



# 1 A global consistent database of plankton and detritus from in situ 2 imaging by the Underwater Vision Profiler 5

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30 **Abstract.** Plankton and detritus are essential components of the Earth's oceans influencing biogeochemical cycles  
 31 and carbon sequestration. Climate change impacts their composition and marine ecosystems as a whole. To improve  
 32 our understanding of these changes, standardized observation methods and integrated global datasets are needed to  
 33 enhance the accuracy of ecological and climate models. Here, we present a global dataset for plankton and detritus  
 34 obtained by two versions of the Underwater Vision Profiler 5 (UVP5). This release contains the images classified in  
 35 33 homogenized categories, as well as the metadata associated with them, reaching 3,114 profiles and ca. 8 million  
 36 objects acquired between 2008-2018 at global scale. The geographical distribution of the dataset is unbalanced, with  
 37 the Equatorial region (30° S - 30° N) being the most represented, followed by the high latitudes in the northern  
 38 hemisphere and lastly the high latitudes in the Southern Hemisphere. Detritus is the most abundant category in terms  
 39 of concentration (90%) and biovolume (95%), although its classification in different morphotypes is still not well  
 40 established. Copepoda was the most abundant taxa within the plankton, with *Trichodesmium* colonies being the second  
 41 most abundant. The two versions of UVP5 (SD and HD) have different imagers, resulting in a different effective size  
 42 range to analyse plankton and detritus from the images (HD objects >600 µm, SD objects >1 mm) and morphological



properties (grey levels, etc.) presenting similar patterns, although the ranges may differ. Therefore, recommendations are provided for the appropriate use of this data when conducting studies. A large number of images of plankton and detritus will be collected in the future by the UVP5, and the public availability of this dataset will help it being utilized as a training set for machine learning and being improved by the scientific community. This will reduce uncertainty by classifying previously unclassified objects and expand the classification categories, ultimately enhancing biodiversity quantification. The dataset that constitutes this first release is available at SEANOE.

**Key words:** plankton, images, Underwater Vision Profiler, Global Ocean

## 1 Introduction

Plankton and particulate matter play a crucial role in natural biogeochemical cycles and provide essential ecosystem services, including carbon sequestration, nutrient cycling, and primary food source for various marine organisms (Turner, 2015; Boyd et al., 2019; Stemmann and Boss, 2012; Steinberg and Laundry, 2017). These components of the marine environment are significantly influenced by climate change, which alters ocean temperature, acidity, oxygenation, and circulation patterns (Doney et al., 2012; Constable et al., 2014). Such changes may, in turn, impact plankton composition by affecting their growth and survival (Hays et al., 2005; Dam and Baumann, 2017; Yebra et al., 2022), which will impact the production and composition of their solid waste, hereafter referred as detritus, with potential consequences for elemental biogeochemical cycles and deep-sea ecosystems (Du Pontavice et al., 2020). Zooplankton biomass and diversity, as well as bulk particulate matter, were identified as essential ocean, biodiversity, and climate variables by the Global Ocean Observing System (Chiba et al., 2018; Batten et al., 2019).

The size range of plankton and detritus spans from pico- to macro-sizes, influencing the methods required for their observation. Different technologies, including microscopy, optical imaging, sediment traps, and remote sensing, are employed depending on the target size class (Karsenti et al., 2011; Davis et al., 2005; Stemmann et al., 2008; Möller et al., 2012; Siegel et al., 2024). Many planktonic organisms and detritus are fragile and their abundances decrease with their size, making their collection and analysis challenging (Stemmann et al., 2008; Soviadan et al., 2024; Atherden et al., 2024). Consequently, obtaining homogeneous global datasets remains difficult notably if different instruments that measure various properties are used (Moriarty and O'brien, 2013). Furthermore, plankton exhibits a patchy distribution in the ocean due to physical, chemical, and biological processes (Suthers et al., 2019). These constraints demand increased sampling efforts and the development of standardized methodologies to enhance comparability of global plankton and detritus observations. For plankton, quantitative imaging has been identified as the best means to do so (Lombard et al 2019). Addressing these challenges is critical for improving our understanding of oceanic carbon cycling and ecosystem responses to environmental change.

According to specific objectives, biogeochemists focus on biomass as a measure of carbon content, while ecologists are primarily interested in community composition and biodiversity. Conventional sampling methods such as sediment traps, nets, pumps, and bottles further contribute to these differences, as they may not provide data that is directly comparable across disciplines. Modelers, on the other hand, require integrative datasets that bridge these gaps,



77 enabling accurate simulations of biogeochemical cycles, ecosystem dynamics, and carbon fluxes (Ratnarajah et al.,  
 78 2023). They are particularly interested in harmonized data formats, conversion factors between abundance and  
 79 biomass, and standardized methodologies to improve the predictive power of ecological and climate models.  
 80 Addressing these disparities is essential for advancing interdisciplinary research and improving our understanding of  
 81 oceanic processes.

82 Predicting the future state of plankton biodiversity and/or biomass with habitat or biogeochemical models is  
 83 difficult and requires data obtained synoptically and regularly. Models could be partially fed by Continuous Plankton  
 84 Recorder surveys (Batten et al., 2019) and satellite programs, such as the recently launched PACE (Gorman et al.,  
 85 2019), but in both cases data on plankton is limited to the surface layer. Deep collection by net tows followed by the  
 86 identification with microscopes provides the classical taxonomic identification, but this sampling methodology  
 87 destroys fragile organisms and requires dedicated ship time (Calbet, 2024; Soviadan et al., 2024; Giering et al., 2022).  
 88 In addition, this classical classification is time-consuming and susceptible to human bias, which can hinder effective  
 89 quality control (Goswami, 2004). Sensors mounted on conventional in situ platforms may provide better spatial  
 90 resolution and have less impact on the observed target. In situ acoustics with broadband systems allow for broad  
 91 taxonomic identification but are not adapted for detritus due to their poor scattering. In situ imaging methods are better  
 92 suited to recognize plankton and detritus, but detect them in a smaller volume. Considering the advantages and  
 93 limitations of each method, we can conclude that in situ imaging is particularly suited for observing plankton and  
 94 detritus, using their shape to infer taxonomy or morphological attributes (Picheral et al., 2010; Stemmann et al., 2012;  
 95 Lombard et al., 2019). However, there is a compromise between the observed volume and the resolution of the camera  
 96 that defines a size range and taxonomic resolution for each type of sensor. Recent publications have suggested  
 97 (Lombard et al., 2019, Kiko et al., 2023) and showcased (Drago et al. 2022, Clements et al. 2023, Laget et al. 2024)  
 98 that cooperative observation of plankton with imaging systems can allow an upscaling of regional observations by  
 99 independent observers to global scale. Thus, plans are now underway to measure these variables with imaging systems  
 100 on large-scale observing programs, like the Bio-GO-SHIP (Clayton et al., 2022) or the BGC-Argo (Claustre et al.,  
 101 2020; Picheral et al., 2022) programs.

102 The Underwater Vision Profiler camera system (UVP, version 5) is an optical imaging instrument that  
 103 captures digital images of a defined volume of seawater, provisioned with LED lights. It was designed to automatically  
 104 detect, size, and count biotic and abiotic particles in the marine environment as it profiles through the water column  
 105 (Picheral et al., 2010). To date, the 25 units of the UVP5 have been used at 14,462 sites since 2008 providing 94.8  
 106 millions of images and ~1,000 new profiles every year. The UVP5 optimal size range to identify and measure  
 107 morphological traits is in the order of 600  $\mu\text{m}$  to a few millimeters (Picheral et al., 2010; Drago et al., 2022). The  
 108 number of taxa that can be recognized is over 200, whereas robust data is available for the 30 most abundant taxa  
 109 (Drago et al., 2022; Panaïotis et al., 2023). Semi-supervised classification (automatic prediction followed by human  
 110 validation) of plankton images (Irisson et al., 2022) was performed at regional scales, because of the time required to  
 111 analyse all profiles and the lack of dedicated work flow to deal with millions of images. Recently the development of  
 112 the collaborative platform Ecopart (<https://ecopart.obs-vlfr.fr/>) for UVP metadata and data curation, and EcoTaxa



(<https://ecotaxa.obs-vlfr.fr/>) for supervised image classification facilitated data treatment for the users. The data, initially recorded to address regional questions, can then be aggregated in global datasets to investigate global distributions of counts, size and vertical flux of all detected objects (Kiko et al., 2022; Clements et al., 2022; Clements et al., 2023). Furthermore, these platforms allow global studies on macrozooplankton biomass (Drago et al., 2022), and community composition (Panaïotis et al., 2023), as well as on the community proportions of Rhizaria (Biard et al., 2016), their impact on carbon flux attenuation, and silicification (Laget et al., 2024). To date, these studies provided the following data products: global plankton biomass, counts and group specific carbon demand data. In addition, to facilitate data access for modelers, all UVP5 particle size profiles, which were analysed without image recognition, were released at a 1 degree spatial resolution (Kiko et al., 2022). A more recent database obtained by combining images from Imaging Flow CytoBot, Zooscan and UVP5 to provide data products on plankton and detritus biomass binned in 1 degree resolution was also released (Dugenne et al., 2024). An attempt was made to classify detritus images based on their morphology (Trudnowska et al., 2022) but given their high number and the lack of consistent shapes, their classification at global scale is still challenging.

Here, we present the global dataset comprising approximately 8 millions validated images of plankton and detritus, rigorously verified by expert taxonomists, associated with relevant metadata and their morphological measurements, that was not released in earlier studies. The dataset is of interest for marine ecologists interested in plankton biogeography and biogeochemical modeling and for computer scientists developing Artificial Intelligence methods to classify images. While the current plankton classification has been homogenized for 33 categories, there remains potential for further improvement to increase the list of taxa consistent at a global scale. Classification of detritus was not performed, as existing methodologies developed at regional scale must be improved for a global approach. By providing the raw images for plankton and detritus, we hope to foster the development of new algorithms to sort and analyze them. The manuscript is organized as follows: details about the UVP5, the inter-calibration and quality control procedures, as well as the dataset structure are provided in the Material and Methods section. Maps of dataset distribution, summarizing statistics regarding taxa composition, size spectrum, description of global detritus and plankton distribution in the Results section. Finally, recommendations for dataset use and potential future expansions of it are provided in the Discussion section.

## 2 Material and methods

### 2.1 Sampling sites

Data from the Underwater Vision Profiler 5 (UVP5), an in-situ imaging system designed to detect, measure, and quantify the distribution of zooplanktonic organisms and marine particles (Picheral et al., 2010), were used. The UVP5 dataset (Figure 2) was compiled from observations across all oceans over a 10-year period (2008–2018) through a collaboration of international partners. It includes 3,114 profiles from 62 EcoTaxa projects conducted during research cruises by different institutions (Table A1), each aimed at addressing specific local or basin-scale research questions.



## 147 2.2 Image acquisition with UVP5

### 148 2.2.1 UVP5 description

149 The UVP5 was commercialized in 2010 and produced until 2021. The standard definition (SD) version with  
 150 a 1.3 Megapixel greyscale camera was produced between 2008 and 2016 (serial numbers 000 to 011) and the high-  
 151 definition (HD) version with a 4 Megapixel greyscale camera was produced between 2016 and 2021 (serial numbers  
 152 200 to 223). In the standard definition setting, the UVP5 images a volume of about 1 L at a frequency of 5 to 20 Hz  
 153 and can be deployed down to 6000 m depth. The UVP5 is mostly integrated in the CTD-Rosette and therefore its  
 154 results (particle and plankton counts) can be related to environmental data obtained with other sensors. All particles  
 155 larger than appr. 100  $\mu\text{m}$  Equivalent Spherical Diameter (ESD) are sized and counted but their images are not stored  
 156 because the low number of pixels precludes any recognition of the particles. Recognized objects larger than 30 pixels  
 157 (UVP5SD) or 80 pixels (UVP5HD) are automatically cropped and the resulting vignettes stored and further analyzed.

### 158 2.2.2 Image analysis by zooprocess

159 Vignettes contain plankton organisms, detritus and artefacts larger than approximately 1 mm ESD for  
 160 UVP5SD and 600  $\mu\text{m}$  for UVP5HD. Pixel size-to-millimeter conversions for UVP5SD and UVP5HD are derived  
 161 from objects of 30 and 80 pixels, respectively. For simplicity, we converted the surface area in pixels to its Equivalent  
 162 Spherical Diameter. In addition, the released dataset contains 42 morphological features characterizing each object.  
 163 Metadata collection (geographic location, date, etc.) and processing of all 8.46 million images was carried out using  
 164 the ZooProcess software. The processing included the segmentation of the object from the obtained raw vignette and  
 165 the measurement of 42 morphological features associated with each object (area, major and minor axis, grey level  
 166 etc.; Picheral and Mérieux, 2025; Table A2). For user convenience, the images were inverted to show dark objects  
 167 on a white background and a scale bar was added to each vignette. Images and metadata were imported into EcoTaxa  
 168 (Picheral et al., 2017), an application which allows a taxonomic classification of images via supervised learning  
 169 algorithms, followed by manual validation (Irisson et al., 2022).

170 The two versions of the UVP5 have been inter-calibrated based on the size spectrum of all particles measured  
 171 concurrently in natural conditions (Kiko et al., 2022). Therefore, size is a conservative property between the  
 172 instruments and plankton community composition can be inter-compared. While all parameters, except for shape-  
 173 based ones, differ between the SD and HD versions due to the distinct dynamics of their imagers, other morphological  
 174 properties, such as opacity, brightness, etc., may not be conservative, requiring caution in their joint analysis. For the  
 175 HD version, measurements are taken from images with a dark field, which are then converted to a light field  
 176 comparable to those in the SD version. Additionally, only images from a continuous descent of the instrument were  
 177 retained.



### 2.2.3 Semi-automatic image identification

Using EcoTaxa (<https://ecotaxa.obs-vlfr.fr/>), we applied semi-automated recognition in two steps following the third strategy path defined in Figure 2 by Irisson et al. (2022). First, features obtained using Zooprocess software or features re-calculated by a convolutional neural network (CNN) are considered to automatically predict object identities using a Random Forest algorithm. Therefore, various image learning sets were used independently on a project-by-project basis by different users. In this way, plankton organisms can be successfully classified into a few broad categories from the overdominant category of detritus with reasonable success. In a second step, manual validation and further sorting ensured correct classification of all plankton categories and detritus. In general the sorting was performed by various users and institutions. To reduce the risk of wrong identification, a shared UVP5 taxonomic guide was used by all annotators to homogenize image sorting (EcoTaxaGuide application). In addition, annotations were rigorously quality checked by the authors. The classification of all images was strictly based on the morphological features of the objects in most cases. However, few annotators have used contextual information provided by other means (e.g., bottle or net collection, depth, GPS) to classify objects with inadequate morphological attributes into taxa.

## 2.3 Dataset

The UVP5 image dataset comprises vertical profiles validated to more than 99%, with only near-completely validated profiles retained for subsequent analysis. Currently, the different annotators of the 62 EcoTaxa projects chosen have sorted the image datasets in more than 250 categories from species level to phyla or by adding morphological attributes and life stages. Such detailed classification is not possible over the whole dataset with high accuracy because recognition and sorting of organisms can be a source of bias depending on the levels of perception and experience of the people who perform them. Several cognitive biases exist, such as boredom, fatigue or a classification biased towards the most used groups (Culverhouse, 2007; Culverhouse et al., 2014). Currently 71 different annotators have classified the images, potentially introducing substantial classification variability and errors (Table A3).

In the present dataset, to reduce these errors and ensure classification homogeneity, the images were thereafter grouped into only 33 broader taxonomic groups for living organisms by combining all vignettes within the children categories (Table 1), while in EcoTaxa the original categories are maintained. The broad taxonomy list was established following recent published works using UVP5 data (Drago et al., 2022; Panaïotis et al., 2023; Laget et al., 2024) to ensure a minimum of 50 images per category and ecological patterns at regional or global scales for specific groups or community composition. For example, rhizarians were well studied at local (Biard and Ohman, 2020) and global scales (Biard et al., 2016; Laget et al., 2024) while polychetes were mostly studied in the tropical Atlantic (Christiansen, 2016). Community composition analyses have been conducted at local scales by Forest et al. (2012) and Barth et al. (2020), and at a global scale by Panaïotis et al. (2022). Global biomass was estimated in Drago et al. (2022). Inter-comparison with net data at global scale (Soviadan et al., 2024) allowed to set confidence in the abundance assessments of many of these groups.



Images of unidentifiable objects exhibiting probable biological characteristics (e.g., symmetry, appendages, or tentacles) were classified into the “plankton-like” category. Thus, the term -like in this dataset refers to all objects that have a structured morphology (appendages, symmetry) that make them resemble an organism but that can’t be classified as such. This labeling includes issues due to bad focus, low image resolution, incomplete organism capture, or the unknown nature of the organisms to the annotator that hinder the accurate taxonomic identification. For example, copepoda-like contains all vignettes of copepods for which the antenna are not visible while the body shape resembles typical prosome and urosome. If many identical objects were common but not evenly distributed and not yet classified, new temporary categories were created and labeled temporary (‘tmpXXX’). When contextual information to identify them was used, “contextual” was added to their names. All images of non-living organisms, including poor-quality images (blurred, low grey level, bubbles, artificial nature, badly segmented images) or particles (aggregates, fibers, pellets) were sorted into several categories without consistency criteria across the consortium. Low-quality images (mostly objects not completely in focus) were set in a category named “artefact” that includes bad focus particles, plankton and real artefact, while particles were grouped into a single detritus category.

To ensure data consistency across projects, a subset of the dataset was reviewed to verify the accuracy of taxonomic sorting within each group. A total of 200 images were extracted from each category, or all available images if fewer than 200 were present. For detritus, 10 % of the total images were selected and independently reviewed after the final dataset was assembled. The error or uncertainty rate was under 8 % for taxa classification. Among the detritus, less than 0.1 % were plankton organisms. The resulting global dataset consisted of 7.05 million detritus images and 734 thousands plankton images from 3,114 profiles.

Due to the large number of plots required to display the results for all categories, results are shown for a subset of the original categories. From the 33 initial categories derived from taxonomy (Table 1), five taxonomic groups were formed for further analysis based on feeding strategies and/or their ecology. These groups can be highly useful as input for biogeochemical or community ecology models. The groups include Crustacea (Copepoda and Malacostraca), gelatinous filter-feeders (Appendicularia, Salpida), gelatinous carnivores (Chaetognatha, Narcomedusae, Siphonophorae, Ctenophora), Rhizaria (Collodaria, colonial Collodaria, other Rhizaria, Foraminifera, Phaeodaria, Acantharea), and *Trichodesmium*. From these groups, biovolumes were calculated along with concentrations at different depth layers and for the analysis of the average grey level.

### 2.3.1 Data output format

We provide four tables with numerical data (in .tsv format), as well as all images (with .jpg extension) while their full description is available in the information documents (UVP5\_dataset\_organization.txt) during the download. Briefly, ‘objects.tsv’ is the data table which contains each object information. The three first columns refer to the unique object ID given by Ecotaxa, original object classification in Ecotaxa and object classification according to Table 1. The following 42 columns contain all the object features obtained from the image (see Table A2). The ‘samples.tsv’ file is the table with the metadata corresponding to all the coordinates (latitude, longitude, date and time) and acquisition details (pixel size and UVP model). The ‘samples\_volume.tsv’ is a table that contains the volume of





water in which the object was extracted and that is used to calculate concentrations (middle depth bin and volume imaged). Finally, ‘properties\_per\_bin.tsv’ is a table with concentrations, biovolume, average size (ESD) of the 33 consistent categories per depth bin and per sample. The native size bin of the dataset is 5 m, but to reduce the size of the file and avoid too many zeros, we provide 25 m depth bins in the surface ocean (up to 200 m), 50 m depth bins (up to 500 m), 100 m bins (up to 1000 m) to 250 m below 1000 m. We provide the code in R to recalculate the table with a depth bin of 5 m. The two files ‘ODV\_biovolumes.txt’ and ‘ODV\_concentrations.txt’ contains the same information as ‘properties\_per\_bin.tsv’ but in the ODV format.

**Table 1: List of categories that are quantitatively consistent in this work and names of children categories not homogeneous among the projects. Categories also defined by their morphotypes or life stages are in bold. The number of images in each category is given in parenthesis. The published dataset contains both the broad and the children categories. When available publications using the dataset at regional scales are listed. Exponent numbers denote the association with an image example in Figure 1.**

| Broad categories selected in this work (number of vignettes) | If more than 50 vignettes, children categories (number of vignettes) proposed by data owners but not yet homogenized  | Reference   |
|--|---|---|
| Acantharia (12,616) <sup>1</sup>                             |   |   |
| Actinopterygii (482) <sup>2</sup>                            |   |   |
| Annelida (3,261) <sup>3</sup>                                | Alciopidae (74), Phyllostodidae (53), Tomopteridae (237), Acrociridae (218), Flota (185), Poeobius (358)  | Christiansen et al., 2022; Barth and Stone, 2022                  |
| Appendicularia (9,354) <sup>4</sup>                          | <i>body</i> (540), <i>house</i> (836)   | Forest et al., 2012   |
| Bacillariophyta-contextual (44,637) <sup>5</sup>             | All objects classified as such using contextual data  |   |
| Chaetognatha (6,793) <sup>6</sup>                            | <b>tail</b> <Chaetognatha (64), <b>head</b> <Chaetognatha (62)  | Forest et al., 2012; Barth and Stone, 2022                        |
| other Collodaria (7,644) <sup>7</sup>                        | <i>Solitaryblack</i> (5,905), <i>solitaryglobule</i> (1,738)  | Biard and Ohman, 2020   |
| colonial Collodaria (2,329) <sup>8</sup>                     |   | Biard and Ohman, 2020   |
| Copepoda (173,909) <sup>9</sup>                              | like<Copepoda (72,284), Eucalanidae (782), like<Temoridae (280), <i>ovigerous</i> (117), <i>ovigerous_side</i> (3,484), <i>ovigerous_top-bottom</i> (287), Pseudocalanus (99), Harpacticoida (65) | Forest et al., 2012; Vilgrain et al., 2021; Barth and Stone, 2022 |
| Ctenophora (2,743) <sup>10</sup>                             | Beroida (238), Cestidae (66), Lobata (111), <b>tentacle</b> <Ctenophora (65)  | Forest et al., 2012   |
| Doliolida (1,304) <sup>11</sup>                              |   |   |
| Eumalacostraca (11,160) <sup>12</sup>                        | Amphipoda (434), Pleuroncodes (258), Euphausiace (531), Munnopsis (70)  | Forest et al., 2012; Barth and Stone, 2022                        |
| Foraminifera (3,014) <sup>13</sup>                           |   |   |
| Gasteropoda (1,500) <sup>14</sup>                            | Gymnosomata (87), Limacinidae (1015), Creseidae (120), other Thecosomata (198)  |   |





|   |   |   |
|---|---|---|
| Narcomedusae (2,938) <sup>15</sup>                      | <i>Solmundella bitentaculata</i> (1,791), Solmaris (203)  |   |
| Nostocales (1,369) <sup>16</sup>                        | Aphanizomenon (689), Dolichospermum (179), Nodularia (501)  |   |
| Ostracoda (8,427) <sup>17</sup>                         |   | Barth and Stone, 2022                   |
| Phaeodaria (65,030) <sup>18</sup>                       | Castanellidae (2,405), Circoporidae (271), Tuscaroridae (85), Aulacanthidae (236), Aulacantha (18,572), Aulographis (827), Coelodendrum (283), Coelographis (2,474), <b>leg</b> <Coelodendridae (1,513), Medusettidae (459), Aulosphaeridae (31,616), Aulatractus (217), <b>colonial</b> <Aulosphaeridae (906), like<colonial (1,381), Cannosphaeridae (1,416), <b>colonial</b> <Cannosphaeridae (84) | Biard and Ohman, 2020                   |
| Pyrosoma (334) <sup>19</sup>                            |   |   |
| Salpida (592) <sup>20</sup>                             |   |   |
| Siphonophorae (2,557) <sup>21</sup>                     | <b>part</b> <Siphonophorae (380), <b>tentacle</b> <Siphonophorae (1,381)  |   |
| Trachymedusae (462) <sup>22</sup>                       | Halimnionidae (184), <i>Aglaia digitale</i> (100)   |   |
| <i>Trichodesmium</i> (88,676) <sup>23</sup>             | Puff (41,936), Tuff (46,740)  | Guidi et al., 2012; Sandel et al., 2015 |
| <i>Trichodesmium</i> -contextual (98,956) <sup>24</sup> | all objects were classified as such by using contextual data, 95% are from the same cruise  | Dupouy et al., 2018                     |
| other Cnidaria (1,994) <sup>25</sup>                    | Ceriantharia (74)   |   |
| other Crustacea (4,569) <sup>26</sup>                   | <b>larvae</b> <Crustacea (58), <b>tail</b> <Crustacea (122)   |   |
| other Hydrozoa (8,659) <sup>27</sup>                    | Leptothecata (96)   |   |
| other Mollusca (3,833) <sup>28</sup>                    | veliger (3,092)   |   |
| other Rhizaria (9,911) <sup>29</sup>                    | contains plankton resembling to Rhizaria  |   |
| tentacle of Cnidaria (8,008) <sup>30</sup>              |   |   |
| plankton-like (74,241) <sup>31</sup>                    | contains all structured objects most probably living organisms, which are in multiple categories to check (35,083), possibly plankton (18,289), to resort (15,535), other living (5,868) and to rename (2,942)  |   |
| detritus (7,055,138) <sup>32</sup>                      | contains particles such as aggregates, filaments, mucus net which are not yet sorted  | Trudnowska et al., 2021                 |
| Artefact (49,636) <sup>33</sup>                         | bubble (8,332), turbid (9,507)  |   |

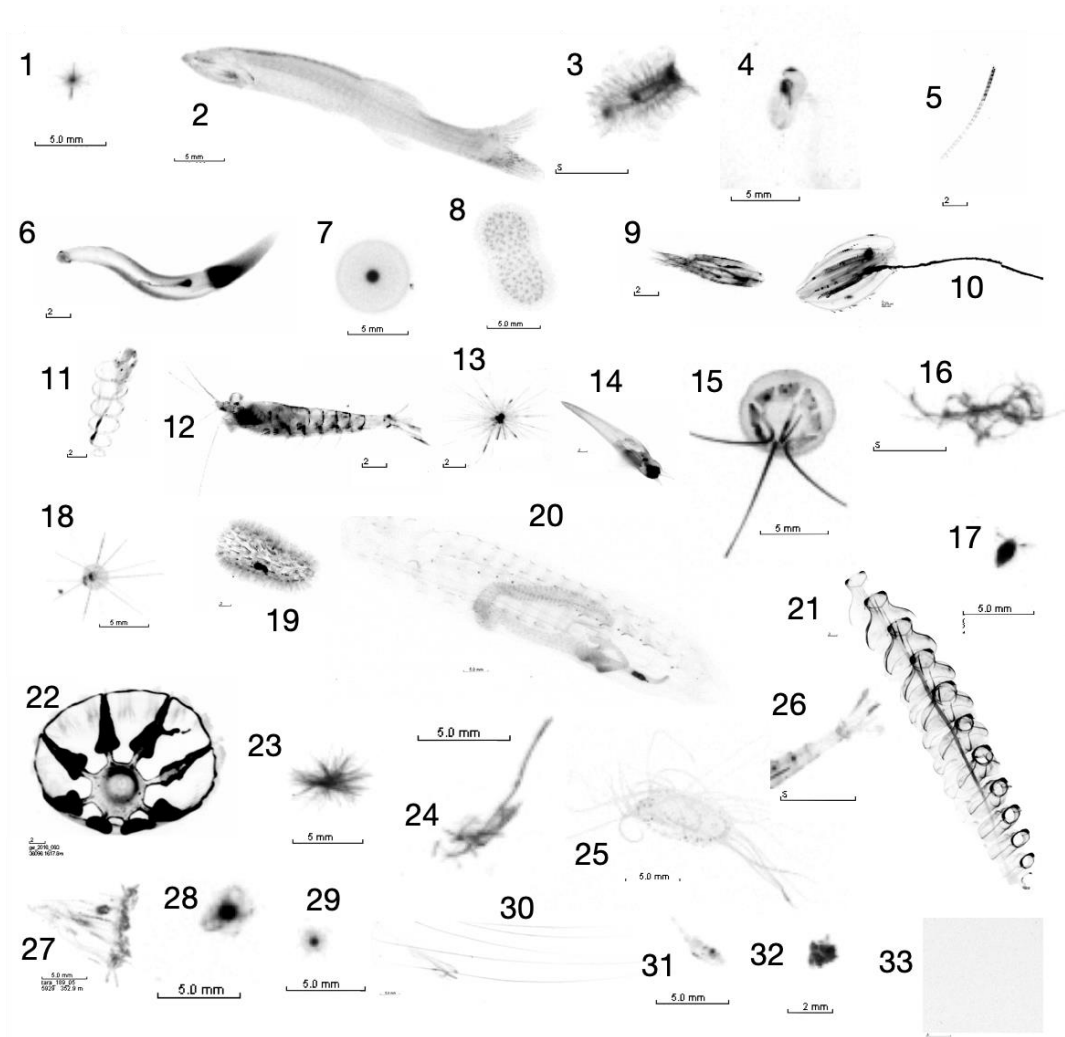


Figure 1: Examples of images for each category (Table 1) present in the dataset.

### 3 Results

#### 3.1 Data coverage in time and space

The dataset presented here reveals a heterogeneous global distribution. The map in Figure 2 illustrates the profiles collected using two different versions of the UVP5 (SD and HD) during deployments across the global ocean between 2008 and 2018. While sampling efforts cover the global ocean, UVP5 deployments predominantly occurred in the northern hemisphere (66.3% of profiles), especially in tropical and temperate latitudes where seasonal variations are well observed. In contrast, the southern hemisphere has fewer observations (33.7% of profiles), resulting in a less



comprehensive dataset. The equatorial region (30° S-30° N) represents the highest proportion of UVP5 profiles (47.25%). The vertical profiles distribution reveals a predominant concentration of measurements in the upper 1000 m of the water column, representing 40.6% and 46.8% for the SD and HD models respectively, with a peak occurring within the upper 500 m. This indicates that most deployments targeted shallower depths. The SD version of the UVP5 demonstrates the most extensive coverage, with profiles spanning the entire latitudinal range. Meanwhile, the HD version UVP5 profiles are more prevalent in the Northern Hemisphere. Importantly, the two versions were rarely co-located in space and time, preventing meaningful inter-comparison of zooplankton community composition. These patterns provide valuable insights into the sampling strategies and data representation of UVP5 profiles across the global ocean. They also highlight the seasonal variability observed primarily in the northern hemisphere and the under-sampling of the southern hemisphere and the deep sea.

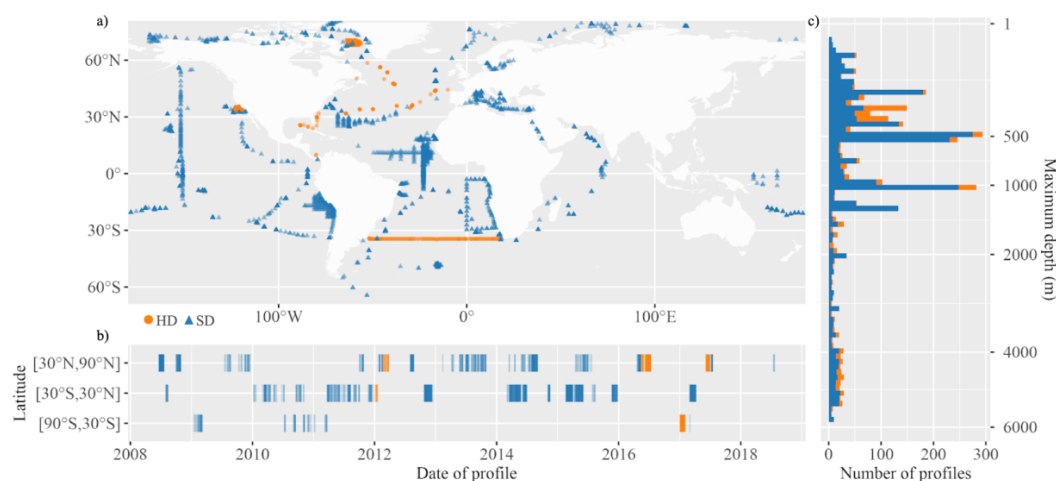
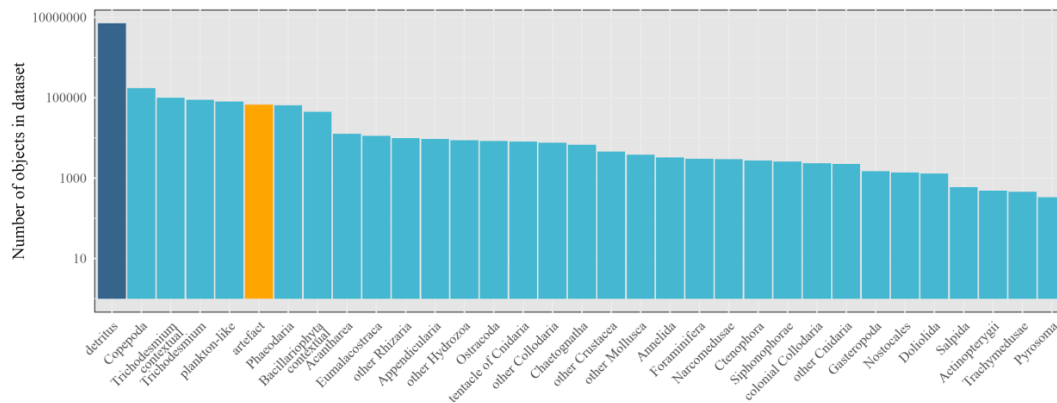


Figure 2: a) Global sampling effort for the two versions of UVP5 (SD and HD, blue and orange, respectively) between 2008-2018 in different oceans. b) Indicates the latitudinal distribution of UVP5 profiles and c) shows the maximum vertical extent of the profiles.

### 3.2 Detritus and plankton composition in the whole dataset

Figure 3 presents the relative abundance of dataset categories, with the six most prevalent in decreasing order being: detritus, Copepoda, *Trichodesmium*-contextual, *Trichodesmium*, plankton-like, and Phaeodaria. Detritus is the most abundant category by far (90.5% of total images), indicating a substantial presence of non-living particulate organic material. Copepoda dominates the zooplankton and represents a significant fraction of the dataset (2.2%), followed by *Trichodesmium* (1.3%) and the *Trichodesmium*-contextual (1.1%) categories. The category plankton-like reflecting some level of taxonomic uncertainty in the dataset, is the fourth category in terms of images within the dataset (1%). The artefact category, including bubbles and artefact, occupied the sixth position (0.9%). This category is followed by Phaeodaria (0.8%) and Bacillariophyta-contextual (0.6%). Each of the remaining groups represent less than 0.16% of the total number of objects.

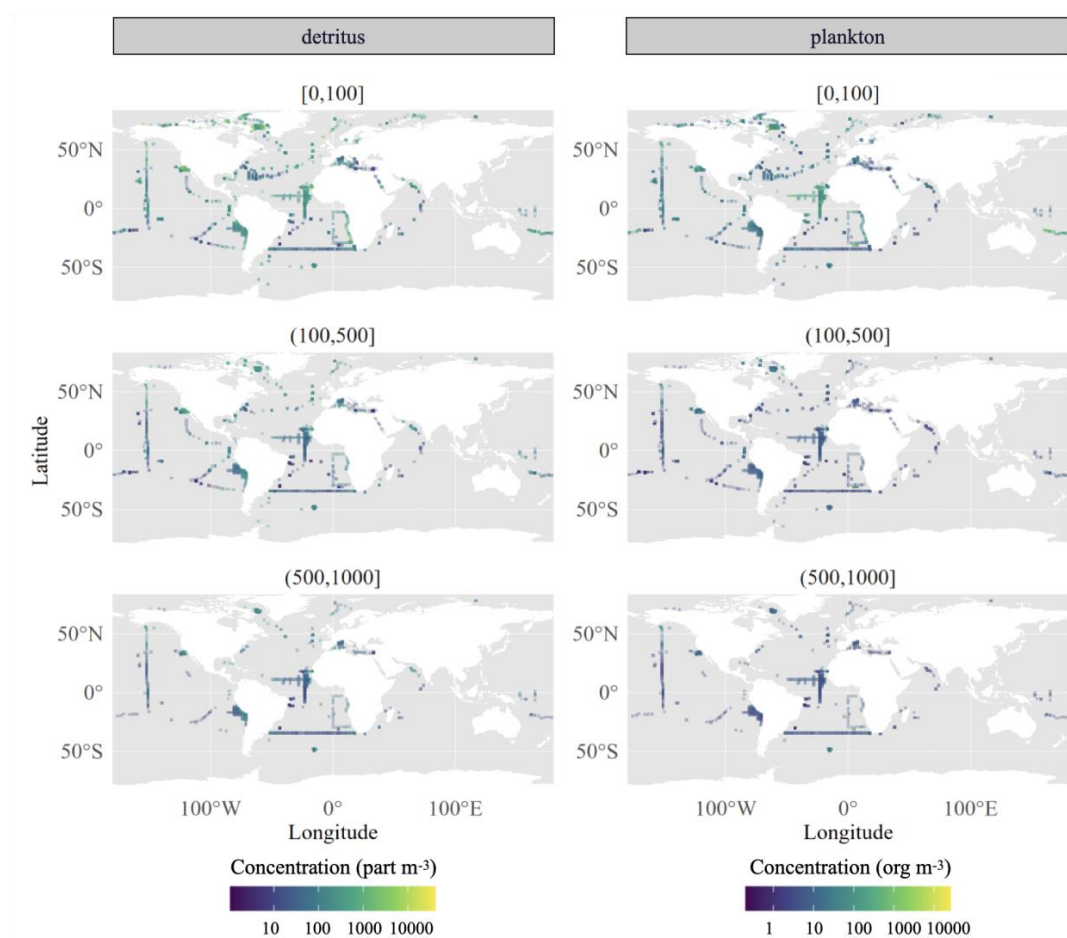


293

294 **Figure 3: Barplot with total number of objects in logarithmic scale available in the dataset per category (detritus and**  
295 **plankton) for the 33 consistent categories. The categories are ordered by descending count, with specific categories**  
296 **("detritus" and "artefact") highlighted in distinct colors. Note that the artefact category represented here contains all**  
297 **vignettes including turbid, bubbles, badfocus (see section 2.3.1).**

### 298 3.3 Global distribution of plankton and detritus

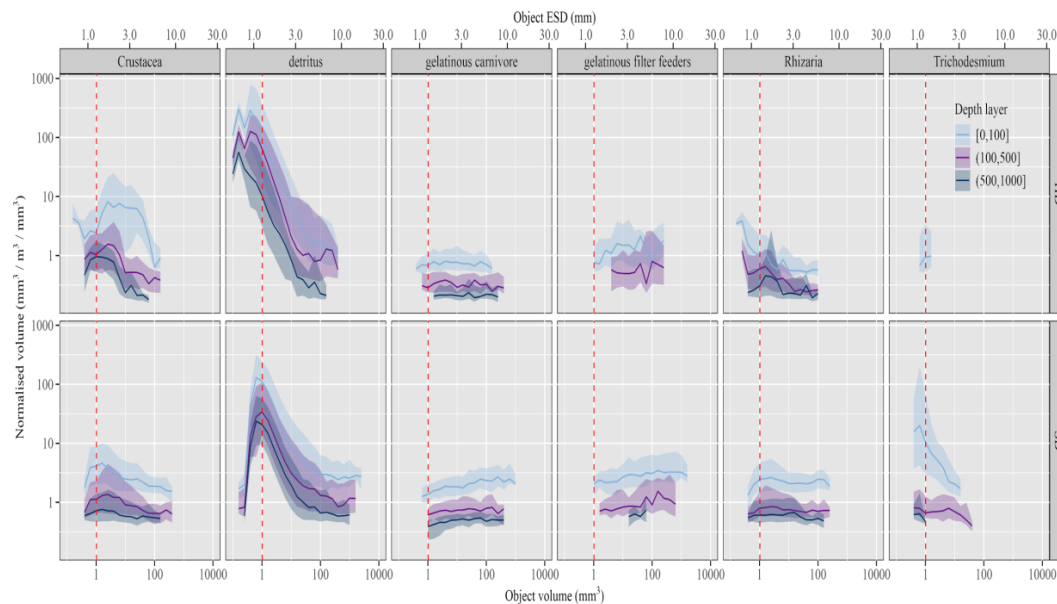
299 The spatial patterns of plankton and detritus concentrations in the global ocean vary significantly with depth  
300 and location. Figure 4 depicts the global distribution of abundances in three depth layers. Across all depth layers,  
301 plankton and detritus abundances are increased (up to 5 plankton/m<sup>3</sup> and 60 detritus/m<sup>3</sup> in the 0-100 m depth layer  
302 and up 2 plankton/m<sup>3</sup> and 20 detritus/m<sup>3</sup> in the 500-1000 m depth layer) in eastern boundary current systems  
303 (especially in the Californian and Senegal upwelling systems) or in coastal seas. Minimum values (up to 2 plankton/m<sup>3</sup>  
304 and 20 detritus/m<sup>3</sup> in the 0-100 m depth layer and up 1 plankton/m<sup>3</sup> and 1 detritus/m<sup>3</sup> in the 500-1000 m depth layer)  
305 are usually found in the center of large ocean gyres (notably in the South Pacific and the Indian Ocean).



**Figure 4: Map of the global detritus and plankton concentration (log particles/organism m<sup>-3</sup>) in three layers (0-100, 100-500 and 500-1000 m).**

### 3.4 Average size distribution of SD and HD versions

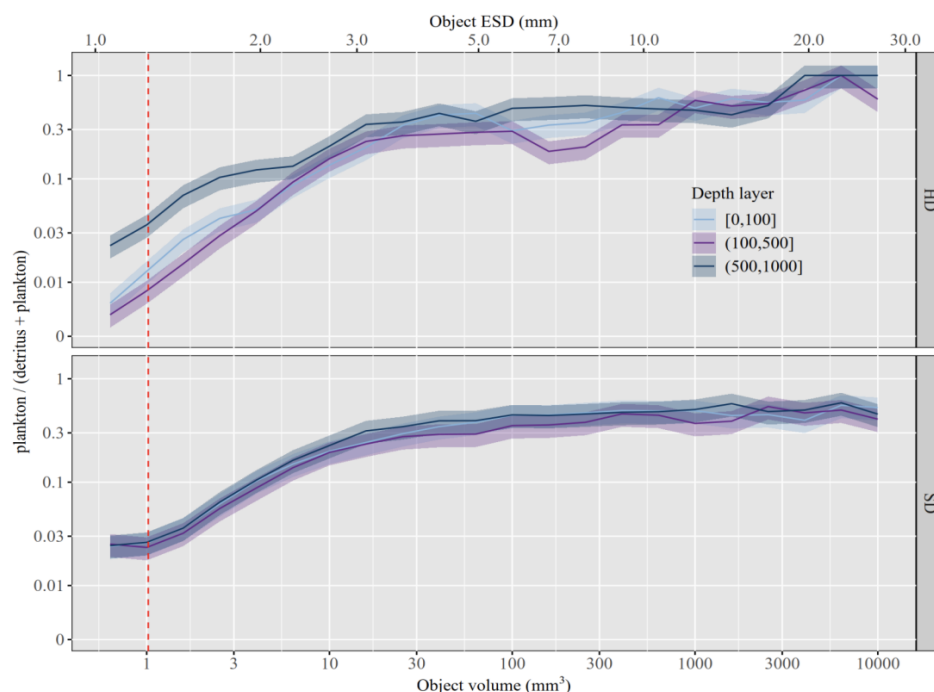
The distribution of detritus, the most abundant category, follows a dome-like pattern (criteria from Drago et al., 2022), with UVP5 showing a lower detection threshold for particles below 1 mm for UVP5 SD and 600  $\mu$ m for UVP5 HD. In contrast, taxonomic categories exhibit a much flatter normalized biomass/biovolume size spectra (NBSS) distribution compared to detritus. Detritus dominates the smaller size range while the contribution of plankton increases relative to detritus in the larger size range. Except for Rhizaria, smaller planktonic organisms were more abundant in the upper 100 m in the UVP5hd dataset. This analysis emphasizes the differing detection limits of the two camera versions in the small size range for both plankton and detritus (Figure 5). Consequently, we recommend a standardized size cutoff of 1.02 mm for ecological studies focusing on abundance and biomass integrating both data sets, while studies utilizing solely UVP5 HD should employ a 600  $\mu$ m size threshold.



**Figure 5: Average size distribution of detritus and plankton categories obtained by the SD and HD version of the UVP5 in three depth layers. Red dashed line indicates the threshold value (1.02 mm) below which UVP5 detection of organisms is difficult to quantify properly. Crustacea (Copepoda and Malacostraca), gelatinous filter-feeders (Appendicularia, Salpida), gelatinous carnivores (Chaetognatha, Narcomedusae, Siphonophorae, Ctenophora), Rhizaria (Collodaria, colonial Collodaria, other Rhizaria, Foraminifera, Phaeodaria, Acantharea), and *Trichodesmium*.**

### 3.5 Plankton to detritus ratio as a function of size

For both UVPs, the proportion of plankton images on all images (detritus and plankton) increases with size from 0.01 at 500 $\mu$ m to about 0.8 for objects larger than 3 mm in ESD. The two instruments display a similar pattern with a plateau in this ratio starting at about 3 mm. In general, the ratio does not vary with depth and remains at 0.8 for objects larger than 3mm.

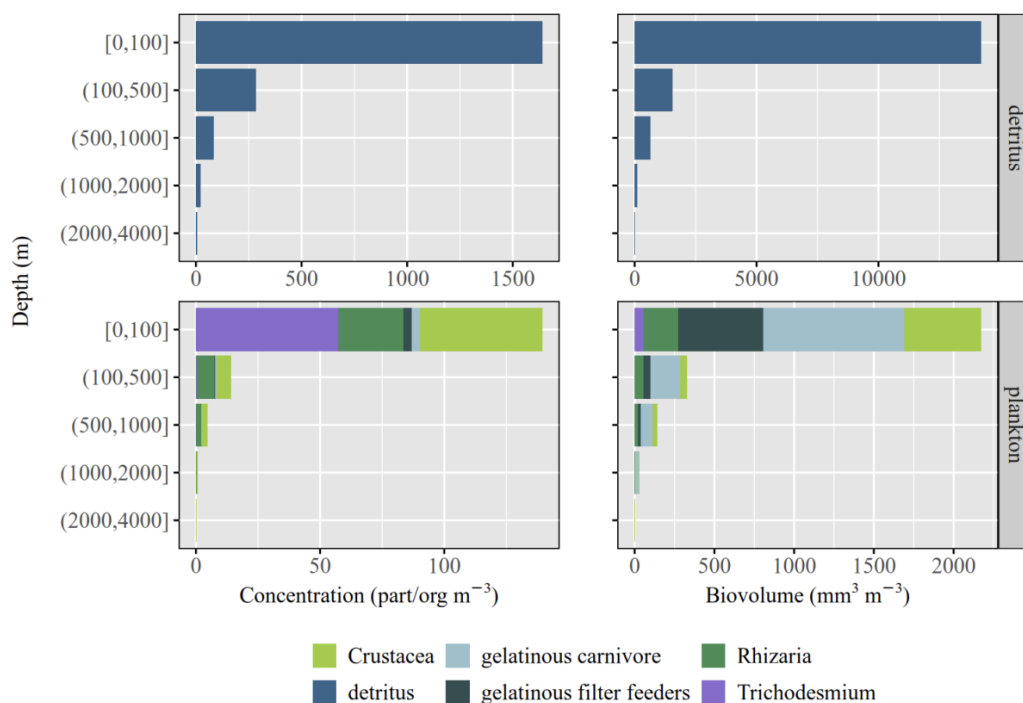


**Figure 6: Abundance ratio of plankton (without plankton-like) over all images as a function of size for the two different UVP5 versions. Red dashed line indicates the threshold value (1.02 mm) below which a robust detection of organisms is limited.**

### 3.6 Vertical profiles of detritus and plankton concentrations

When objects are analyzed across different vertical layers (Figure 7), detritus exhibits increased concentrations, primarily within the uppermost 100 m of the water column. The biovolume distribution mirrors the concentration pattern, with detritus exhibiting substantially higher values—several orders of magnitude greater—compared to living plankton organisms. Trichodesmium (not including Trichodesmium-contextual) was the most abundant group in terms of concentration within the upper 100 meters, with its presence decreasing with depth. However, in terms of biovolume, due to their smaller size (Figure 4), it represented a relatively small proportion compared to other groups analyzed. Crustacea, on the other hand, were present in similar proportions in both concentration and biovolume, showing a high abundance in the surface layer and a decline with depth. In the case of gelatinous plankton, both with a filter-feeding (Appendicularia, Salpida) and carnivores strategies (Chaetognatha, Narcomedusae, Siphonophorae, Ctenophora), their concentrations were very low in all analyzed depths, while their biovolume became significantly important, particularly in the upper 100 m of the water column. Both parameters showed a decrease with depth for these groups, similar to the patterns observed for the aforementioned groups. Rhizaria, in turn, showed peak concentrations in the upper 100 m, which gradually decreased to 1000 m. Their biovolume values exhibited patterns similar to those observed for concentration.

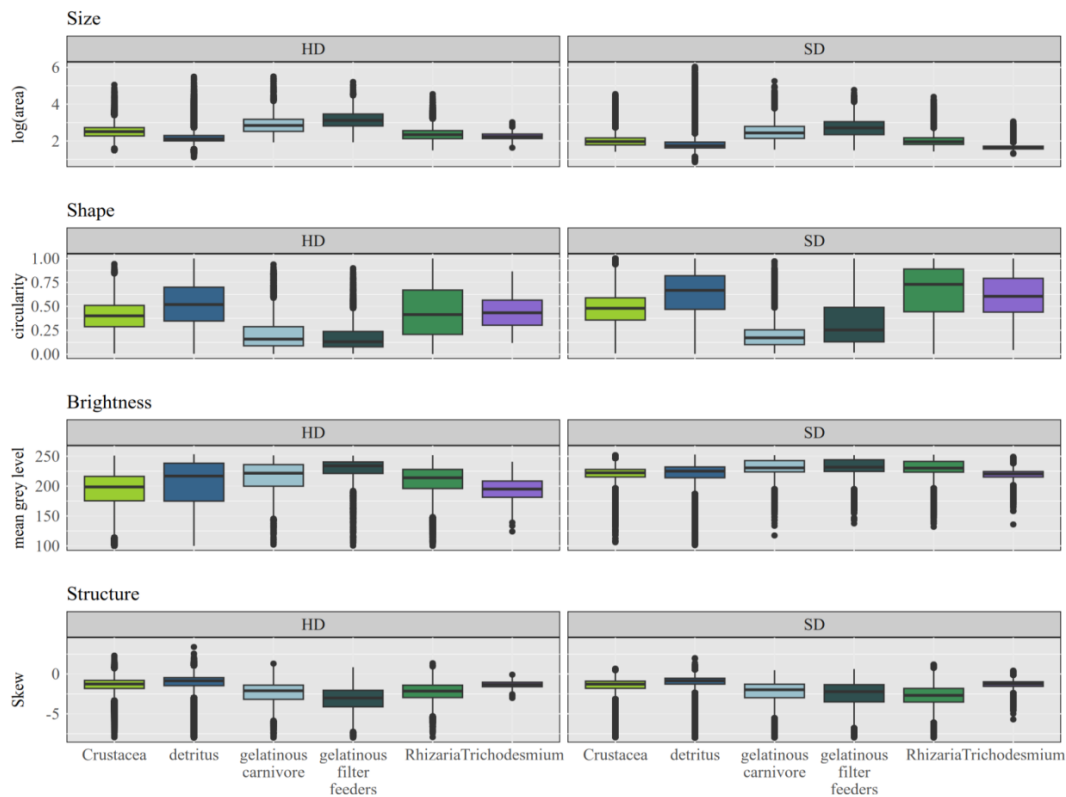




**Figure 7: Global vertical profiles of concentration and biovolume for detritus and the five plankton functional groups.**part: particles of detritus; org: organisms from different taxa categories. Crustacea (Copepoda and Malacostraca), gelatinous filter-feeders (Appendicularia, Salpida), gelatinous carnivores (Chaetognatha, Narcomedusae, Siphonophorae, Ctenophora), Rhizaria (Collodaria, colonial Collodaria, other Rhizaria, Foraminifera, Phaeodaria, Acantharea), and *Trichodesmium*.

### 3.7 Morphological properties

The key morphological properties for detritus and the five taxonomic groups presented similar patterns between the two UVP5 versions (SD and HD). In some cases, the range differed between versions for the groups depending on the property examined (Figure 8). Gelatinous groups were the biggest categories in terms of area measured, while they presented the lowest circularity, compared to the remaining groups. Crustacea was the darkest group, whilst gelatinous were the lightest ones.



**Figure 8: Box plots of key exemplary morphological properties (area, circularity, mean grey level, skew) for detritus and the five taxonomic groups (Crustacea, gelatinous carnivore, gelatinous filter feeders, Rhizaria and *Trichodesmium*) for the two UVP5 versions (HD and SD). Crustacea (Copepoda and Malacostraca), gelatinous filter-feeders (Appendicularia, Salpida), gelatinous carnivores (Chaetognatha, Narcomedusae, Siphonophorae, Ctenophora), Rhizaria (Collodaria, colonial Collodaria, other Rhizaria, Foraminifera, Phaeodaria, Acantharea), and *Trichodesmium*.**



## 367 4 Discussion

368 The availability and accessibility of both the images and their associated metadata present new challenges,  
 369 encouraging the scientific community to leverage and refine them for future research. With this objective in mind,  
 370 the dataset presented here serves as a starting point for further exploration of the current database (10,000 profiles  
 371 not fully validated) and future data incorporated in EcoTaxa (approximately 500 profiles annually on average over  
 372 the last three years for UVP5 versions only), covering both plankton and detritus. In this context, we discuss the  
 373 strengths and limitations of the dataset, as well as its potential applications and future directions for the UVP  
 374 database.

### 375 4.1 Consideration related to spatial coverage and instruments performances

376 The dataset covers all major ocean basins, though the southern hemisphere and the deep sea remains largely  
 377 undersampled. The list of profiles included are primarily confined to the mid- to high-latitudes of the northern  
 378 hemisphere (Figure 2). Some specific regions are well sampled as they belong to monitoring or time-series programs  
 379 developed during the last decades, such as the California Current, the Western Mediterranean, and the Equatorial  
 380 region, through initiatives like the CCELTER program (e.g., Biard and Ohman, 2020), ongoing observational efforts  
 381 of MOOSE (e.g., Llopis Monferrer et al., 2022), and long-term studies in the Equatorial region (e.g., Kiko et al.  
 382 2017, Fernández-Carrera et al., 2023). The even distribution across ocean basins is mostly the result of the Tara  
 383 Ocean Expeditions (2009-2013). Altogether, this dataset represents global coverage over a decade, with  
 384 homogeneous and intercomparable methodologies across UVP5 versions. Despite this extensive coverage, there are  
 385 some potential issues regarding data quality. For instance, the small volume sampled by UVP5s can lead to  
 386 challenges in detecting rare groups, and the quality of the images may affect the accuracy of object identification.  
 387 Additionally, "zeros" might not always reflect the actual absence of objects or natural patchiness of plankton and  
 388 particles, but rather a result of low volume sampled or detection limits of the instrument.

389 At larger size ranges, as organisms become rare, statistical analysis may be challenging and required to  
 390 aggregate counts across thick ocean layers. At the surface, abundant taxa may permit 10 m binning intervals (Dupouy  
 391 et al., 2018; Biard and Ohman., 2020), though 20 m bins are typically more appropriate (Picheral et al., 2010). For  
 392 rare taxonomic groups, aggregation across several hundred meters may be necessary to achieve statistically  
 393 meaningful analyses (Hauss et al., 2016; Panaïotis et al., 2023). In this study, we employed a depth-layered approach  
 394 to mitigate variability in rare taxonomic groups, aggregating data into five progressively wider depth layers (0-100,  
 395 100-500, 500-1000, 1000-2000 and 2000-4000 m). While previous publications have chosen 0-200 m and 200-500  
 396 m (Drago et al., 2022) or a dynamic one, for the first layer, set as the deepest value among the mixed layer depth and  
 397 the euphotic depth (median 88 m, Panaïotis et al., 2023). Such a choice depends on the scientific question.

398 The two versions of the UVP5 are inter-calibrated based on the size spectrum of all objects obtained from  
 399 repeated joint profiles at the same location (Picheral et al., 2010; Kiko et al., 2022). A peak in the size spectrum at



the lower size range generally reflects the minimum size of efficient detection by in situ imaging, while high variability in the large size range due to the poor ability to detect rare large objects results in constant (e.g., flatter) distribution (Stemmann and Boss, 2012). In between these two size ends, a steep decrease is generally observed which translates to a quasi-linear decrease on a log scale. The detritus size spectra (Figure 5) clearly show that the HD version, presenting a smaller pixel size, has a better detection than the SD version for detritus smaller than 1 mm, but that above a threshold size of 1 mm both versions are efficient up to a few mm as the slopes get flatter.

#### 4.2 Considerations related to classification

Variations in taxonomic classification depth among annotators necessitated the merging of certain groups, resulting in a final list of 33 taxa with a lower common denominator (Table 1). To validate classification accuracy, a random subset of images (200 from each taxon and 10,000 for detritus) was independently reviewed by a single operator. Error rates were consistently below 10% across all groups, with most groups exhibiting error rates under 2.5%. An additional source of error in the case of detritus is that many annotators had moved un-focused particles into the category artefact. This category includes various sources and types of images that may result from factors such as a poorly tied rope, which was recorded during the image capture, as well as seals or cables. On the other hand, it may also consist of effects like bubbles caused by the descent through the water column and the air intrusion, as well as artifacts resulting from image blurriness where no actual object or non-recognizable object is present (real artifact). When analyzing the artefact category which contains some subjectivity, considering for example that bad-focused could be very transparent objects, we found that it consists of 85% of real detritus shaped objects, 5% of plankton and 10% of artefact. Sorting them in the EcoTaxa database system would be too time-consuming, so users must decide whether to include or exclude them during the analysis. Here, we merged them with detritus as they would become a minority (less than few percents). However, it is important to note that objects classified as artefacts (not shown in Figure 7) represent less than 1% of the total dataset when the concentration is calculated, but contribute to more than 200% when volume is considered. Their impact is high mostly on large objects because many artefacts are observed in very turbid water where particles are badly segmented. This highlights the significance of their removal.

##### *Plankton-like*

We have gathered in plankton-like all objects that are most probably plankton. Inside this category, ~60% of the objects in the possibly plankton group may be Rhizaria, but the image definition impairs their recognition. It is important to keep this category apart from the detritus category because they represent a significant fraction of the total plankton count and probably living biomass. Although a significant portion indicates uncertain classification, it is also in this category that most new findings can be performed.

##### *Plankton contextual*

The term -contextual in this dataset refers to objects that were classified as a taxonomic category but do not have the morphology of the taxa. Scientists on oceanographic surveys who had other contextual information



have created the category *Trichodesmium*-contextual and Bacillariophyta-contextual for objects that aren't formally identified as *Trichodesmium* puff and tuff or as diatom chains. In this context, the classification is supported by many other optical observations (for example see Dupouy et al., 2018 for *Trichodesmium*). Due to the risk of misinterpreting ecological results or impairing the quality of the learning set, it is better to distinguish them from objects whose images are typical of plankton organisms. In the case of the two categories, *Trichodesmium* and *Trichodesmium*-contextual, they contain nearly the same number of objects, so including both would double the amount of data for this group. On the other hand, Bacillariophyta-contextual represents the sole category for this genus within the dataset, where its inclusion or not could lead to its complete exclusion.

#### 4.3 Scientific relevance

Overall, this dataset provides a unique and comprehensive tool for the scientific community, enabling researchers to examine the intricate relationships between plankton, detritus, and ecosystem function. The detailed classification of plankton and detritus provides a valuable resource for understanding ecological dynamics at global (Drago et al., 2022; Panaïotis et al., 2023; Laget et al., 2024) and local scales (see references in Table 1) within marine systems as already shown in several studies. By analyzing the abundance, diversity, and distribution of planktonic organisms, researchers can gain insights into food web structures, energy flow, and the interactions between different trophic levels (e.g., D'Alelio et al., 2016; Perhirin et al., 2025). The data allows ecologists the monitoring of plankton communities, which are essential indicators of environmental health and can reflect changes in water quality, nutrient levels, and ecosystem productivity (Muller-Karger et al., 2018). The dataset is equally valuable in biogeochemical studies, so researchers can better predict how changes in plankton community composition and detritus abundance influence nutrient cycling and the overall functioning of marine ecosystems. The complex relationships between these components provide critical insights into primary productivity, nutrient fluxes, and the potential impacts of environmental stressors such as climate change and ocean acidification. Additionally, biogeochemical models that integrate this dataset can help refine predictions of carbon sequestration in the ocean, an essential process for regulating atmospheric CO<sub>2</sub> levels and mitigating global warming. As plankton and detritus play pivotal roles in the ocean's carbon pump, understanding their dynamics is crucial for assessing the resilience of marine ecosystems and their ability to mitigate or exacerbate climate change effects (e.g., Le Quéré et al., 2016; Rohr et al., 2023). Through these integrated models, the dataset offers a robust framework for exploring the complex feedback mechanisms between marine biological communities and biogeochemical processes, advancing our ability to predict future changes in marine ecosystems under varying environmental scenarios.

#### 4.4 Moving forward: future development

The published dataset (3,114 profiles, 7,000,000 images) is publicly available, while only validated images are visible from the remaining profiles in EcoTaxa (6,000 profiles, 83,000,000 images), although not yet publicly accessible to be downloaded and/or re-processed. Therefore, future users should contact the data owners through the Ecopart and EcoTaxa platforms to request access to the complete datasets.



468 Despite the efforts of the data owners, many images in the EcoTaxa database for UVP5 were not recognized  
 469 in any taxonomic group, so were categorized under a plankton-like category. The challenge of further sorting  
 470 plankton-like organisms is significant due to its great abundance, but it also provides an opportunity to increase the  
 471 biodiversity within the dataset and refine the taxonomy.

472 Future classification should focus its efforts on sorting the detritus category, as a recent study highlights  
 473 that sinking speed estimated for various particle categories varies with particle morphology (Trudnowska et al., 2022;  
 474 Soviadan et al., 2025) or that mesoscale activities mix them vertically according to their morphology (Accardo  
 475 et al., 2025). In addition, they represent a very high proportion in abundance and probably of organic biomass  
 476 compared to the plankton. Additionally, the specific classification of detritus particles, such as fecal pellets and their  
 477 associated zooplankton communities, will enhance our understanding of their dynamics, improve the accuracy of  
 478 carbon flux calculations, and refine their representation in biogeochemical models (Perhirin et al., 2025). Therefore,  
 479 classifying particles based on their traits could provide valuable new insights into their origin and role in the carbon  
 480 cycle through the gravitational pump (Boyd et al., 2019). When it comes to zooplankton, recent studies revealed that  
 481 analyzing their traits, such as body size, morphology, and metabolic rates, which significantly influence their  
 482 ecological roles and interactions within marine ecosystems, provides a framework for linking organismal  
 483 characteristics to ecosystem functions, offering deeper insights into zooplankton behavior, community structure, and  
 484 their contributions to biogeochemical cycles (Titocci et al., 2025).

485 Transfer learning has shown promising potential in leveraging existing datasets to enhance training for  
 486 improved recognition tasks. By utilizing the present dataset as a model training for machine learning and artificial  
 487 intelligence (AI) techniques can be employed to better classify and extract relevant features (Lumini and Nanni,  
 488 2019; Irisson et al., 2022). Specifically, AI could play a key role in sorting and categorizing the 7,000,000 detritus  
 489 images in the dataset, enhancing the processing of the remaining 5,598 UVP5 profiles and their corresponding 70  
 490 million images. Furthermore, integrating new profiles, such as the anticipated 500 annually for UVP5 and 5,000  
 491 annually for the new version of the UVP (UVP6lp and hd; Picheral et al., 2022), into the observation system will  
 492 enhance the robustness of the database, ensuring its adaptability and scalability for future data handling and feature  
 493 extraction. This observation effort is supported by GOOS (Dexter and Summerhayes, 2010) and already in operation  
 494 in large-scale observing programs, like the Bio-GO-SHIP (Clayton et al., 2022) or possibly by the BGC-Argo  
 495 (Claustre et al., 2020) program. Having better embedded classifiers is particularly important for BGC Argo for which  
 496 the floats and the cameras are not recovered. Given the versatility of the new UVP6, which can be attached or  
 497 associated with different platforms, it has allowed for widespread distribution and, consequently, a significant  
 498 deployment of profiles (~23 thousands on EcoTaxa), reaching 87 millions of images in the last 4 years since its  
 499 commercialization in 2020. These advancements will ultimately contribute to more efficient data analysis and  
 500 support the continuous improvement of environmental monitoring systems.

## 501 **Data availability**

502 The dataset is available on SEANOE at <https://doi.org/10.17882/107583> (Nocera et al., 2025).



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522 **Author contribution method**

523 ACN, JOI and LS formulated the goals for dataset presentation, quality control and the publication of a global UVP5  
524 dataset and their respective images. LS and MP led the community effort supported by all UVP managers and quality  
525 control endeavours, supported by all co-authors. ACN and LS conceived and drafted the article. All authors  
526 participated in writing the article.

527 **Conflict of interest statement**

528 The authors declare no conflicts of interest.





529 **Appendix A**

530 **Table A1. List of all projects included in the present study (N = 62), with their corresponding project identification (pprojid),**  
531 **title (ptitle), data owner and license registered on EcoTaxa (<https://ecotaxa.obs-vlfr.fr/>).**



| pprojid | ptitle                            | data_owner                     | projid | title   | license      |
|---------|-----------------------------------|--------------------------------|--------|---|--------------|
| 1       | uvp5_sn003_jerico_2017            | lars.stemmann@obs-vlfr.fr      | 578    | UVP5 JERICO 2017  |              |
| 2       | uvp5_sn000_boum2008               | lars.stemmann@obs-vlfr.fr      | 122    | UVP5 BOUM 2008  |              |
| 3       | uvp5_sn000_ccelter_2012           | tristan.biard@univ-littoral.fr | 45     | UVP5 CCELTAR (Iter2008 ccelter_2012 ccelter_2014)                   | CC BY-NC 4.0 |
| 4       | uvp5_sn000_lohafex2009            | lars.stemmann@obs-vlfr.fr      | 109    | UVP5 LOHAFEX 2009   | CC0 1.0      |
| 5       | uvp5_sn000_iter2008               | tristan.biard@univ-littoral.fr | 45     | UVP5 CCELTAR (Iter2008 ccelter_2012 ccelter_2014)                   | CC BY-NC 4.0 |
| 6       | uvp5_sn000_tara2009               | lars.stemmann@obs-vlfr.fr      | 579    | UVP5 Tara Oceans (tara2009, tara2010, tara2011, tara2012, tara2013) | CC BY 4.0    |
| 7       | uvp5_sn000_tara2010               | lars.stemmann@obs-vlfr.fr      | 579    | UVP5 Tara Oceans (tara2009, tara2010, tara2011, tara2012, tara2013) | CC BY 4.0    |
| 9       | uvp5_sn000_tara2012               | lars.stemmann@obs-vlfr.fr      | 579    | UVP5 Tara Oceans (tara2009, tara2010, tara2011, tara2012, tara2013) | CC BY 4.0    |
| 10      | uvp5_sn002_moose_dyf_2013         | lars.stemmann@obs-vlfr.fr      | 35     | UVP5 Dyfamed 2013 2014 2015 2016 2017 2018                          |              |
| 11      | uvp5_sn002_moose_dyf_2014         | lars.stemmann@obs-vlfr.fr      | 35     | UVP5 Dyfamed 2013 2014 2015 2016 2017 2018                          |              |
| 12      | uvp5_sn002_moose_dyf_2015         | lars.stemmann@obs-vlfr.fr      | 35     | UVP5 Dyfamed 2013 2014 2015 2016 2017 2018                          |              |
| 13      | uvp5_sn002_moose_dyf_2016         | lguidi@obs-vlfr.fr             | 35     | UVP5 Dyfamed 2013 2014 2015 2016 2017 2018                          |              |
| 14      | uvp5_sn002_moose_dyf_2017         | coppola@obs-vlfr.fr            | 35     | UVP5 Dyfamed 2013 2014 2015 2016 2017 2018                          |              |
| 15      | uvp5_sn002_moose_ge_2013          | lars.stemmann@obs-vlfr.fr      | 30     | UVP5 MooseGE 2012 2013 2014 2015 2016                               | CC BY-NC 4.0 |
| 16      | uvp5_sn000_malina2009             | lars.stemmann@obs-vlfr.fr      | 28     | UVP5 MALINA 2009  | CC BY 4.0    |
| 17      | uvp5_sn002_moose_ge_2014          | lars.stemmann@obs-vlfr.fr      | 30     | UVP5 MooseGE 2012 2013 2014 2015 2016                               | CC BY-NC 4.0 |
| 20      | uvp5_sn000_operex2008             | lars.stemmann@obs-vlfr.fr      | 110    | UVP5 OPEREX 2008  | CC BY 4.0    |
| 21      | uvp5_sn001_2012_moose_ge          | stemmann@obs-vlfr.fr           | 30     | UVP5 MooseGE 2012 2013 2014 2015 2016                               | CC BY-NC 4.0 |
| 22      | uvp5_sn001_2012_msm22             | rkiko@geomar.de                | 593    | UVP5 Geomar 2012 msm22  |              |
| 23      | uvp5_sn001_2012_msm23             | rkiko@geomar.de                | 584    | UVP5 Geomar 2012 msm23  |              |
| 31      | uvp5_sn002_moose_ge_2015_filtered | lars.stemmann@obs-vlfr.fr      | 30     | UVP5 MooseGE 2012 2013 2014 2015 2016                               | CC BY-NC 4.0 |
| 32      | uvp5_sn002_somba_ge_2014          | lars.stemmann@obs-vlfr.fr      | 36     | UVP5 SOMBA 2014   | CC BY 4.0    |



|    |                                |                                     |     |   |              |
|----|--------------------------------|-------------------------------------|-----|---|--------------|
| 41 | uvp5_sn003_cassiopee_2015      | lars.stemmann@obs-vlfr.fr           | 38  | UVP5 CASSIOPEE 2015   | CC BY 4.0    |
| 42 | uvp5_sn003_ccelter_2014        | tristan.biard@univ-littoral.fr      | 45  | UVP5 CCELTER (Iter2008 ccelter_2012 ccelter_2014)                   | CC BY-NC 4.0 |
| 43 | uvp5_sn003_ccelter_2016        | tristan.biard@univ-littoral.fr      | 156 | UVP5 CCELTER 2016   | CC BY-NC 4.0 |
| 44 | uvp5_sn003_dewex_spring_2013   | lars.stemmann@obs-vlfr.fr           | 4   | UVP5 DEWEX 2013 (spring)  | CC BY 4.0    |
| 46 | uvp5_sn003_outpace_2015        | lguidi@obs-vlfr.fr                  | 37  | UVP5 OUTPACE 2015   | CC BY 4.0    |
| 47 | uvp5_sn003_sargasso_a          | lombard@obs-vlfr.fr                 | 22  | UVP5 Sargasso 2014 (sargasso_a sargasso_b)                          | CC BY 4.0    |
| 48 | uvp5_sn003_sargasso_b          | lombard@obs-vlfr.fr                 | 22  | UVP5 Sargasso 2014 (sargasso_a sargasso_b)                          | CC BY 4.0    |
| 49 | uvp5_sn003_tara2013            | lars.stemmann@obs-vlfr.fr           | 579 | UVP5 Tara Oceans (tara2009, tara2010, tara2011, tara2012, tara2013) | CC BY 4.0    |
| 51 | uvp5_sn003zp_tara2012          | lars.stemmann@obs-vlfr.fr           | 579 | UVP5 Tara Oceans (tara2009, tara2010, tara2011, tara2012, tara2013) | CC BY 4.0    |
| 53 | uvp5_sn005_dewex_2013_winter   | lars.stemmann@obs-vlfr.fr           | 3   | UVP5 DEWEX 2013 (winter)  | CC BY 4.0    |
| 54 | uvp5_sn005_dy032_2015_filtered | francois.carlotti@mio.osupytheas.fr | 40  | UVP5 DY032 205  | CC BY 4.0    |
| 55 | uvp5_sn005_moose_ge_2013       | lars.stemmann@obs-vlfr.fr           | 30  | UVP5 MooseGE 2012 2013 2014 2015 2016                               | CC BY-NC 4.0 |
| 56 | uvp5_sn008_an1304              | marcel.babin@takuvik.ulaval.ca      | 41  | UVP5 AN Arctique (an1304 an1405 an1407) part A to validate          |              |
| 57 | uvp5_sn008_an1405              | marcel.babin@takuvik.ulaval.ca      | 41  | UVP5 AN Arctique (an1304 an1405 an1407) part A to validate          |              |
| 58 | uvp5_sn008_an1406              | marcel.babin@takuvik.ulaval.ca      | 618 | UVP5 AN 1406  | CC BY-NC 4.0 |
| 59 | uvp5_sn008_an1407              | marcel.babin@takuvik.ulaval.ca      | 41  | UVP5 AN Arctique (an1304 an1405 an1407) part A to validate          |              |
| 60 | uvp5_sn008_green_2015_i_cecamp | marcel.babin@takuvik.ulaval.ca      | 42  | UVP5 GREEN EDGE Ice Camp 2015                                       | CC BY-NC 4.0 |
| 61 | uvp5_sn008_green_2016_i_cecamp | marcel.babin@takuvik.ulaval.ca      | 315 | UVP5 GREEN EDGE Ice Camp 2016                                       | CC BY-NC 4.0 |
| 62 | uvp5_sn008_subice_2014         | stemmann@obs-vlfr.fr                | 107 | UVP5 Subice 2014  |              |
| 64 | uvp5_sn009_2015_p16n           | amcdonnell@alaska.edu               | 43  | uvp5_sn009_2015_p16n  | CC BY 4.0    |
| 65 | uvp5_sn009_2015_p16n_goa       | amcdonnell@alaska.edu               | 43  | uvp5_sn009_2015_p16n  | CC BY 4.0    |
| 68 | uvp5_sn010_2014_m105           | rkiko@geomar.de                     | 602 | UVP5 Geomar 2014 m105   |              |
| 69 | uvp5_sn010_2014_m106           | rkiko@geomar.de                     | 595 | UVP5 Geomar 2014 m106   |              |



|     |                                    |   |      |   |              |
|-----|------------------------------------|---|------|---|--------------|
| 70  | uwp5_sn010_2014_m107               | rkiko@geomar.de                         | 559  | UVP5 Geomar 2014 m107   |              |
| 71  | uwp5_sn010_2014_m108               | rkiko@geomar.de                         | 586  | UVP5 Geomar 2014 m108   |              |
| 72  | uwp5_sn010_2014_ps88b              | rkiko@geomar.de                         | 558  | UVP5 Geomar 2014 ps88b  |              |
| 73  | uwp5_sn010_2015_m116               | rkiko@geomar.de                         | 589  | UVP5 Geomar 2015 m116   |              |
| 76  | uwp5_sn010_2015_m121               | rkiko@geomar.de                         | 560  | UVP5 Geomar 2015 m121   |              |
| 81  | uwp5_sn201_2016_naames_02          | lee.karp-boss@maine.edu                 | 146  | UVP5hd NAAMES02   | CC BY-NC 4.0 |
| 83  | uwp5_sn201_ccelter_2017            | tristan.biard@univ-littoral.fr          | 627  | UVP5hd CCELTTER 2017  | CC BY-NC 4.0 |
| 86  | uwp5_sn203_greenedge_2016          | lars.stemmann@obs-vlfr.fr               | 149  | UVP5hd GreenEdge 2016   | CC BY-NC 4.0 |
| 87  | uwp5_sn203_greenedge_2016_1b       | lars.stemmann@obs-vlfr.fr               | 149  | UVP5hd GreenEdge 2016   | CC BY-NC 4.0 |
| 92  | uwp5_sn201_2017_naames_03          | lee.karp-boss@maine.edu                 | 637  | UVP5hd NAAMES03   | CC BY-NC 4.0 |
| 95  | uwp5_sn010_2017_m135               | rkiko@geomar.de                         | 548  | UVP5 Geomar 2017 m135   |              |
| 99  | uwp5_sn000_tara2011                | lars.stemmann@obs-vlfr.fr               | 579  | UVP5 Tara Oceans (tara2009, tara2010, tara2011, tara2012, tara2013) | CC BY 4.0    |
| 105 | uwp5_sn008_ips_amundsen_2018       | marcel.babin@takuvik.ulaval.ca          | 1165 | UVP5 IPS Amundsen 2018  | CC BY 4.0    |
| 109 | uwp5_sn010_2014_eddy               | rkiko@geomar.de                         | 881  | UVP5 Geomar 2014 eddy   |              |
| 166 | uwp5_sn002_moose_dyf_2018          | coppola@obs-vlfr.fr                     | 35   | UVP5 Dyfamed 2013 2014 2015 2016 2017 2018                          |              |
| 231 | uwp5_sn202_msm060_filtered         | andreas.rogge@awi.de, anya.waite@dal.ca | 351  | UVP5hd msm060 (2017)  | Copyright    |
| 236 | uwp5_sn201_2018_naames_04_filtered | lee.karp-boss@maine.edu                 | 1252 | UVP5hd Naames 04  | CC BY-NC 4.0 |



533 **Table A2. Morphological features computed for the object (not the background). Areas and/or length units are expressed**  
534 **in pixels.**

| Feature         | Description   |
|-----------------|---|
| objid           | unique object identifier in EcoTaxa (integer number)  |
| area            | surface area of the object (integer number)   |
| mean            | average grey value within the object; sum of the grey values of all pixels in the object divided by the number of pixels  |
| stddev          | standard deviation of the grey value used to generate the mean grey value   |
| mode            | modal grey value within the object  |
| min             | minimum grey value within the object (0 = black)  |
| max             | maximum grey value within the object (255 = white)  |
| perim           | the length of the outside boundary of the object  |
| width           | width of the smallest rectangle enclosing the object  |
| height          | height of the smallest rectangle enclosing the object   |
| major           | primary axis of the best fitting ellipse for the object   |
| minor           | secondary axis of the best fitting ellipse for the object   |
| angle           | angle between the primary axis and a line parallel to the x-axis of the image   |
| circ            | circularity = $(4 * \pi * \text{Area}) / \text{Perim}^2$ ; a value of 1 indicates a perfect circle, a value approaching 0 indicates an increasingly elongated polygon                     |
| feret           | maximum feret diameter, i.e., the longest distance between any two points along the object boundary   |
| intden          | integrated density, corresponding to the sum of the grey values of the pixels within the object (i.e. = $\text{Area} * \text{Mean}$ )   |
| median          | median grey value within the object   |
| skew            | skewness of the histogram of grey level values  |
| kurt            | kurtosis of the histogram of grey level values  |
| %area           | percentage of object's surface area that is comprised of holes, defined as the background grey level  |
| area_exc        | surface area of the holes in the object, in square pixels ( $= \text{Area} * (1 - (\% \text{area} / 100))$ )  |
| fractal         | fractal dimension of object boundary (Berube and Jebrak, 1999)  |
| skelarea        | surface area of skeleton in pixels. In a binary image, skeleton is obtained by repeatedly removing pixels from the edges of objects until they are reduced to the width of a single pixel |
| slope           | slope of the grey level normalized cumulative histogram   |
| histcum 1, 2, 3 | grey level value at the first, second and third quartile of the normalized cumulative histogram of grey levels  |
| nb1, nb2, nb3   | number of remaining objects in the image after thresholding on level histcum1, 2 and 3  |
| symetrie h      | bilateral horizontal symmetry index   |
| symetrie v      | bilateral vertical symmetry index   |
| symetrie hc     | symmetry of the largest remaining object in relation to the horizontal axis after thresholding at the grey level histcum1 value   |
| symetrie vc     | symmetry of the largest remaining object in relation to the vertical axis after thresholding at the grey level histcum1 value   |
| convperim       | the perimeter of the smallest polygon within which all points in the object fit   |
| convarea        | the area of the smallest polygon within which all points in the object fit  |
| fcons           | measure of contrast based in the texture feature descriptor (Amadasun and King, 1989)   |
| thickr          | thickness ratio: relation between the maximum thickness of an object and the average thickness of the object excluding the maximum  |



|                     |   |
|---------------------|---|
| elongation          | major/minor   |
| range               | grey max - grey min   |
| meanpos             | (mean-max)/(mean-min)   |
| cv                  | 100*(stddev/mean)   |
| sr                  | 100*(stddev/(max-min))  |
| perimareaexc        | perim/(sqrt(area_exc))  |
| feretareaexc        | feret/(sqrt(area_exc))  |
| perimferet          | perim/feret   |
| perimmajor          | perim/major   |
| circex              | $(4 \cdot \pi \cdot \text{area\_exc}) / (\text{pow}(\text{perim}, 2))$      |
| kurt_mean           | mean kurtosis of the histogram of grey level values                         |
| skew_mean           | mean Skewness of the histogram of grey level values                         |
| convperim_perim     | perimeter of the smallest polygon within which all points in the object fit |
| convarea_area       | area of the smallest polygon within which all points in the object fit      |
| symetrie_h_area     | symetrie_h/area   |
| symetrie_v_area     | symetrie_v/area   |
| nb1, nb2, nb3_area  | nb1, nb2, nb3/area  |
| nb1, nb2, nb3_range | nb1, nb2, nb3/range   |
| median_mean         | median/grey mean  |
| median_mean_range   | (median-grey mean)/range  |
| skeleton_area       | skelarea/area   |



536 **Table A3. List of annotators involved in the classification of objects within the current UVP5 dataset.**

| Annotator name       | Annotator name ( <i>cont.</i> ) | Annotator name ( <i>cont.</i> ) |
|----------------------|---------------------------------|---------------------------------|
| Alikacem Yasmine     | Gawinski Christine              | Panchal Aakash                  |
| Andersson Bjoern     | Gremion Gwenaelle               | Panaïotis Thelma                |
| Anhaus Philipp       | Guérin Sébastien                | Pelletier Noemie                |
| Arboit Genevieve     | Hasnain Sarah                   | Perhirin Margaux                |
| Aubry Cyril          | Hauss Helena                    | Perron Christophe               |
| Bansept Marc-Antoine | Hintringer Moritz               | Picheral Marc                   |
| Benoît-Gagné Maxime  | Irisson Jean-Olivier            | Pretty Jess                     |
| Berline Léo          | Jalabert Laetitia               | Prondzinsky Alannah Paulina     |
| Berrenger Hugo       | Karp Boss Lee                   | Reimer Jody                     |
| Biard Tristan        | Kiko Rainer                     | Renaut Sophie                   |
| Blanc Benjamin       | Lafond Augustin                 | Rogge Andreas                   |
| Blunck Frauke        | Leguen Guillaume                | Singh Akanksha                  |
| Bourdin Guillaume    | Lekanoff Rachel                 | Singh Arvind                    |
| Caray-Counil Louis   | Leroux Riwan                    | Spahic Susanne                  |
| Chawarski Julek      | Levesque-Desrosiers Felix       | Stemmann Lars                   |
| Christiansen Svenja  | Li Juan                         | Suwaki Caroline                 |
| Courchet Lucas       | Lombard Fabien                  | Toullec Jordan                  |
| Cram Jacob           | Lopes Rubens                    | Trudnowska Emilia               |
| Denoso Ferez Katty   | Maps Frédéric                   | Vilgrain Laure                  |
| Donggyun Kim         | Marec Claudie                   | Wauthy Maxime                   |
| Drago Laetitia       | Mcdonnell Andrew M. P.          |                                 |





|                  |                 |
|------------------|-----------------|
| Dullaert Emma    | Migotto Alvaro  |
| Elineau Amanda   | Motreuil Solène |
| Faustmann Jannik | Nagata Renato   |
| Fowler Victoria  | Oliveira Bruna  |

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