0.037 - 0.076	0.472	0.947	0.266 - 0.677	0.054	0.52	0.042 - 0.15	5
Annelida	Polynoidae	<i>Eunoe</i> sp., <i>Phyllodoce</i> sp.	0.074	0.409	0.007 - 0.141	0.745	0.840	0.500 - 0.991	0.121	0.722	- 0.178	1
Arthropoda	Aegidae	<i>Aegiochus</i> sp.	0.280	0.806	0.214 - 0.346	0.765	0.964	0.694 - 0.836	0.384	0.924	- 0.437	2
Arthropoda	Colosseididae	<i>Colossendeis</i> sp.	0.240	0.459	-0.553 - 1.034	0.921	0.999	0.826 - 1.015	0.270	0.490	0.567 - 1.106	4
Arthropoda	Lithodidae	<i>Lithodes</i> sp.	0.227	1.000	0.220 - 0.233	0.656	1.000	0.633 - 0.678	0.346	1.000	- 0.348	3
Arthropoda	Nymphonidae	<i>Nymphon</i> sp.	0.260	0.740	0.144 - 0.376	0.879	0.994	0.829 - 0.929	0.293	0.733	- 0.427	1
Bivalvia	Cancellytididae	<i>Terebratulina</i> sp.	0.018	0.666	0.008 - 0.028	0.054	0.793	0.031 - 0.076	0.280	0.584	- 0.472	1
Bryozoa	Horneridae	<i>Hornera</i> sp.	0.045	0.992	0.042 - 0.048	0.046	0.992	0.042 - 0.049	0.977	0.99	- 0.899	1
Bryozoa	Phidoloporidae	<i>Reteporella</i> sp.	0.037	0.872	0.027 - 0.048	0.043	0.859	0.030 - 0.056	0.870	0.991	- 0.931	1
Chordata	Didemnididae	<i>Didemnum</i> sp.	0.048	0.760	0.026 - 0.070	0.329	0.766	0.181 - 0.477	0.139	0.904	- 0.176	1
Chordata	Styelidae	<i>Kukenthalia</i> sp.	0.112	0.813	-0.573 - 0.798	0.686	0.984	-0.433 - 1.804	0.171	0.901	- 0.892	3
Cnidaria	Actinostolididae	<i>Stomphia</i> sp.	0.095	1.000	0.095 - 0.096	0.846	1.000	0.818 - 0.873	0.113	1.000	- 0.108	3



Phylum	Family	Genus	WM-to- AFD M CF	WM-to- AFD M R ²	WM-to- AFD M CI	DM-to- AFD M CF	DM-to- AFD M R ²	DM-to- AFD M CI	WM-to- DM CF	WM-to- DM R ²	WM-to- DM CI	N
Cnidaria	Aglaopheniidae	<i>Aglaophenopsis</i> sp.	0.046	0.534	-0.499 -0.591	0.597	0.998	0.223	0.080	0.583	0.782	3
Cnidaria	Capnelidae	<i>Drifa</i> sp., <i>Duva</i> sp., <i>Pseudodrifa</i> sp.	0.092	0.931	0.083 -0.101	0.378	0.897	-	0.330	0.993	0.231 -0.245	3 2
Cnidaria	Coralliidiae	<i>Paragorgia</i> sp.	0.054	0.980	0.040 -0.068	0.141	0.985	-	0.109	0.999	-	5
Cnidaria	Hormathidae	<i>Actinauge</i> sp., <i>Hormathia</i> sp.	0.089	0.911	0.076 -0.103	0.755	0.992	-	0.723	0.954	-	2 1
Cnidaria	Liponematidae	<i>Liponema</i> sp.	0.151	0.992	0.135 -0.167	0.805	0.995	-	0.740	0.998	-	7
Cnidaria	Sertulariidae	<i>Diphasia</i> sp., <i>Thuiaria</i> sp.	0.301	0.993	0.253 -0.349	0.640	1.000	-	0.869	0.996	-	5
Echinodermata	Antedonidae	<i>Heliometra</i> sp.	0.063	0.883	0.031 -0.095	0.267	0.923	-	0.160	0.981	-	6
Echinodermata	Benthoplectinidae	<i>Pontaster</i> sp.	0.097	0.969	0.081 -0.112	0.264	0.964	-	0.219	0.988	-	9
Echinodermata	Echinasteridae	<i>Henricia</i> sp.	0.222	0.997	0.212 -0.231	0.707	0.998	-	0.678	0.998	-	1 0
Echinodermata	Gorgonocephalidae	<i>Gorgonocephalus</i> sp.	0.167	0.949	0.122 -0.211	0.383	0.916	-	0.250	0.994	-	7
Echinodermata	Laetmogonidae	<i>Laetmogone</i> sp.	0.020	0.963	0.017 -0.023	0.216	0.851	-	0.142	0.775	-	1 0
Echinodermata	Molpadiidae	<i>Molpadia</i> sp.	0.026	0.962	0.022 -0.031	0.195	0.979	-	0.172	0.962	-	1 0



Phylum	Family	Genus	WM-to-AFD M CF	WM-to-AFD M R ²	WM-to-AFD M CI	DM-to-AFD M CF	DM-to-AFD M R ²	DM-to-AFD M CI	WM-to-DM CF	WM-to-DM R ²	WM-to-DM CI	N
Echinodermata	Ophiacanthidae	<i>Ophiosabine</i> sp.	0.062	0.669	0.018 - 0.105	0.168	0.819	0.089	0.351	0.747	0.147	8
Echinodermata	Ophiodolidae	<i>Ophiopholis</i> sp.	0.101	0.931	0.080 - 0.121	0.186	0.977	0.165	0.524	0.899	0.392	1
Echinodermata	Poraniidae	<i>Poraniomorpha</i> sp.	0.083	0.951	0.067 - 0.098	0.340	0.979	0.300	0.242	0.961	0.202	1
Echinodermata	Pterasteridae	<i>Diplopteraster</i> sp., <i>Hymenaster</i> sp., <i>Pteraster</i> sp.	0.132	0.982	0.125 - 0.140	0.681	0.998	0.667	0.195	0.992	0.188	2
Echinodermata	Solasteridae	<i>Crossaster</i> sp.	0.118	0.959	0.103 - 0.132	0.456	0.975	0.412	0.258	0.990	0.243	1
Echinodermata	Strongylocentrotidae	<i>Strongylocentrotus</i> sp.	0.011	0.213	-0.001 - 0.022	0.140	0.901	0.114	0.077	0.230	0.000	1
Mollusca	Buccinidae	not further identified	0.162	0.962	0.135 - 0.188	0.330	0.866	0.224	0.458	0.967	0.389	1
Mollusca	Hanleyidae	<i>Hanleya</i> sp.	0.176	0.981	0.156 - 0.196	0.432	0.947	0.435	0.395	0.977	0.526	0
Porifera	Ancorinidae	<i>Stelletta</i> sp.	0.120	0.961	0.100 - 0.139	0.554	0.981	0.348	0.217	0.987	0.345	1
Porifera	Bubaridae	<i>Phakellia</i> sp.	1.867x10⁻⁴	0.867	-0.001 - 0.001	0.214	0.454	-0.274	2.260x10⁻⁴	0.128	0.007	3
Porifera	Coelosphaeridae	<i>Histodermella</i> sp.	0.080	0.763	0.060 - 0.100	0.332	0.792	0.255	0.243	0.979	0.227	2
Porifera	Euplectellinae	<i>Euplectella</i> sp.	0.001	0.696	-0.001 - 0.004	0.280	0.998	0.410	0.245	0.657	0.005	4
								0.315	0.004	-	0.014	



Phylum	Family	Genus	WM-to-AFD M CF	WM-to-AFD M R ²	WM-to-AFD M CI	DM-to-AFD M CF	DM-to-AFD M R ²	DM-to-AFD M CI	WM-to-DM CF	WM-to-DM R ²	WM-to-DM CI	N	
Porifera	Geodidae	<i>Geodia</i> sp.	0.101	0.871	0.119	0.539	0.983	0.506	-	0.187	0.887	0.155	2
								0.573				0.219	1
								0.511				0.206	1
Porifera	Mycalidae	<i>Mycale</i> sp.	0.112	0.995	0.106 - 0.118	0.517	1.000	-	0.217	0.996	-	0.227	1
								0.523				0.227	1
Porifera	Polymastidiidae	<i>Polymastia</i> sp., <i>Tentorium</i> sp.	0.113	0.993	0.110 - 0.116	0.580	0.994	-	0.194	0.995	-	0.189	4
								0.566				0.198	4
Porifera	Rossellidae	<i>Asconema</i> sp.	0.022	0.899	0.017 - 0.026	0.206	0.675	-	0.081	0.794	-	0.053	1
								0.111				0.108	3
Porifera	Tetillidae	<i>Craniella</i> sp.	0.145	0.990	0.133 - 0.157	0.592	0.984	-	0.244	0.997	-	0.233	1
								0.531				0.254	0
Porifera	Theneidae	<i>Thenea</i> sp.	0.071	0.890	0.050 - 0.091	0.385	0.957	-	0.188	0.978	-	0.165	1
								0.653				0.211	0
								0.318					
								0.451					



4.2 Body size-to-weight relationships and conversion factors

155 Spearman ranks correlations indicate that body size predicted biomass measures in most families: 31 of 43 families showed significant body size to WM correlations, and 32 families showed significant body size to DM and AFDM relationships (FAO, 2025) (Table 4 and S2) (p -value > 0.05). As with weight-to-weight models, regression explained variance well ($R^2 > 0.5$) even where rank correlations were insignificant for families, indicating that allometric models provide robust body size to biomass conversion factors.

160 Reliable body size-to-WM and size-to-DM conversion factors ($R^2 > 0.5$) were obtained for 32 of the 43 families, while size-to-AFDM conversion factors were reliable for 31 families (Table 4). These conversion factors include key VME indicator taxa such as feather stars, cauliflower soft corals, gorgonian soft corals, demosponges, bryozoans, and, for size-to-DM, also glass sponges. Conversion factors were unreliable ($R^2 < 0.5$) for body size-to-WM and body size-to-DM in 11 families, and for body size-to-AFDM in 12 families. (Table 4). For some families belonging to the classes of hydroids, feather stars, 165 bristle worms and to the order of anemones, the body size-to-weight conversion factor was negative (Table 4). For these families, the R^2 of the conversion factors was lower than 0.5 and Spearman Ranks correlations tests indicated no significant correlation between body size and biomass.

170 It should be noted that significant Spearman rank correlations, as well as conversion factors relating to weight-to-weight and body size-to-weight relationships for the families Actinostolidae (sea anemones), Aglaopheniidae (hydroids), Bubaridae, Lithodidae (king crabs), and Styelidae (tunicates) should be interpreted with caution, as they were calculated from only three samples.

175 Several factors could explain the lack of a significant linear relationship between body mass metrics, as well as body size and body mass. For families like Strongylocentrotidae, for example, gonad production or recent feeding events, constitutes a major component of their body mass, potentially distorting any weight to size relationships (Blicher et al., 2007). Furthermore, sea cucumbers can expel their internal organs (evisceration) as defense strategy, though strategies vary across sea cucumbers (Emson and Wilkie, 1980; García-Arrarás and Greenberg, 2002; Hyman, 1955). The genera *Laetmogone* sp. and *Molpadia* sp. may use this defense during the sampling event, which could impact biomass estimates in the present 180 study. Non-linear and colonial growth patterns, such as found in some sponges, hydroids and bryozoans, may limit the use of size for specific taxa as a biomass indicator. Additionally, it was often impossible to collect entire specimens of taxa with delicate structures that easily break during sampling, such as feather stars or sponges of the family Bubaridae. This may have introduced a certain degree of bias in the results.



185 **Table 4: Body size [mm]-to-weight [g] (wet mass (WM), dry mass (DM), and ash-free dry mass (AFDM)) conversion factors (CF) were calculated for 43 benthic families, including 95 % confidence intervals (CI) and R² values. For each family, the column “N” represents the number of samples used to determine the conversion factors, along with the genera included in the calculations. The column “Morphometrics” indicates the body part measured to evaluate organism size, which varies according to the morphology of organisms within a family. Conversion factors of families with significant Spearman rank correlation tests (p-value < 0.05) are shown in bold text. The full table of Spearman rank correlation results is provided in the Supplements (Table S2).**

Phylum	Family	Genus	Morphometrics	Size-to-WM CF	Size-to-WM R ²	Size-to-WM CI	Size-to-DM CF	Size-to-DM R ²	Size-to-DM CI	Size-to-AFD M CF	Size-to-AFD M R ²	Size-to-AFD M CI	N
Ann elida	Aphro ditidae	<i>Laetmonice</i> sp., one more not further identified	Body length	0.192	0.286	0.079 - 0.463	0.028	0.15 9 - 0.08 5	0.02 9 - 0.08 5	0.019	0.179	-0.018 - 0.057	9
Ann elida	Golfin giidae	<i>Golfingia</i> sp.	Body length	0.069	0.830	0.012 - 0.126	0.005	0.72 7	0.00 1 - 0.01	0.002	0.532	-0.001 - 0.006	5
Ann elida	Polynoidae	<i>Eunoe</i> sp., <i>Phyllodoce</i> sp.	Body length	0.031	0.210	0.018 - 0.08	0.001	0.00 6	0.00 7 - 0.00 8	-0.002	0.096	-0.008 - 0.003	11
Arthropoda	Aegidae	<i>Aegiochus</i> sp.	Body length	0.131	0.846	0.104 - 0.158	0.051	0.81 9	0.04 0 - 0.06 3	0.039	0.758	0.028 - 0.049	21
Arthropoda	Colossendeidae	<i>Colossendeis</i> sp.	Carpax length	0.065	0.848	0.286 - 0.417	0.027	0.78 4	0.15 5 - 0.21	0.025	0.753	-0.155 - 0.205	4
Arthropoda	Lithodidae	<i>Lithodes</i> sp.	Carpax length	2.516	0.980	2.036 - 7.067	0.870	0.98 0	0.69 8 - 2.43 7	0.570	0.980	-0.477 - 1.618	3
Arthropoda	Nymphonidae	<i>Nymphon</i> sp.	Carpax length	0.019	0.740	0.011 - 0.028	0.005	0.50 5	0.00 1 - 0.01 0	0.005	0.537	0.002 - 0.008	11
Brachiopoda	Cancellothyrididae	<i>Terebratulina</i> sp.	Body length	0.019	0.035	0.061 - 0.099	0.010	0.07 4	0.01 9 - 0.03 9	0.001	0.017	-0.001 - 0.002	10



Phylum	Family	Genus	Morphometrics	Size-to-WM CF	Size-to-WM R ²	Size-to-WM CI	Size-to-DM CF	Size-to-DM R ²	Size-to-DM CI	Size-to-AFD M CF	Size-to-AFD M R ²	Size-to-AFD M CI	N	
Bryozoa	Horneridae	<i>Hornera</i> sp.	Body length	0.035	0.249	0.014	0.036	0.276	0.012	0.002	0.268	-0.001	10	
						-			0.083			0.004		
Bryozoa	Phidoloporidae	<i>Reteporella</i> sp.	Body length	0.083	0.751	0.047	0.073	0.758	0.042	0.003	0.586	0.001	11	
						-	0.119		0.103			0.005		
Chordata	Didemnidiae	<i>Didemnum</i> sp.	Body length	0.067	0.285	-0.02	0.011	0.358	0.001	0.004	0.361	0.000	10	
						-	0.154		0.023			0.009		
Chordata	Styelidae	<i>Kukenthalia</i> sp.	Body diameter	0.012	0.325	0.211	0.003	0.639	0.026	0.002	0.756	-0.014	3	
						-	0.235		0.032			0.019		
Cnidaria	Actinostolidae	<i>Stomphia</i> sp.	Body length	0.576	1.000	0.513	0.065	1.000	0.060	0.055	1.000	0.049	3	
						-	0.639		0.069			0.061		
Cnidaria	Aglaopheniidae	<i>Aglaophenopsis</i> sp.	Body length	-	0.038	0.333	-	0.712	-	0.028	-0.004	0.959	-0.015	3
						-	0.637		0.017			0.007		
Cnidaria	Capnelidae	<i>Drifa</i> sp., <i>Duva</i> sp., <i>Pseudodrifa</i> sp.	Body length	0.482	0.511	0.310	0.114	0.494	0.072	0.052	0.617	0.037	3	
						-	0.655		0.157			0.067		
Cnidaria	Corallidiidae	<i>Paragorgia</i> sp.	Body length	19.217	0.875	5.862	7.370	0.871	2.160	1.072	0.915	0.474	5	
						-	32.572		0.125			1.671		
Cnidaria	Hormathiidae	<i>Actinauge</i> sp., <i>Hormathia</i> sp.	Body length	0.911	0.767	0.669	0.103	0.641	0.066	0.074	0.578	0.044	21	
						-	1.151		0.140			0.104		
Cnidaria	Liponematidae	<i>Liponema</i> sp.	Body diameter	1.057	0.959	0.862	0.010	0.018	0.075	-0.010	0.019	-0.061	9	
						-	1.253		0.055			0.045		



Phylum	Family	Genus	Morphometrics	Size-to-WM CF	Size-to-WM R ²	Size-to-WM CI	Size-to-DM CF	Size-to-DM R ²	Size-to-DM CI	Size-to-AFD M CF	Size-to-AFD M R ²	Size-to-AFD M CI	N
Cnidaria	Sertulariidae	<i>Diphasia</i> sp., <i>Thuiaria</i> sp.	Body length	0.007	0.920	0.003 - 0.012	0.004	0.94 4	0.00 2 - 0.00 5	0.002	0.952	0.001 - 0.003	5
Echi node rmat a	Antedonidae	<i>Helio metra</i> sp.	Body length	0.029	0.549	0.001 - 0.06	0.003	0.04 5	0.01 2 - 0.01 8	-0.002	0.045	-0.013 - 0.009	7
Echi node rmat a	Benthopectinidae	<i>Pontaster</i> sp.	Outer diameter	0.108	0.762	0.059 - 0.157	0.037	0.66 7	0.01 6 - 0.05 9	0.009	0.442	0.001 - 0.017	10
Echi node rmat a	Echinasteridae	<i>Henricia</i> sp.	Outer diameter	0.089	0.969	0.077 - 0.101	0.028	0.97 3	0.02 5 - 0.03 1	0.020	0.959	0.016 - 0.023	11
Echi node rmat a	Gorgonocephalidae	<i>Gorgonocephalus</i> sp.	Disk diameter	0.152	0.346	0.088 - 0.393	0.060	0.28 4	0.04 9 - 0.16 7	0.030	0.454	-0.008 - 0.068	7
Echi node rmat a	Laetmogonidae	<i>Laetmogone</i> sp.	Body length	0.601	0.930	0.467 - 0.736	0.047	0.76 6	0.02 6 - 0.06 9	0.012	0.884	0.008 - 0.015	10
Echi node rmat a	Molpadiidae	<i>Molpadia</i> sp.	Body length	0.890	0.963	0.747 - 1.033	0.120	0.93 3	0.09 4 - 0.14 6	0.023	0.909	0.017 - 0.029	10
Echi node rmat a	Ophiacanthidae	<i>Ophiosabine</i> sp.	Disk diameter	0.066	0.067	-0.18 - 0.311	0.005	0.00 3	0.09 8 - 0.10 8	0.005	0.079	-0.013 - 0.024	8
Echi node rmat a	Ophiodolidae	<i>Ophiopholis</i> sp.	Disk diameter	0.219	0.607	0.086 - 0.352	0.121	0.60 7	0.04 8 - 0.19 5	0.024	0.675	0.011 - 0.037	11
Echi node rmat a	Poraniidae	<i>Poraniomorpha</i> sp.	Outer diameter	0.584	0.903	0.428 - 0.739	0.148	0.95 5	0.12 2 - 0.17 4	0.051	0.943	0.040 - 0.061	10



Phylum	Family	Genus	Morphometrics	Size-to-WM CF	Size-to-WM R ²	Size-to-WM CI	Size-to-DM CF	Size-to-DM R ²	Size-to-DM CI	Size-to-AFD M CF	Size-to-AFD M R ²	Size-to-AFD M CI	N
Echinodermata	Pterasteridae	<i>Diplopteraster</i> sp., <i>Hymenaster</i> sp., <i>Pteraster</i> sp.	Outer diameter	1.463	0.917	-	0.285	0.90	7 - 8	0.193	0.894	-	2
						1.648			0.324			0.221	6
Echinodermata	Solasteridae	<i>Crossaster</i> sp.	Outer diameter	0.126	0.862	0.095 - 0.156	0.033	0.880	6 - 0.040	0.015	0.824	0.011 - 0.019	15
Echinodermata	Strongylocentrotidae	<i>Strongylocentrotus</i> sp.	Body diameter	0.374	0.430	0.137 - 0.610	0.082	0.816	1 - 0.104	0.012	0.796	0.009 - 0.015	17
Mollusca	Buccinidae	not further identified	Body length	0.450	0.978	0.395 - 0.505	0.206	0.945	5 - 0.246	0.073	0.947	0.059 - 0.087	10
Mollusca	Hanleyidae	<i>Hanleya</i> sp.	Body length	0.482	0.945	0.387 - 0.577	0.191	0.926	7 - 0.235	0.084	0.906	0.062 - 0.106	10
Porifera	Ancorinidae	<i>Stelletta</i> sp.	Body diameter	17.564	0.822	10.900 - 24.228	3.802	0.807	2.286 - 5.318	2.129	0.809	1.286 - 2.971	10
Porifera	Bubaridae	<i>Phakellia</i> sp.	Body length	2.481	0.975	2.597 - 7.56	0.001	0.252	0.017 - 0.018	0.001	0.955	-0.001 - 0.002	3
Porifera	Coelosphaeridae	<i>Histodermella</i> sp.	Body diameter	0.112	0.577	0.069 - 0.156	0.026	0.537	0.015 - 0.038	0.010	0.419	0.004 - 0.013	23
Porifera	Euplectellinae	<i>Euplectella</i> sp.	Body length	2.276	0.783	1.371 - 5.924	0.011	0.630	0.015 - 0.038	0.003	0.653	-0.004 - 0.01	4
Porifera	Geodidae	<i>Geodia</i> sp.	Body diameter	67.517	0.832	52.959 - 82.074	11.849	0.653	7.699 - 15.998	6.332	0.631	4.006 - 8.658	21
Porifera	Myciliidae	<i>Mycale</i> sp.	Body length	1.593	0.833	1.056 - 2.130	0.337	0.791	0.206 - 0.468	0.173	0.783	0.104 - 0.242	11



Phylum	Family	Genus	Morphometrics	Size-to-WM CF	Size-to-WM R ²	Size-to-WM CI	Size-to-DM CF	Size-to-DM R ²	Size-to-DM CI	Size-to-AFD M CF	Size-to-AFD M R ²	Size-to-AFD M CI	N
Porifera	Polymastiidae	<i>Polymastia</i> sp., <i>Tentorium</i> sp.	Body diameter	2.565	0.723	2.071 - 3.060	0.505	0.74 3 3	0.41 3 - 0.59 8	0.285	0.701	0.227 - 0.344	4 4
Porifera	Rossellidae	<i>Asconema</i> sp.	Body length	0.086	0.191	0.031 - 0.202	0.013	0.55 3 3	0.00 5 - 0.02 1	0.002	0.170	-0.001 - 0.005	1 3
Porifera	Tetillidae	<i>Craniella</i> sp.	Body diameter	3.779	0.922	2.883 - 4.675	0.921	0.92 0 0	0.70 0 - 1.14 2	0.546	0.907	0.404 - 0.688	1 0
Porifera	Theneidae	<i>Thenea</i> sp.	Body diameter	0.990	0.742	0.514 - 1.466	0.184	0.70 9 9	0.08 8 - 0.28 0	0.063	0.532	0.015 - 0.111	1 0



5 Conclusion

This study provides robust weight-to-weight conversion factors and body size-to-weight conversion factors for some of the 195 most dominant boreal and Arctic shelf megafauna, including critical Vulnerable Marine Ecosystem (VME) taxa, for the first time. We demonstrate that WM is a reliable predictor of DM and AFDM for most families in this study. In addition, we establish robust relationships between body size and biomass, showing that body size can serve as a reliable proxy for WM, DM, and AFDM in many benthic families. Together, these conversion factors strengthen the potential for non-destructive sampling approaches, enabling effective monitoring of ecosystem changes over time while minimizing disturbance to 200 sensitive habitats. This dataset provides a baseline, which can be repeatedly updated and to which further existing data can be added to improve the taxonomic resolution and its usefulness for future work. As pressure on scientific resources increases and the ethics of destructive sampling of the few remaining pristine habitats are more and more questionable, datasets like these will help to streamline and improve future sampling and monitoring events in the Arctic (Javed and Hamid, 2025; Marlow et al., 2024).

205 Authors contributions

Johanna Behrisch led the writing of the original draft, curated the data, performed formal analyses, and created visualizations. Nadescha Zworschke contributed to writing through review and editing, secured funding, curated data, and did the conceptualization of the study.

Competing interests

210 The authors declare that they have no conflict of interest.

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References

Behrisch, J. and Zworschke, N.: Conversion factors for Greenland shelf benthos: Weight-to-weight and body size-to-weight relationships [Data set], Zenodo, <https://doi.org/10.5281/zenodo.17714017>, 2025.

225

Blicher, M. E., Rysgaard, S., and Sejr, M. K.: Growth and production of sea urchin *Strongylocentrotus droebachiensis* in a high-Arctic fjord, and growth along a climatic gradient (64 to 77 °N), *Mar Ecol Prog Ser*, 341, 89–102, <https://doi.org/10.3354/meps341089>, 2007.

230

Bluhm, B. A., Ambrose, W., Bergmann, M., Clough, L., Gebruk, A., Hasemann, C., Iken, K., Klages, M., Macdonald, I., Renaud, P., Schewe, I., Soltwedel, T., and Włodarska-Kowalcuk, M.: Diversity of the arctic deep-sea benthos, *Marine Biodiversity*, 41, 87–107, <https://doi.org/10.1007/s12526-010-0078-4>, 2012.

235

Bluhm, B. A., Janout, M. A., Danielson, S. L., Ellingsen, I., Gavrilo, M., Grebmeier, J. M., Hopcroft, R. R., Iken, K. B., Ingvaldsen, R. B., Jørgensen, L. L., Kosobokova, K. N., Kwok, R., Polyakov, I. V., Renaud, P. E., and Carmack, E. C.: The Pan-Arctic Continental Slope: Sharp Gradients of Physical Processes Affect Pelagic and Benthic Ecosystems, <https://www.frontiersin.org/articles/10.3389/fmars.2020.544386>, 2020.

240

Brey, T., Müller-Wiegmann, C., Zittler, Z. M. C., and Hagen, W.: Body composition in aquatic organisms — A global data bank of relationships between mass, elemental composition and energy content, *J Sea Res*, 64, 334–340, <https://doi.org/https://doi.org/10.1016/j.seares.2010.05.002>, 2010.

245

Buhl-Mortensen, L., Vanreusel, A., Gooday, A. J., Levin, L. A., Priede, I. G., Buhl-Mortensen, P., Gheerardyn, H., King, N. J., and Raes, M.: Biological structures as a source of habitat heterogeneity and biodiversity on the deep ocean margins, *Marine Ecology*, 31, 21–50, <https://doi.org/https://doi.org/10.1111/j.1439-0485.2010.00359.x>, 2010.

Dodge, Y.: Spearman Rank Correlation Coefficient., in: *The Concise Encyclopedia of Statistics*, Springer New York, New York, NY, 502–505, https://doi.org/10.1007/978-0-387-32833-1_379, 2008.

250



Eklöf, J., Austin, Å., Bergström, U., Donadi, S., Eriksson, B., Hansen, J., and Sundblad, G.: Size matters: relationships between body size, dry weight and ash-free dry weight of common coastal, aquatic invertebrates in the Baltic Sea, <https://doi.org/10.7287/PEERJ.PREPRINTS.2368V1>, 2016.

255 Emson, R. H. and Wilkie, I. C.: Fission and autotomy in echinoderms., *Oceanogr Mar Biol Annu Rev*, 155–250, 1980.

FAO: VME indicators, thresholds and encounter responses adopted by R(F)MOs in force during 2019, Available from: <https://www.fao.org/in-action/vulnerable-marine-ecosystems/vme-indicators/en/>, (Accessed 23 December 2025), 2025.

260 Fox, J.: *Applied Regression Analysis, Linear Models, and Related Methods*, SAGE Publications, 1997.

Fox, J.: *Applied Regression Analysis and Generalized Linear Models*, Third Edit., SAGE Publications, 2011.

Fox, J. and Weisberg, S.: *An R Companion to Applied Regression*, Third Edit., SAGE Publications, 2019.

265

García-Arrarás, J. and Greenberg, M.: Visceral Regeneration in Holothurians, *Microsc Res Tech*, 55, 438–451, <https://doi.org/10.1002/jemt.1189>, 2002.

270 Glasser, G. J. and Winter, R. F.: Critical values of the coefficient of rank correlation for testing the hypothesis of independence, *Biometrika*, 48, 444–448, <https://doi.org/10.1093/biomet/48.3-4.444>, 1961.

275 Gogina, M., Zettler, M. L., Vanaverbeke, J., Dannheim, J., Van Hoey, G., Desroy, N., Wrede, A., Reiss, H., Degraer, S., Van Lancker, V., Foveau, A., Braeckman, U., Fiorentino, D., Holstein, J., and Birchenough, S. N. R.: Interregional comparison of benthic ecosystem functioning: Community bioturbation potential in four regions along the NE Atlantic shelf, *Ecol Indic*, 110, 105945, <https://doi.org/10.1016/j.ecolind.2019.105945>, 2020.

Gogina, M., Zettler, A., and Zettler, M. L.: Weight-to-weight conversion factors for benthic macrofauna: recent measurements from the Baltic and the North seas, *Earth Syst Sci Data*, 14, 1–4, <https://doi.org/10.5194/essd-14-1-2022>, 2022.

280



285 Grebmeier, J. M., Bluhm, B. A., Cooper, L. W., Danielson, S. L., Arrigo, K. R., Blanchard, A. L., Clarke, J. T., Day, R. H., Frey, K. E., Gradinger, R. R., Kędra, M., Konar, B., Kuletz, K. J., Lee, S. H., Lovvorn, J. R., Norcross, B. L., and Okkonen, S. R.: Ecosystem characteristics and processes facilitating persistent macrobenthic biomass hotspots and associated benthivory in the Pacific Arctic, *Prog Oceanogr*, 136, 92–114, [https://doi.org/https://doi.org/10.1016/j.pocean.2015.05.006](https://doi.org/10.1016/j.pocean.2015.05.006), 2015.

290

Hewitt, J. E., Anderson, M. J., and Thrush, S. F.: Assessing and monitoring ecological community health in marine systems, *Ecological Applications*, 15, 942–953, <https://doi.org/10.1890/04-0732>, 2005.

Hyman, L. H.: The invertebrates: echinodermata, *The coelomate bilateria*, 4, 1–763, 1955.

295 Javed, H. and Hamid, N.: The Study Methods of the Marine Benthic Community Structures BT - Recent Trends in Marine Toxicological Assessment, edited by: Chuan, O. M., Hamid, N., and Ghazali, A., Springer Nature Switzerland, Cham, 51–89, https://doi.org/10.1007/978-3-031-75713-6_3, 2025.

300 Kornder, N. A., Cappelletto, J., Mueller, B., Zalm, M. J. L., Martinez, S. J., Vermeij, M. J. A., Huisman, J., and de Goeij, J. M.: Implications of 2D versus 3D surveys to measure the abundance and composition of benthic coral reef communities, *Coral Reefs*, 40, 1137–1153, <https://doi.org/10.1007/s00338-021-02118-6>, 2021.

305 Lappalainen, A. and Kangas, P.: Littoral Benthos of the Northern Baltic Sea II. Interrelationships of wet, dry and ash-free dry weights of macrofauna in the Tvärminne Area, *Internationale Revue of Hydrobiology*, 60, 297–312, <https://doi.org/https://doi.org/10.1002/iroh.19750600302>, 1975.

Levene, H.: Robust tests for equality of variances, *Contributions to Probability and Statistics: Essays in Honor of Harold Hotelling*, 2, 278–292, 1960.

310 Marlow, J., Halpin, J. E., and Wilding, T. A.: 3D photogrammetry and deep-learning deliver accurate estimates of epibenthic biomass, *Methods Ecol Evol*, 15, 965–977, <https://doi.org/https://doi.org/10.1111/2041-210X.14313>, 2024.

Petersen, G. and Curtis, M.: Differences in energy flow through major components of subarctic, temperate and tropical marine shelf ecosystems, *Dana*, 1, 1980.

315

Piepenburg, D.: Recent research on Arctic benthos: Common notions need to be revised, *Polar Biol*, 28, 733–755, <https://doi.org/10.1007/s00300-005-0013-5>, 2005.



320 Piepenburg, D., Archambault, P., Ambrose, W., Blanchard, A., Bluhm, B., Carroll, M., Conlan, K., Cusson, M., Feder, H., Grebmeier, J., Jewett, S., Lévesque, M., Petryashev, V., Sejr, M., Sirenko, B., and Włodarska-Kowalczuk, M.: Towards a pan-Arctic inventory of the species diversity of the macro- and megabenthic fauna of the Arctic shelf seas, *Marine Biodiversity*, 41, 51–70, <https://doi.org/10.1007/s12526-010-0059-7>, 2010.

325 Pörtner, H.-O., Scholes, R. J., Arneth, A., Barnes, D. K. A., Burrows, M. T., Diamond, S. E., Duarte, C. M., Kiessling, W., Leadley, P., Managi, S., McElwee, P., Midgley, G., Ngo, H. T., Obura, D., Pascual, U., Sankaran, M., Shin, Y. J., and Val, A. L.: Overcoming the coupled climate and biodiversity crises and their societal impacts, *Science* (1979), 380, eabl4881, <https://doi.org/10.1126/science.abl4881>, 2024.

330 R Core Team: R: A Language and Environment for Statistical Computing, R Foundation for Statistical Computing, Vienna, 2022.

335 Rantanen, M., Karpechko, A. Yu., Lippinen, A., Nordling, K., Hyvärinen, O., Ruosteenoja, K., Vihma, T., and Laaksonen, A.: The Arctic has warmed nearly four times faster than the globe since 1979, *Commun Earth Environ*, 3, 168, <https://doi.org/10.1038/s43247-022-00498-3>, 2022.

Ricciardi, A. and Bourget, E.: Weight-to-weight conversion factors for marine benthic macroinvertebrates, *Mar Ecol Prog Ser*, 163, 245–251, <https://doi.org/10.3354/meps171245>, 1998.

340 Rombouts, I., Beaugrand, G., Artigas, L. F., Dauvin, J.-C., Gevaert, F., Goberville, E., Kopp, D., Lefebvre, S., Luczak, C., Spilmont, N., Travers-Trolet, M., Villanueva, M. C., and Kirby, R. R.: Evaluating marine ecosystem health: Case studies of indicators using direct observations and modelling methods, *Ecol Indic*, 24, 353–365, <https://doi.org/https://doi.org/10.1016/j.ecolind.2012.07.001>, 2013.

345 Rumohr, H., Brey, T., and Ankar, S.: A compilation of biometric conversion factors for benthic invertebrates of the Baltic Sea, Institut für Meereskunde, 1987.

350 Shapiro, S. S. and Wilk, M. B.: An Analysis of Variance Test for Normality (Complete Samples), *Biometrika*, 52, 591–611, <https://doi.org/10.2307/2333709>, 1965.

Sherman, K.: Sustainability, biomass yields, and health of coastal ecosystems: an ecological perspective, *Mar Ecol Prog Ser*, 112, 277–301, <https://doi.org/10.3354/meps112277>, 1994.



355 Souster, T. A., Barnes, D. K. A., Primicerio, R., and Jørgensen, L. L.: Quantifying zoobenthic blue carbon storage across habitats within the Arctic's Barents Sea, *Front Mar Sci*, 10, <https://doi.org/10.3389/fmars.2023.1260884>, 2024.

Spearman, C.: The Proof and Measurement of Association between Two Things, *Am J Psychol*, 15, 72–101, <https://doi.org/10.2307/1412159>, 1904.

360 Stratmann, T., van Oevelen, D., Martínez Arbizu, P., Wei, C.-L., Liao, J.-X., Cusson, M., Scrosati, R. A., Archambault, P., Snelgrove, P. V. R., Ramey-Balci, P. A., Burd, B. J., Kenchington, E., Gilkinson, K., Belley, R., and Soetaert, K.: The BenBioDen database, a global database for meio-, macro- and megabenthic biomass and densities, *Sci Data*, 7, 206, <https://doi.org/10.1038/s41597-020-0551-2>, 2020.

365 Tumbiolo, M. L. and Downing, J. A.: An empirical model for the prediction of secondary production in marine benthic invertebrate populations, *Mar Ecol Prog Ser*, 114, 165–174, 1994.

WoRMS Editorial Board: World register of marine species., <https://www.marinespecies.org>, VLIZ, 2025.

370 Zwierschke, N., Arboe, N. H., Behrisch, J., Blicher, M., and Barnes, D. K. A.: Towards a regional baseline of Greenland's continental shelf seabed biodiversity, *J Environ Manage*, 382, 125285, <https://doi.org/https://doi.org/10.1016/j.jenvman.2025.125285>, 2025.