

We thank the reviewers for their thoughtful feedback, which has helped us to further improve and strengthen the manuscript. Our detailed responses to the final reviewer comments are provided below in blue. All revisions are visible in the track-changes version of the revised manuscript included after the responses.

Reviewer #1

This manuscript describes the U.S. DOE ARM program's immersion-mode ice-nucleating particle (INP) dataset, spanning fixed observatories and ARM Mobile Facility (AMF) deployments, as well as vertically resolved sampling with tethered balloon systems (TBS). The paper documents sampling protocols, INS processing (including heat and H₂O₂ treatments), and automated analysis with the OLAF DaQ INS software, and points readers to data access via ARM Data Discovery. The work represents a highly valuable, standardized, multi-site record with strong potential for model evaluation, parameterization development, and process studies.

Specific comments A. Major issues

1. Sampling duration vs. frequency (Table 1; Section 2.2.1, lines 177–181): Table 1 reports ~6-day collection frequency at several sites, whereas Section 2.2.1 states that sampling is “typically after 24 hours”. Please reconcile this apparent contradiction explicitly in the text and encode, per site and period, the actual filter exposure duration in the metadata. If some sites use multi-day exposures, discuss the potential for on-filter chemical aging (photo-/heterogeneous oxidation) to bias INP spectra—particularly depressing labile organics at warm T (–5 to –15 °C) and enriching refractory INPs. Consider a sensitivity check comparing daily versus 6-day exposures during IOPs.

This wording may have been unclear as originally written. We collect 24-hour filters approximately every 6 days, resulting in ~5-day gaps between collections. To clarify, we specified this in the Table 1 footer and added the following sentence to Section 2.2.1: “As detailed in Table 1, the 24-hour samples are collected either daily or roughly every 6 days, depending on the goals and duration of the measurement campaign.” Because these are discrete 24-hour samples rather than multi-day exposures, they are not subject to potential issues of on-filter chemical aging.

2. Coordinate error (Section 2.1.1, lines 82–84; Table 1): The SGP site is given as 97.488° E, but it should be 97.488° W. Please correct this typo and ensure it propagates to Table 1, Figure 1, and metadata exports.

This was just a typo in Section 2.1.1. We have corrected it there. It was already the correct coordinate (in °W) in the map and in metadata exports.

3. Automated monotonicity correction (Section 3.3, lines 297–302; Fig. 3): Section 3.3 describes adjusting bins to enforce monotonicity when blank subtraction yields decreasing K(T). Later it says bins exceeding a threshold are “flagged with an error signal”. Please quantify how often this correction is applied per sample/site, expose a counter and QC flag in the NetCDF, and—if possible—retain pre-correction values.

As noted in Section 3.3, bins that exceed the error threshold during blank subtraction are assigned an error value of –9999. We have revised the text in this section to clarify this assignment. Regarding the retention

of pre-corrected values, these are not included in the final data product to avoid potential confusion or inadvertent use of uncorrected data instead of the finalized QA/QC-processed values.

It is important to note that OLAF was developed and fully operational within the past year, and has only been applied to recent datasets, including CRG, BNF, and recent SGP samples; earlier datasets were processed manually. We already stated this in the manuscript at the beginning of Section 3.3 but included this statement in the newly-added text for clarity: “This correction was not applied prior to OLAF when files were generated manually.”

We agree that implementing a monotonicity QC flag in the NetCDF files is a valuable improvement and will begin adding this feature to existing OLAF-generated files, as well as all future datasets. We added a statement to Section 3.3 delineating this: “We are currently developing an additional QC flag for OLAF-generated data files to indicate which data points were adjusted due to stochasticity-related corrections.”

4. Uncertainty budget (Equation 1, Section 3.1, lines 232–239): Beyond Agresti–Coull intervals, please include systematic and volumetric terms: flow meter accuracy ($\pm 2\%$), temperature measurement (± 0.2 °C, including block gradients), droplet volume tolerance, and suspension volume uncertainties. Discuss edge cases (0/32 and 32/32) and how LOD/LOQ are reported. Providing per-bin combined uncertainties will improve usability.

We appreciate the reviewer’s suggestion to expand the uncertainty budget beyond the Agresti–Coull intervals. We have now included additional sources of systematic and volumetric uncertainty at the end of Section 3.1. Specifically:

- Flow meter accuracy: $\pm 4\%$, based on the 5200 Series Gas Flow Multi-Meter Operation and Service Manual (TSI, US).
- Thermocouple accuracy: Type T thermocouples are estimated at ± 0.5 °C, though relative temperature uncertainty between data points is closer to ± 0.1 °C (Perkins et al., 2020).
- Block temperature gradients: No measurable gradient has been observed in previous lab tests. Uncertainty is sufficiently captured by thermocouple range.
- Droplet volume tolerance:
 - Larger pipette used for DI water in dilutions: $\pm 1.3\%$
 - Smaller pipette used for “sample” in dilutions: $\pm 2.5\%$
 - Multipipetter: $\pm 1.8\%$
- Suspension volume uncertainty: Currently dispensed using 20 mL syringes; a gravimetric test was conducted in the CSU lab to quantify this uncertainty and ranges from $\pm 0.1\%$ to $\pm 0.8\%$ depending on the dispensed volume.

We also added clarification on edge cases and detection limits:

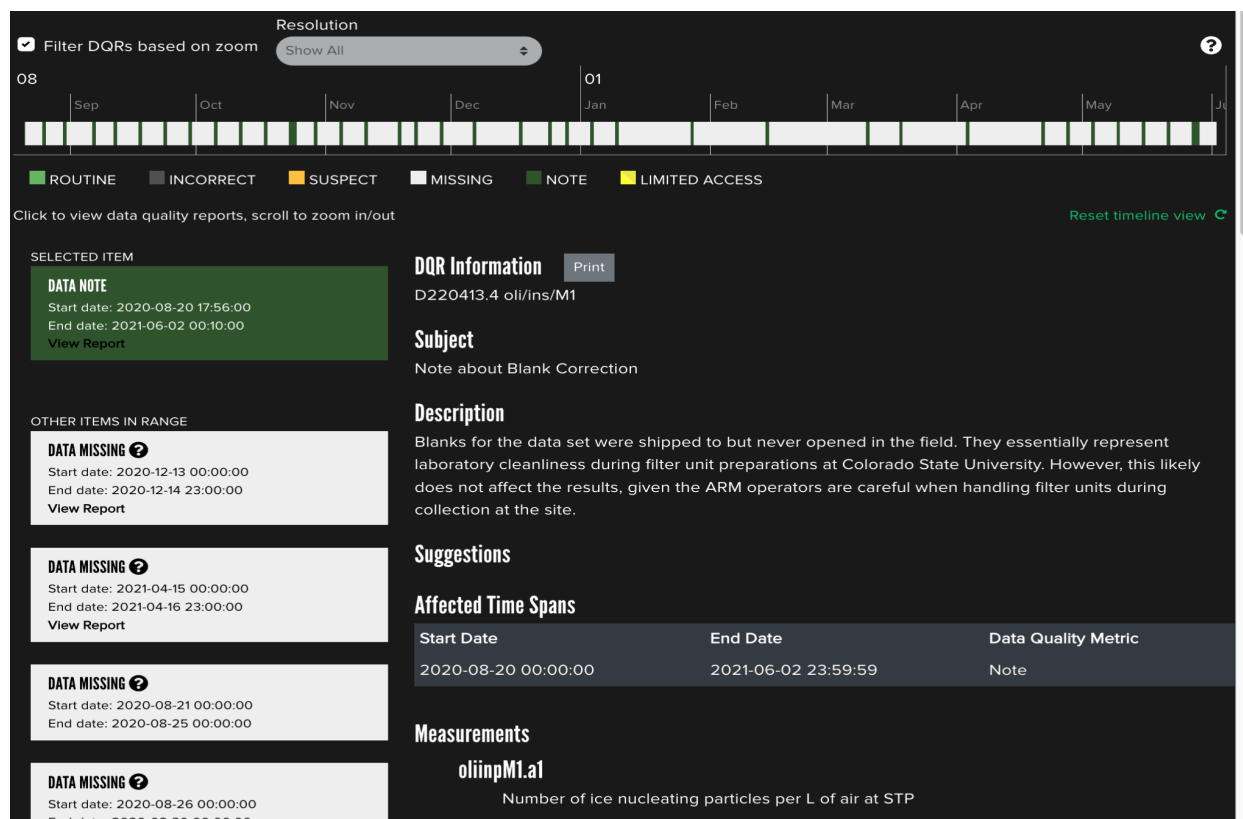
- 0/32 and 32/32 freezing cases are not reported.
- The LOD varies by sample depending on the total air and suspension volumes. Values below detection are reported as –9999.

Perkins, R. J., Gillette, S. M., Hill, T. C. J., and DeMott, P. J.: The Labile Nature of Ice Nucleation by Arizona Test Dust, ACS Earth Space Chem., 4, 133–141, <https://doi.org/10.1021/acsearthspacechem.9b00304>, 2020.

- Blank strategy and Oliktok (Sections 2.1.2–2.1.3, Table 1; Section 3.2.1, lines 254–255): Clarify, in a table and in the data files, which sites have field versus lab blanks. For Oliktok (OLI), state explicitly that only lab blanks were available and flag affected samples in the data (e.g., “blank_type = lab-only; use with caution”).

All sites include both field and deionized water blanks; the only exception is OLI, where field blanks were not collected due to operational issues by the technicians at the site. In that case, laboratory blanks were used instead. Since this information would be largely redundant in the table, we instead stated it at the end of Section 3.2.1: “All sites undergo field blank collection approximately once per month, with the exception of OLI M1, which is detailed in a Data Quality Report (DQR) that can be viewed under the “Description” for this site’s INP data on the ARM Data Discovery Portal. Moving forward, all existing and new sites in the program will include routine monthly field blanks.”

In lieu of a flag, under “Description” for OLI after selecting “Details” and clicking on the timeline, we already note on ARM Data Discovery: “Blanks for the data set were shipped to but never opened in the field. They essentially represent laboratory cleanliness during filter unit preparations at Colorado State University. However, this likely does not affect the results, given the ARM operators are careful when handling filter units during collection at the site.” See screen shot of the website below from <https://adc.arm.gov/discovery/#!/results/s::inp>.



6. TBS sampling details (Section 2.1.3, lines 144–152; Table 2): For TBS deployments, please document the standard conditions implied by the logged volumetric flow (Slpm) and any conversions applied to STP volumes at altitude. Include face velocity across the filter, residence time per filter/altitude, flow stability during ascents/descents, and the barometric/temperature corrections used. If these are detailed in Dexheimer et al. (2024), cross-reference explicitly.

Since this paper has undergone review, we have published a separate paper detailing the TBS system, its deployments, and associated data (Creamean et al., 2025). Consequently, we removed Table 2 and revised the final sentence of Section 2.1.3 to: “The TBS INP sampler design, filter preparations, deployments, and available data are described in detail in Creamean et al. (2025) and are only briefly mentioned here.” We also removed Section 2.2.2, which focused on filter preparations for TBS deployments, as this is fully covered in our more recent paper. Information on how to access TBSINP data was retained, as it is essential to the available dataset and relevant to this data paper.

Creamean, J. M., Dexheimer, D., Hume, C. C., Vazquez, M., Hess, B. T. M., Longbottom, C. M., Ruiz, C. A., and Theisen, A. K.: Reaching new heights: A vertically-resolved ice nucleating particle sampler operating on Atmospheric Radiation Measurement (ARM) tethered balloon systems, EGUsphere, 1–22, <https://doi.org/10.5194/egusphere-2025-5000>, 2025.

B. Moderate suggestions

Positive control (Section 3.2.3, lines 272–283): Report periodic measurements of a standard INP material (e.g., Snomax, illite NX) to track sensitivity drift.

We do not use positive controls with a standard INP material to assess instrumental drift. Instead, potential drift or other issues (including those related to deionized water quality) are monitored through the deionized water blanks that are run alongside every sample. Thus the deionized water blanks serve a dual purpose. If a deionized water blank appears abnormal relative to expectations, we investigate potential issues by re-running the sample and water blank, evaluating possible sources of contamination, and/or checking whether INS components (i.e., thermocouples, cameras, etc.) may be malfunctioning. To make this clear, we added the following to Section 3.2.3: “The deionized water blanks run with every sample serve a dual purpose: they act as a positive control as well and help monitor instrument drift, potential contamination, and the proper functioning of INS components (e.g., thermocouples, cameras).”

Cross-site comparability (Figures 6–7; Section 4.3, lines 358–364): Daily sampling resolves episodic events better than 6-day routine sampling. Note this caveat and consider a sensitivity test subsampling daily periods to pseudo-6-day to quantify biases in seasonal boxplots.

See response to Comment 1. The samples are not 6-day integrations; rather, they are 24-hour samples collected approximately every 6 days. Therefore, episodic events (at least, those more than 24-hours) are already captured within these figures.

Treatment fractions (Section 2.3.1, line 224–225; Fig. 5, lines 342–346): State, per site and season, the fraction and number of samples undergoing heat and H₂O₂ treatments. Add treatment identifiers to records.

Due to the ongoing nature of this program, the numbers of treatments change continuously on a weekly basis. Readers can, however, view real-time statistics of the treatments at each site and the exact dates

when they were applied via the field log on our website. For example, the first entry for BNF as of 2 Oct 2025 shows totals for base (no treatments), heat, and peroxide at the top of the log. Because of this constant evolution, including treatment numbers in the manuscript would be outdated by the time of publication and thus not meaningful. To clarify this in the text, we added the following sentences to the end of Section 2.3.1, in the paragraph describing processing of one-third of the samples at most sites: “Due to the ongoing nature of this program, the numbers of treatments conducted on samples from any given site evolve continuously on a weekly basis. Exact sample sites, dates, identifiers, and other metadata regarding which samples undergo treatments can be accessed in real time via the publicly available field log on the INS website.”

Treatment identifiers are already included in the records/data files as flags, as described in Section 3.1. We have added clarification in that section specifying the meaning of each flag: untreated = 0, heat treated = 1, and peroxide treated = 2.

52	40	40	8	8	nr = not recorded nc = not collected red = value was high blue = value was low																
filter #	data processed	base	heat	H2O2	checklist date	start (UTC)	end (UTC)	A start flow (slpm)	A start P (kPa)	A start T (C)	A end flow (slpm)	A end P (kPa)	A end T (C)	A total vol (L)	B start flow (slpm)	B start P (kPa)	B start T (C)	B end flow (slpm)	B end P (kPa)	B end T (C)	B total vol (L)
1	✓	✓	□	□	10/16/24	10/01/24 14:05	10/02/24 14:05	14.46	47.80	22.8	11.87	42.93	23.90	1,998.90	14.70	46.90	24.4	12.25	39.95	23.8	19,940.8
2	✓	✓	✓	□	10/16/24	10/07/24 14:03	10/08/24 14:03	13.70	48.59	23.3	12.29	43.44	22.80	19,976.00	14.61	50.57	24.1	14.11	42.33	23.2	22,948.5
3	✓	✓	□	□	10/16/24	10/15/24 14:09	10/16/24 13:58	13.12	54.96	22.9	13.58	45.97	23.40	20,433.60	12.32	54.64	23.4	13.14	42.29	24.1	19,893.5
4	✓	✓	✓	✓	10/22/24	10/21/24 14:07	10/22/24 14:02	14.56	53.65	19.7	12.13	44.32	21.90	21,592.60	13.99	53.47	20.4	9.93	38.77	22.7	20,315.0
5	✓	✓	✓	□	10/28/24	10/27/24 14:01	10/28/24 13:57	12.74	55.62	20.9	8.52	39.77	23.60	17,544.00	15.80	55.06	21.5	10.25	38.96	24.0	21,314.9
6	✓	✓	✓	✓	11/04/24	11/02/24 14:10	11/03/24 14:10	13.03	49.08	23.2	7.01	37.81	22.90	15,854.50	14.83	51.61	23.9	9.29	38.34	23.5	18,920.7
7	✓	✓	□	□	11/21/24	11/08/24 14:54	11/09/24 15:00	14.68	47.95	21.0	10.82	43.08	23.10	18,955.80	15.36	46.46	21.6	11.55	40.58	23.5	20,146.0
8	✓	✓	✓	✓	11/21/24	11/14/24 14:55	11/15/24 14:55	9.27	51.34	21.1	9.44	41.48	22.90	14,192.50	13.58	54.75	21.7	13.58	43.15	22.6	21,331.7
9	✓	✓	□	□	11/21/24	11/20/24 14:59	11/21/24 15:00	8.68	47.90	21.1	9.60	42.45	22.50	13,966.20	14.66	51.00	21.6	15.35	45.60	22.3	22,593.3
10	✓	✓	✓	✓	12/03/24	12/02/24 14:46	12/03/24 14:56	9.21	50.55	22.3	9.05	43.36	19.90	13,905.60							
11	✓	✓	✓	✓	12/09/24	12/08/24 16:24	12/09/24 16:23	9.36	44.39	20.4	7.64	38.92	24.30	12,125.60	14.36	44.51	20.2	9.50	38.01	24.4	16,915.0
12	✓	✓	□	□	12/23/24	12/20/24 14:40	12/21/24 14:40	14.84	51.58	21.7	12.49	46.62	19.70	20,522.60	14.41	50.10	21.8	15.89	45.73	22.4	155.8

Software versioning (Section 3.3, line 286): Cite a DOI/Zenodo release or commit hash for OLAF DaQ INS and include version in dataset metadata.

We do not have a DOI or Zenodo release for OLAF, so we provided the GitHub link where it is already available as open-source software. We clarified this by revising the sentence in the sentence in Section 3.3 to: “More information and the software itself are available at: <https://github.com/SiGran/OLAF>.”

C. Minor and editorial corrections

DOIs (lines 432 and 450): Several DOIs are written as <https://doi.org/...> (missing colon). Please correct to <https://doi.org/...>

Done.

Typo (Table 3): Remove duplicated word 'the the ARM G-1'.

Done.

Purge gas (line 213): Replace 'pre-cooled slightly above block temperature' with 'pre-cooled near block temperature'.

Done.

Figures: 4 (lines 318–330): Add number of samples per line.

Each line is based on the same number of samples from SGP, as these are data points from cumulative spectra. We now indicate in the figure caption that these data are from 247 total processed filter samples.

Fig. 5 (lines 342–346): Clarify number of treated samples.

We now indicate in the figure caption that these data are from 84 treated samples (34% of the processed SGP samples in Figure 4).

Figs. 6–7 (lines 358–364): Add sample counts per boxplot and note filter durations.

Done. All filter durations are 24-hour as indicated in the comments above.

Tables: Table 1: add typical duration per filter and blank type.

Added to footnote.

Table 2 (line 153): add residence-time assumptions.

See response to Comment #6.

I recommend “major revisions.” The manuscript’s core contribution is strong, but resolving sampling-duration ambiguity, fully documenting automated corrections/QC, and expanding the uncertainty budget are essential for a durable ESSD dataset paper.

Reviewer #2

This data description paper by Creamean et al. offers a bodacious ice-nucleating particle dataset. The manuscript is well-written and fulfills the journal's scope. This reviewer recommends publishing this paper for ESSD after the authors address the following comments.

The authors did a good job showing applications of their INP data. How about limitations? What do the data users need to be aware of when they apply the offered data in observation-driven Earth system models etc. on different spatial-temporal scales? Perhaps, the authors might address it according to the challenges discussed in Burrows et al. (2022; DOI - <https://doi.org/10.1029/2021RG000745>)? This reviewer believes that clearly stating limitations is as important as demonstrating the applicability of any data.

The reviewer brings up a very valid point. We changed the title of Section 5 to “Community use and limitations of ARM INP data” and have added the following text: “The DOE ARM baseline INP measurements provide valuable long-term and IOP-based observations but have several limitations that users should be aware of. First, these measurements do not account for time dependence in freezing behavior, which is generally less significant than temperature dependence (Ervens and Feingold, 2013). Second, sampling assumes collection of the total aerosol size distribution; however, this has not been

explicitly tested, so the exact size range collected is uncertain. The 0.2- μm filters we use have reduced transmission efficiency for particles around 150 nm (down to 65–78%), but generally exhibit very high collection efficiency at most sizes (Spurny and Lodge, 1972). Third, because filters are collected over 24 hours, typically every six days, short-term or episodic INP events may be missed, although higher-frequency sampling can be requested. Fourth, these measurements are made at the surface and may not fully represent the INP population at cloud level, though cloud-surface coupling analyses (e.g., Creamean et al., 2021; Griesche et al., 2021) and TBS INP data (Creamean et al., 2025) can help bridge this gap. Lastly, not all samples are subjected to treatments unless requested, and, as noted by Burrows et al. (2022), the absence of co-located baseline measurements of aerosol composition (particularly dust, sea spray, and primary biological aerosol particles) limits the ability to fully constrain INP sources and improve parameterizations in models.”

L66-67: It seems controversial that the authors raise the concern of “not routine” INP measurements here, while they offer several single IOP data in this manuscript. The authors might want to rephrase this sentence; otherwise, clarify the concern rigorously.

Good point. What we intended to emphasize is that ARM now offers INP measurements as a baseline product. Accordingly, we revised the sentence to: “While INP measurements have been conducted at various ARM sites in the past, they were primarily user-driven and not part of the baseline measurement suite.” We also changed “routine” on line 72 to “baseline.”

L180-181: Has the time span between collection and analysis been consistent for all samples? If not, this reviewer would like to see if the authors can discuss the impact of various sample storage intervals.

The time between collection and analysis has varied from as little as one week to over a year. Beall et al. (2020) reported no significant differences in INP concentrations for samples stored between 1 and 166 days. For select samples re-analyzed as part of our internal quality checks, we observed minimal differences (<1%) after one to two and a half years of storage at -20°C . In cases where re-analysis was required due to issues with the original measurements, differences of 20 to 60% were observed at some temperature bins; however, these represent outliers associated with problematic initial samples. We added this text to Section 2.3: “The time between collection and analysis has ranged from one week to over a year. Beall et al. (2020) reported no significant differences in INP concentrations for samples stored between 1 and 166 days. In our internal quality checks, select samples stored for 1–2.5 years at -20°C showed minimal differences (<1%), while larger differences (20–60%) were observed only in outlier cases associated with problematic original measurements.”

L220-221: What is the rational procedure of H_2O_2 application for organic removal? This reviewer is aware that the H_2O_2 concentration ranges (e.g., Perkins et al., 2020: DOI - 10.1021/acsearthspacechem.9b00304). It seems the peroxide digestion method is operational without verification. The authors might consider including a brief yet clear statement of what needs to be investigated in terms of H_2O_2 application down the road in the manuscript. Doing this will benefit the community and mitigate the concerns of readers.

The use of H_2O_2 for degrading organic matter in soils dates back to Robinson (1922), who first introduced it for soil texture analysis (Mikutta et al., 2005). Since then, it has become the most widely used chemical

reagent for organic matter destruction prior to textural and mineralogical analyses. Typical treatments employ 30% (w/w) H₂O₂, though concentrations ranging from 6% to 50% have been used with little difference in carbon removal efficiency (McLean, 1931), likely due to rapid H₂O₂ decomposition and the persistence of mineral-protected organic compounds. Early protocols began at room temperature to accommodate vigorous reactions with easily decomposable material, followed by heating (60–90 °C) to accelerate oxidation (Schultz et al., 1999). Despite its century-long use, there remains no standardized protocol for H₂O₂ treatment in soil science, with reported concentrations, reaction temperatures, and contact times varying widely depending on research objectives and soil properties (e.g., Sequi and Aringhieri, 1977).

In the presence of UV, H₂O₂ photolyzes, producing hydroxyl radicals (H₂O₂ + hv (UV) → 2 ·OH). The resulting hydroxyl radicals are highly reactive and can attack functional groups on polysaccharides, open sugar rings, and oxidize carbonyl (–C=O) or carboxylic acid (–COOH) groups (e.g., Chen et al., 2021; Ofoedu et al., 2021). Such reactions can disrupt molecular integrity and crystallographic order, oxidize or rearrange hydrophilic groups, and alter polymer conformation (e.g., through unwinding, depolymerization, or cross-linking). These structural modifications can diminish ice-nucleating activity of bio-organic materials by reducing the number of effective ice-active sites, or, if extensive enough, completely eliminate activity by destroying the structural motifs responsible for ice templating (DeMott et al., 2023; Gute and Abbatt, 2018).

Within the INP community, treatments with 10% H₂O₂ are most commonly used, although concentrations ranging from 10% to 35% have been reported (e.g., Barry et al., 2023a; Beall et al., 2022; Hill et al., 2016; McCluskey et al., 2018; O’Sullivan et al., 2014; Roy et al., 2021; Suski et al., 2018; Tesla et al., 2024; Testa et al., 2021; Tobo et al., 2019). Perkins et al. (2020) used 15% H₂O₂ and heated to 95 °C, but organics in their ATD tests were found to be thermally stable to 500 °C without peroxide digestion. In our recent tests comparing 10% and 20% treatments on the same samples, we observed minimal differences in ice nucleating activity; however, additional experiments are needed to confirm this result.

As suggested by the reviewer, we have provided this text to the section 2.3.1: “H₂O₂ has long been used as an oxidizing agent for degrading organic matter, originating in soil science nearly a century ago and later adopted across disciplines for removing organic material prior to chemical or physical analyses (McLean, 1931; Mikutta et al., 2005; Robinson, 1922; Schultz et al., 1999; Sequi and Aringhieri, 1977). In the presence of UV light, H₂O₂ photolyzes to form highly reactive hydroxyl radicals that can oxidize and structurally modify organic macromolecules, diminishing or eliminating their ice-nucleating activity (DeMott et al., 2023; Gute and Abbatt, 2020). Within the INP community, H₂O₂ treatments typically range from 10 to 35% (Beall et al., 2022; O’Sullivan et al., 2014; Perkins et al., 2020; Roy et al., 2021; Teska et al., 2024; Tobo et al., 2019). We conducted recent tests showing minimal differences in ice-nucleating activity between 10% and 20% treatments; however, further validation is needed, and future community efforts should aim to establish a standardized protocol and concentration to ensure methodological consistency across studies.”

References:

Beall, C. M., Hill, T. C. J., DeMott, P. J., Könenan, T., Pikridas, M., Drewnick, F., Harder, H., Pöhlker, C., Lelieveld, J., Weber, B., Iakovides, M., Prokeš, R., Sciare, J., Andreae, M. O., Stokes, M. D., and

Prather, K. A.: Ice-nucleating particles near two major dust source regions, *Atmospheric Chemistry and Physics*, 22, 12607–12627, <https://doi.org/10.5194/acp-22-12607-2022>, 2022.

Chen, X., Sun-Waterhouse, D., Yao, W., Li, X., Zhao, M., and You, L.: Free radical-mediated degradation of polysaccharides: Mechanism of free radical formation and degradation, influence factors and product properties, *Food Chemistry*, 365, 130524, <https://doi.org/10.1016/j.foodchem.2021.130524>, 2021.

DeMott, P. J., Hill, T. C. J., Moore, K. A., Perkins, R. J., Mael, L. E., Busse, H. L., Lee, H., Kaluarachchi, C. P., Mayer, K. J., Sauer, J. S., Mitts, B. A., Tivanski, A. V., Grassian, V. H., Cappa, C. D., Bertram, T. H., and Prather, K. A.: Atmospheric oxidation impact on sea spray produced ice nucleating particles, *Environ. Sci.: Atmos.*, 3, 1513–1532, <https://doi.org/10.1039/D3EA00060E>, 2023.

Gute, E. and Abbatt, J. P. D.: Oxidative Processing Lowers the Ice Nucleation Activity of Birch and Alder Pollen, *Geophys. Res. Lett.*, 45, 1647–1653, <https://doi.org/10.1002/2017GL076357>, 2018.

McLean, W.: The Nature of Soil Organic Matter as Shown by the attack of Hydrogen Peroxide, *The Journal of Agricultural Science*, 21, 595–611, <https://doi.org/10.1017/S0021859600009813>, 1931.

Mikutta, R., Kleber, M., Kaiser, K., and Jahn, R.: Review: Organic Matter Removal from Soils using Hydrogen Peroxide, Sodium Hypochlorite, and Disodium Peroxodisulfate, *Soil Science Society of America Journal*, 69, 120–135, <https://doi.org/10.2136/sssaj2005.0120>, 2005.

O’Sullivan, D., Murray, B. J., Malkin, T. L., Whale, T. F., Umo, N. S., Atkinson, J. D., Price, H. C., Baustian, K. J., Browse, J., and Webb, M. E.: Ice nucleation by fertile soil dusts: relative importance of mineral and biogenic components, *Atmospheric Chemistry and Physics*, 14, 1853–1867, <https://doi.org/10.5194/acp-14-1853-2014>, 2014.

Ofoedu, C. E., You, L., Osuji, C. M., Iwouno, J. O., Kabuo, N. O., Ojukwu, M., Agunwah, I. M., Chacha, J. S., Muobike, O. P., Agunbiade, A. O., Sardo, G., Bono, G., Okpala, C. O. R., and Korzeniowska, M.: Hydrogen Peroxide Effects on Natural-Sourced Polysaccharides: Free Radical Formation/Production, Degradation Process, and Reaction Mechanism—A Critical Synopsis, *Foods*, 10, 699, <https://doi.org/10.3390/foods10040699>, 2021.

Robinson, G. W.: Note on the mechanical analysis of humus soils, *The Journal of Agricultural Science*, 12, 287–291, <https://doi.org/10.1017/S0021859600005347>, 1922.

Roy, P., House, M., and Dutcher, C.: A Microfluidic Device for Automated High Throughput Detection of Ice Nucleation of Snomax®, *Micromachines*, 12, 296, <https://doi.org/10.3390/mi12030296>, 2021.

Schultz, M. K., Biegalski, S. R., Inn, K. G. W., Yu, L., Burnett, W. C., Thomas, J. L. W., and Smith, G. E.: Optimizing the removal of carbon phases in soils and sediments for sequential chemical extractions by coulometry, *J. Environ. Monit.*, 1, 183–190, <https://doi.org/10.1039/A900534J>, 1999.

Sequi, P. and Aringhieri, R.: Destruction of Organic Matter by Hydrogen Peroxide in the Presence of Pyrophosphate and Its Effect on Soil Specific Surface Area, *Soil Science Society of America Journal*, 41, 340–342, <https://doi.org/10.2136/sssaj1977.03615995004100020033x>, 1977.

Teska, C. J., Dieser, M., and Foreman, C. M.: Clothing Textiles as Carriers of Biological Ice Nucleation Active Particles, *Environ. Sci. Technol.*, 58, 6305–6312, <https://doi.org/10.1021/acs.est.3c09600>, 2024.

Tobo, Y., Adachi, K., DeMott, P. J., Hill, T. C. J., Hamilton, D. S., Mahowald, N. M., Nagatsuka, N., Ohata, S., Uetake, J., Kondo, Y., and Koike, M.: Glacially sourced dust as a potentially significant source of ice nucleating particles, *Nat. Geosci.*, 12, 253–258, <https://doi.org/10.1038/s41561-019-0314-x>, 2019.

L232: Missing a negative sign on RHS in Eqn (1)?

Thanks for catching that typo. We have fixed it.

L233: How do the authors determine $V_{\text{suspension}}$ values? Please clarify in the manuscript.

We have clarified this in Section 3.1: “Specifically, the $V_{\text{suspension}}$ is the volume of 0.1 μm –filtered deionized water used to resuspend the particles from the filter (7–10 mL).”

1 Long-term measurements of ice nucleating particles at Atmospheric 2 Radiation Measurement (ARM) sites worldwide

3 Jessie M. Creamean¹, Carson Hume¹, Maria Vazquez¹, Adam Theisen²

4 ¹Department of Atmospheric Science, Colorado State University, Fort Collins, Colorado, 80523, USA

5 ²Argonne National Laboratory, Lemont, Illinois, 60439, USA

6 *Correspondence to:* Jessie M. Creamean (jessie.creamean@colostate.edu)

7 Abstract

8 Ice nucleating particles (INPs) play a critical role in cloud microphysics and precipitation formation, yet long-term, spatially
9 extensive observational datasets remain limited. Here, we present one of the most comprehensive publicly available datasets
10 of immersion-mode INP concentrations using a single analytical method, generated through the U.S. Department of Energy's
11 (DOE) Atmospheric Radiation Measurement (ARM) user facility. INP filter samples have been collected across a broad range
12 of environments—including agricultural plains, Arctic coastlines, high-elevation mountain sites, marine regions, and urban
13 areas—via fixed observatories, mobile facility deployments, and vertically-resolved tethered balloon system operations. We
14 describe the standardized processing and quality assurance pipeline, from filter collection and processing using the Ice
15 Nucleation Spectrometer to final data products archived on the ARM Data Discovery portal. The dataset includes both total
16 INP concentrations and selectively treated samples, allowing for classification of biological, organic, and inorganic INP types.
17 It features a continuous 5-year record of INP measurements from a central U.S. site, with data collection still ongoing. Seasonal
18 and site-specific differences in INP concentrations are illustrated through intercomparisons at -10°C and -20°C , revealing
19 distinct regional sources and atmospheric drivers. We also outline mechanisms for researchers to access existing data, request
20 additional sample analyses, and propose future field campaigns involving ARM INP measurements. This dataset supports a
21 wide range of scientific applications, from observational and mechanistic studies to model development, and provides critical
22 constraints on aerosol-cloud interactions across diverse atmospheric regimes.

23 Short summary

24 This study presents a comprehensive, publicly available ice nucleating particles (INP) dataset from the U.S. Department of
25 Energy Atmospheric Radiation Measurement (ARM) user facility across diverse environments, including Arctic, agricultural,
26 urban, marine, and mountainous sites. Samples are collected via fixed and mobile platforms and processed using a standardized
27 pipeline. The dataset supports observational and modelling analyses of seasonal, spatial, and compositional variability in INPs.

28 1 Introduction

29 The formation and microphysical evolution of cloud droplets and ice crystals are strongly influenced by aerosols acting as
30 cloud condensation nuclei (CCN) and ice nucleating particles (INPs). While INP observations remain sparse compared to other
31 aerosol properties, they are essential for understanding aerosol-cloud interactions and their impacts on cloud microphysics and
32 radiative properties. Immersion freezing—where an INP first acts as a CCN before freezing at temperatures above
33 homogeneous freezing (-38°C)—is particularly important for mixed-phase cloud formation (Kanji et al., 2017; Knopf and
34 Alpert, 2023).

35 An aerosol’s ability to serve as an INP depends on temperature, vapor saturation with respect to water and ice, and particle
36 properties such as composition (chemical, mineral, or biological), morphology, and size, all of which are linked to its source
37 (Hoose and Möhler, 2012). Known atmospheric INPs include mineral dust, soil dust, sea spray, volcanic ash, black carbon,
38 and a range of biological particles (e.g., bacteria, fungal spores, pollen, algae, lichens, macromolecules) (e.g., Conen et al.,
39 2011; Creamean et al., 2013, 2019; Cziczo et al., 2017; DeMott, 1990; DeMott et al., 2016, 2018c; Hill et al., 2016; Huang et
40 al., 2021; Kaufmann et al., 2016; Levin et al., 2010; McCluskey et al., 2017; O’Sullivan et al., 2014, 2016). Among natural
41 INPs, mineral dust and biological particles are especially important. Dust is prevalent and typically active below -15°C , while
42 some biological particles, such as specific bacteria, can initiate freezing at temperatures as high as -1.5°C (Després et al.,
43 2012; Huang et al., 2021; Janine Fröhlich-Nowoisky et al., 2016; Schnell and Vali, 1976; Vali et al., 1976). Quantifying total
44 INPs, as well as distinguishing their biological and mineral fractions, provides critical insight into INP sources and atmospheric
45 abundances.

46 Although offline drop freezing assay techniques have been employed for decades, recent intercomparison studies (DeMott et
47 al., 2017, 2018d, 2025a; Lacher et al., 2024; Wex et al., 2015) affirm their effectiveness for ambient INP sampling. These
48 methods are particularly valuable because they often capture INP concentrations across nearly the full heterogeneous freezing
49 temperature range. Their simplicity makes them well-suited for long-term and remote deployments, as filters or other sample
50 types can be easily collected and later analyzed offline. Long-term, multi-year INP records are critical for improving the
51 representation of INP sources and their temporal evolution in earth system models (Burrows et al., 2022). Schrod et al. (2020)
52 presented long-term measurements of deposition and condensation mode INPs from six diverse climatic regions, including the
53 Amazon, Caribbean, central Europe, and the Arctic. Their near-continuous 24-hour samples—analyzed at -20 , -25 , and $-$
54 30°C —spanned over two years in some locations and showed relatively consistent INP concentrations across sites, generally
55 within one order of magnitude. Similarly, Wex et al. (2019) reported comparable INP levels across multiple Arctic coastal
56 sites, though they observed strong seasonal variability spanning several orders of magnitude, largely driven by the presence or
57 absence of snow and sea ice. Freitas et al. (2023) documented a four-year record of Arctic INPs in Svalbard, which peaked
58 during summer in conjunction with increased fluorescent biological particles. Schneider et al. (2021) reported 14 months of
59 INP data from a Finnish boreal forest, showing seasonal alignment with primary biological aerosol particles (PBAPs),

Deleted: freezing

61 including pollen. Gratzl et al. (2025) further linked seasonal INP fluctuations in the European sub-Arctic to fungal spores,
62 particularly Basidiomycota, over the course of a year.

63 As recent studies have shown, long-term INP monitoring is especially powerful when integrated with detailed aerosol
64 properties—such as mass concentration, size distribution, chemical composition, and optical characteristics—routinely
65 measured by global in situ monitoring networks. The U.S. Department of Energy’s Atmospheric Radiation Measurement
66 (ARM) user facility is particularly well-suited for this purpose, with fixed sites and extended-duration mobile deployments
67 that span a range of environments from the Arctic to the midlatitudes and the southern hemisphere. While INP measurements
68 have been conducted at various ARM sites in the past, they were primarily user-driven and not part of the baseline measurement
69 suite. These efforts have provided critical insights, including INP closure studies that reveal discrepancies between observed
70 and predicted INPs, highlighting the need for improved parameterizations that may be missing key INP types (Knopf et al.,
71 2021).

Deleted: routine

72 Recently, ARM has begun implementing baseline INP measurements at select sites, with coverage growing both spatially and
73 temporally. The most extensive record to date spans nearly five years at ARM’s fixed observatory in Oklahoma, USA. This
74 paper outlines the availability of the valuable datasets at ARM sites, describing the sampling and offline analysis methods,
75 data quality assurance pipelines, and access for the broader scientific community. A key aim is to raise awareness of these
76 resources beyond current ARM users and encourage broader utilization by both experimentalists and modelers.

Deleted: routine

Deleted: —

77 **2 Sample collection and processing**

78 **2.1 ARM sites with existing INP measurements**

79 **2.1.1 Fixed sites**

80 Locations where INP measurements have been conducted or are currently underway are shown in Figure 1, with corresponding
81 start and end dates, and filter collection frequency, listed in Table 1. For more up-to-date information on ARM observatories,
82 visit <https://www.arm.gov/capabilities/observatories>. Detailed information on INP sampling, including field logs and filter
83 metadata, is available at <https://www.arm.gov/capabilities/instruments/ins>. Filter samples are currently collected on a routine
84 basis approximately every 6 days at two of the three fixed atmospheric observatories: the Southern Great Plains Central Facility
85 in Lamont, Oklahoma (SGP C1; 314 m AMSL, 36.607° N, 97.488° W) and the North Slope of Alaska Central Facility in
86 Utqiagvik, Alaska (NSA C1; 8 m AMSL, 71.323° N, 156.615° W). Routine filter collections began at SGP C1 in October 2020
87 and are ongoing indefinitely, making it the first site globally with nearly five years of continuous INP measurements. At NSA
88 C1, filter collection commenced in June 2025 and is likewise planned as a long-term effort.

Deleted: E

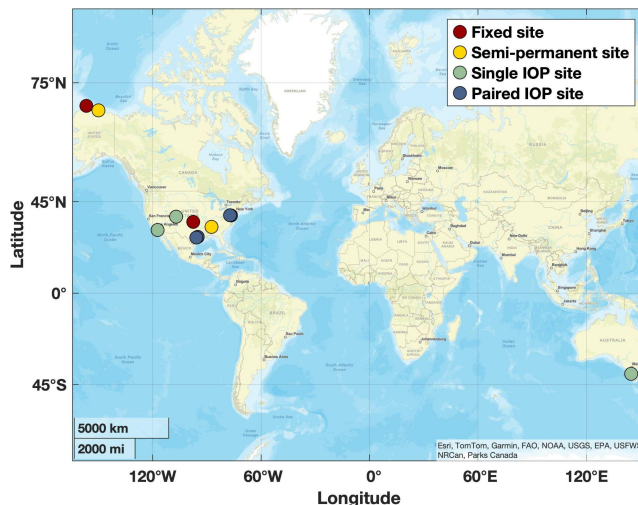


Figure 1. Map of U.S. Department of Energy Atmospheric Radiation Measurement (DOE ARM) user facility sites where routine INP measurements have been established. Red markers show fixed observatories, including Southern Great Plains (SGP C1) and North Slope of Alaska (NSA C1). ARM Mobile Facility (AMF) deployments are shown by yellow markers, while green and blue markers show IOP AMF deployment locations with single and paired sites, respectively. Paired sites indicate IOPs where main and supplemental site locations had simultaneous sample collections. Fixed and semi-permanent sites have single sample collection locations. See Table 1 for site details. Map was generated using Matlab with data from the Environmental Systems Research Institute.

An Intensive Observational Period (IOP) campaign, AGINSGP (Agricultural Ice Nuclei at SGP; Burrows, 2023), was conducted from September 2021 to May 2022. The objective of this deployment was to collect observations to better understand the drivers of variability in INP concentrations at the SGP locale, which are hypothesized to be influenced in part by regional emissions from fertile, organic-rich agricultural soils. Scientific users can submit requests to ARM to implement enhanced sampling strategies—such as increased temporal resolution, additional sampling sites, or entirely new locations—similar to the approach used during AGINSGP. Throughout the campaign, INP filters were collected approximately daily to support case study analyses following the field observations.

Table 1. List of DOE ARM sites with INP measurements. Also included are start and end dates and collection frequency of INP filters. Sites are indicated as either fixed, AMF, or ARM user-requested IOP (Intensive Observing Period). Sites that are continuous are labeled as such in the “INP filter end” column and those with “tbd” indicate an end date has yet to be determined.

Site name	Site type	Site ID	INP filter start	INP filter end	Filter collection frequency
-----------	-----------	---------	------------------	----------------	-----------------------------

Southern Great Plains (SGP) Central Facility	fixed	SGP C1	Oct 2020	continuous	every 6 days
North Slope of Alaska (NSA) Central Facility	fixed	NSA C1	Jun 2025	continuous	every 6 days
Agricultural Ice Nuclei at SGP (AGINSGP)	IOP	SGP C1	Apr 2022	Apr 2022	daily
Oliktok Point (OLI) Main Site	AMF	OLI M1	Aug 2020	Jun 2021	every 6 days
Bankhead National Forest (BNF) Main Site	AMF	BNF M1	Oct 2024	tbd	every 6 days
Surface Atmosphere Integrated Field Laboratory (SAIL) Main Site	AMF	GUC M1	Sep 2021	Oct 2021	every 6 days
Surface Atmosphere Integrated Field Laboratory (SAIL) second Supplemental Facility	AMF	GUC S2	Nov 2021	Jun 2023	every 6 days
TRacking Aerosol Convection interactions ExpeRiment (TRACER) Main Site	AMF	HOU M1	Jun 2022	Sep 2022	daily
TRacking Aerosol Convection interactions ExpeRiment (TRACER) third Supplemental Facility	AMF	HOU S3	Jun 2022	Sep 2022	daily
Eastern Pacific Cloud Aerosol Precipitation Experiment (EPCAPE) Main Site	AMF	EPC M1	Feb 2023	Feb 2024	every 6 days
Cloud And Precipitation Experiment at kennaook (CAPE-k) third Supplemental Facility	AMF	KCG S3	Feb 2023	Oct 2025	every 6 days*
Coast-Urban-Rural Atmospheric Gradient Experiment (CoURAGE) Main Site	AMF	CRG M1	Dec 2024	Nov 2025	every 6 days
Coast-Urban-Rural Atmospheric Gradient Experiment (CoURAGE) second Supplemental Facility	AMF	CRG S2	Dec 2024	Nov 2025	every 6 days
CAPE-K-AEROSOLS	IOP	KCG S3	Feb 2025	Apr 2025	daily

*Filter durations vary due to the INS filter system operating only during clean sector "baseline" sampling periods. As such conditions were not always observed daily, individual 24-hour filter collections typically occur every ~6 days, but may be more or less than 6 days depending on site-specific conditions.

2.1.2 Mobile facility sites

Scientific users can propose field campaigns (<https://www.arm.gov/research/campaign-proposal>) to deploy one of ARM's three Mobile Facilities (AMFs) in undersampled regions around the world. These mobile platforms provide comprehensive atmospheric measurements, including INP filter sampling. Deployments for the first and second mobile facilities (AMF1 and AMF2, respectively) typically span 6–18 months, with the third mobile facility (AMF3) being deployed for up to 5–8 years. Information on ARM INP measurements made at the AMFs is also included in Figure 1 and Table 1.

Deleted: baseline

Deleted: sample

Deleted: durations

Deleted: span

Deleted: ess or moreshorter or longer

24 The first INP filters were collected as a part of the AMF3 at the Main Site in Oliktok Point, Alaska (OLI M1; 2 m AMSL,
25 70.495° N, 149.886° W), from August 2020 to June 2021. AMF3 was then relocated to the southeastern United States, where
26 filter collections began in October 2024 at the Main Site in Bankhead National Forest, Alabama (BNF M1; 293 m AMSL,
27 34.342° N, 87.338° W), and are ongoing.

28 INP filters were collected as a part of the AMF2 during the Surface Atmosphere Integrated Field Laboratory (SAIL; Feldman
29 et al., 2023) campaign in Crested Butte, Colorado. Sampling began at the Main Site (GUC M1; 2886 m AMSL, 38.956° N,
30 106.988° W) in September 2021 and continued through October 2021, before transitioning to the second Supplemental Facility
31 on Mt. Crested Butte (GUC S2; 3137 m AMSL, 38.898° N, 106.94° W), where collections continued for the duration of the
32 campaign from November 2021 to June 2023. AMF2 was subsequently deployed to Australia, where INP filters ~~were~~ collected
33 at the third Supplemental Facility during the Cloud And Precipitation Experiment at kennaook (CAPE-k) campaign, located
34 at the kennaook/Cape Grim Baseline Air Pollution Station on the northwestern tip of Tasmania (KCG S3; 67 m AMSL,
35 40.683° S, 144.690° E). This deployment began in February 2023 and ~~concluded~~ in October 2025. These samples ~~were~~
36 collected during clean sector or “baseline” conditions—when winds originated from the southwest, transporting air masses
37 across the Southern Ocean that ~~were~~ free from local point source contamination. However, select samples ~~were~~ also captured
38 air masses from over Tasmania to help characterize potential local influences. Baseline information indicating when sector-
39 based sampling was active is available through the ARM Data Discovery portal
40 (https://adc.arm.gov/discovery/#/results/instrument_code:baseline).

41 The first INP filters collected using AMF1 were obtained in Texas during the TRacking Aerosol Convection interactions
42 Experiment (TRACER) campaign (Jensen et al., 2023). Filters were collected at both the Main and third Supplemental Facility
43 sites in Houston (HOU M1: 8 m AMSL, 29.670° N, 95.059° W; HOU S3: 20 m AMSL, 29.328° N, 95.741° W) from June to
44 September 2022. The M1 site represented an urban environment, while the S3 site was rural. Due to the short duration of this
45 deployment, filters were collected approximately daily at both locations. Following TRACER, AMF1 was deployed to La
46 Jolla, California, as part of the Eastern Pacific Cloud Aerosol Precipitation Experiment (ECAPE; Russell et al., 2024), where
47 INP filters were collected at the Main Site (EPC M1; 7 m AMSL, 32.867° N, 117.257° W) from February 2023 to February
48 2024. AMF1 ~~resided~~ in Maryland for the Coast-Urban-Rural Atmospheric Gradient Experiment (CoURAGE), where filter
49 collection ~~occurred~~ at both the Main and second Supplemental Facility sites in the Baltimore region (CRG M1: 45 m AMSL,
50 39.317° N, 76.586° W; CRG S2: 158 m AMSL, 39.422° N, 77.21° W). This deployment began in December 2024 and
51 ~~continued~~ through November 2025. As with TRACER, the M1 and S2 sites represent urban and rural environments,
52 respectively.

53 A very recent IOP campaign, known as CAPE-K-AEROSOLS (CAPE-k Summertime Single-Particle and INP Campaign),
54 was conducted from February to April 2025. This campaign aimed to improve understanding and predictability of Southern

Deleted: are being

Deleted: is planned to

Deleted: are

Deleted: are

Deleted: have

Deleted: <https://adc.arm.gov/discovery/>

Deleted: currently

Deleted: s

Deleted: is ongoing

Deleted: is expected to

65 Ocean aerosol concentrations, chemical composition, and sources, as well as their relationships to CCN and INPs. During this
66 period, INP filters were collected approximately daily.

67 **2.1.3 Tethered balloon system (TBS) deployments**

68 ARM operates three TBSs, each capable of carrying payloads up to 50 kg on repeated vertical profiles through the atmospheric
69 boundary layer, reaching elevations of approximately 1500 m AMSL depending on meteorological conditions and regulatory
70 constraints. Detailed descriptions of the TBS systems are provided in Dexheimer et al. (2024). Vertically resolved INP filters
71 have been collected during several ARM TBS deployments through ARM field campaign requests, using a customized
72 miniaturized sampler. ~~The TBS INP sampler design, filter preparations, deployments, and available data are described in detail~~
73 ~~in Creamean et al. (2025) and are only briefly mentioned here.~~

74 **2.2 Filter preparation and sample collection**

75 **2.2.1 Fixed and AMF locations**

76 Filter units are prepared following the methodology outlined in Creamean et al. (2024), with a brief summary provided here.
77 Single-use Nalgene™ Sterile Analytical Filter Units are modified by replacing the original cellulose nitrate filters with 0.2-
78 µm polycarbonate filters, backed by either 10-µm polycarbonate filters (both 47 mm diameter Whatman® Nuclepore™ Track-
79 Etched Membranes) or 1-µm cellulose nitrate filters (47 mm diameter Whatman® non-sterile cellulose nitrate membranes),
80 depending on the anticipated aerosol loading at each site. All components are pre-cleaned in-house following the procedure
81 described in Barry et al. (2021). Filter units are disassembled and reassembled under ultraclean conditions inside a laminar
82 flow cabinet with near-zero ambient particle concentrations, then sealed and stored individually in clean airtight bags until
83 deployment.

84 Each sampling setup consists of the sterile, single-use filter units prepared at CSU, a totalizing mass flow meter (TSI Mass
85 Flow Meter 5200-1 or 5300-1, TSI Inc.), a vacuum pump (Oil-less Piston Compressor/Vacuum Pump, Thomas), connecting
86 tubing, and precipitation shields (Figure 2). Two identical filter assemblies operate in parallel: one collects primary filters for
87 INP analysis, while the other collects duplicate filters, which serve either as backups or as archival samples available for user-
88 requested analysis. The filter units are mounted open-faced and secured to the exterior of the AMF or other fixed-site
89 infrastructure, protected from precipitation by shield covers. Each unit is connected via vacuum line tubing to the flow meter
90 and vacuum pump, which are housed either within the main container or in an external pump enclosure, depending on the
91 available space and site-specific conditions.

92 Upon completion of sampling (typically after 24 hours) the 0.2-µm filters containing the collected aerosol particles are
93 carefully removed from the single-use filter units and stored frozen at approximately −20 °C in individual sterile Petri dishes
94 (Pall®). ~~As detailed in Table 1, the 24-hour samples are collected either daily or roughly every 6 days, depending on the goals~~

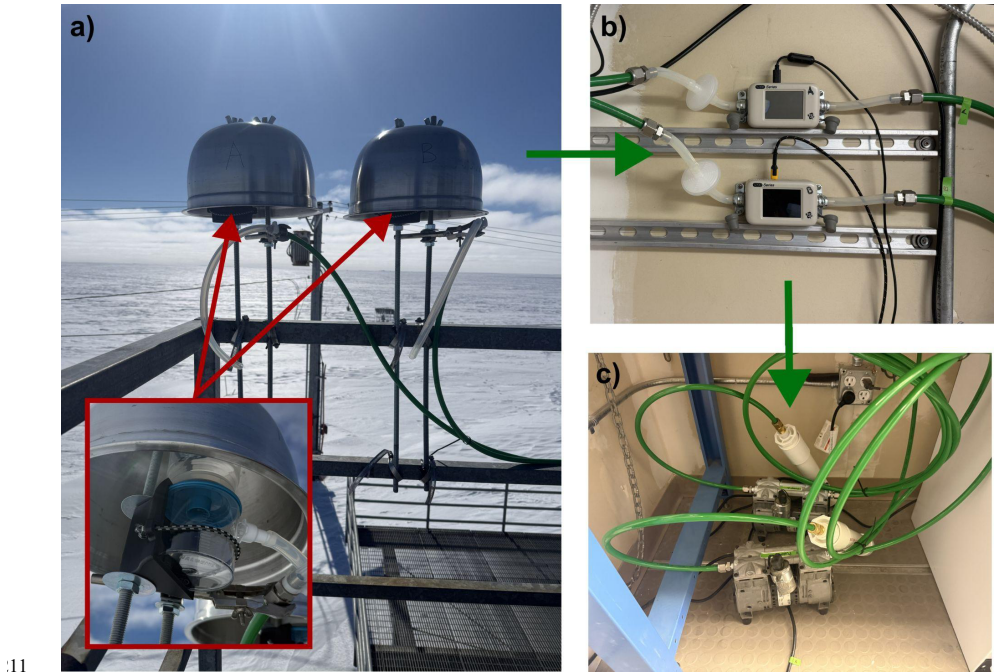
Deleted: Details of t

Deleted: and

Deleted: strategy, will be presented in a forthcoming publication. These past and near-future planned deployments include SGP in April 2022; GUC in May and July 2022, as well as January and April–June 2023; CRG in February and July 2025; and BNF in March, May/June, and August 2025. Deployment timelines and filter collection details are summarized in Table 2.

Deleted: *These samples have not yet been processed, and data are not currently available. Researchers interested in accessing or analyzing these samples may submit a request to ARM (<https://www.arm.gov/guidance/campaign-guidelines/small-campaigns>).

09 and duration of the measurement campaign. These samples are preserved on site until they can be transported in frozen batches
10 to CSU, where they remain frozen until they are processed and analysed.



11 **Figure 2:** Filter unit sampling apparatuses, including a) single-use, open-face filter units under precipitation shields which are
12 connected via tubing to b) the mass flow meters and to c) the vacuum pumps. Flow meters and pumps are always shielded from outside
13 conditions. The inset in a) shows a magnified photo of a filter unit in a custom, 3D-printed filter holder. All photos are from the NSA C1
14 site.
15

16 **2.3 Sample processing with the Ice Nucleation Spectrometer (INS)**

17 The INS mimics immersion freezing of cloud ice through ambient aerosols serving as INPs by way of heterogeneous ice
18 nucleation. This technique provides quantitative information on the population of ambient aerosols that can facilitate cloud ice
19 formation at a wide range of subzero temperatures and, hence, INP concentration (e.g., 6 orders of magnitude). The INS (also
20 known as the Colorado State University (CSU) Ice Spectrometer) is supported with well-established experimental protocols
21 and has been applied in many diverse scenarios (e.g., Beall et al., 2017; DeMott et al., 2017; Hill et al., 2016; Hiranuma et al.,
22 2015; McCluskey et al., 2017, 2018; Suski et al., 2018). It is an offline analytical instrument used to quantify freezing

Deleted: 2.2.2 Filter preparations for TBS deployments

TBS filters are identical in preparation to those used at fixed and mobile sites, 0.2- μm sample filters, but are housed differently to accommodate aerial and/or mobile deployment. Filters are loaded into reusable 47-mm polycarbonate in-line holders (Pall[®]), pre-cleaned with cycles of methanol and deionized water, and stored in foil and sterile plastic bags until use. These are deployed using a miniaturized sampler custom-built for mobile applications, currently operated on ARM's TBS platforms (Dexheimer et al., 2024). The TBS INP sampling system holds up to four in-line filter units, allowing for sampling across three altitude ranges plus one blank. Housed in a 3D-printed enclosure, the sampler is tethered to the TBS and remotely operated from the ground.

Deleted: —

Deleted: —

39 temperature spectra of immersion mode INP number concentrations from collected filter samples (Creamean et al., 2024).
40 Each INS unit consists of two 96-well aluminum incubation blocks originally designed for polymerase chain reaction (PCR)
41 plates, positioned end-to-end and thermally regulated by cold plates encasing the sides and base. Two INS instruments are
42 operated side-by-side to increase sample processing throughput. The temperature measurement range of the INS is between 0
43 °C and approximately −27 to −30 °C.

44 For analysis, each filter is placed in a sterile 50 mL polypropylene tube with 7–10 mL of 0.1 µm-filtered deionized water,
45 depending on expected aerosol loading. Lower volumes are used for cleaner environments to improve sensitivity. Samples are
46 re-suspended by rotating the tubes end-over-end for 20 minutes. Dilution series are prepared using the suspensions and 0.1 µm-
47 filtered deionized water, typically including 11-fold dilutions. Each suspension and its dilutions are dispensed into blocks of
48 32 aliquots (50 µL each) in single-use 96-well PCR trays (Optimum Ultra), alongside a 32-well negative control of filtered
49 deionized water. The trays are placed in the aluminum blocks of the INS and cooled at 0.33 °C min^{−1}. Freezing is detected
50 optically using a CCD camera with 1-second data resolution. HEPA-filtered N₂, pre-cooled near block temperature,
51 continuously purges the headspace to prevent condensation build-up and warming of the aliquots.

52 The time between collection and analysis has ranged from one week to over a year. Beall et al. (2020) reported no significant
53 differences in INP concentrations for samples stored between 1 and 166 days. In our internal quality checks, select samples
54 stored for 1–2.5 years at −20 °C showed minimal differences (<1%), while larger differences (20–60%) were observed only in
55 outlier cases associated with problematic original measurements.

56 2.3.1 Heat and peroxide treatments

57 Thermal and hydrogen peroxide (H₂O₂) treatments are used to probe INP composition, specifically targeting biologically-
58 derived materials (Maki et al., 1974). Heat treatment involves heating 2.5 mL of sample suspension to 95 °C for 20 minutes to
59 denature heat-labile INPs, such as proteins (Barry et al., 2023b, a; Hill et al., 2016, 2023; McCluskey et al., 2018b, c, a; Moore
60 et al., 2025; Suski et al., 2018; Testa et al., 2021). Peroxide digestion is performed on a separate 2 mL aliquot by adding 1 mL
61 of 30% H₂O₂ (Sigma-Aldrich®) to deionized water to create a 10% solution, followed by heating to 95 °C for 20 minutes
62 under UVB illumination to generate hydroxyl radicals. Residual H₂O₂ is then neutralized using catalase (MP Biomedicals™,
63 bovine liver). This process removes bio-organic INPs, as detailed in McCluskey et al. (2018c), Suski et al. (2018), and Testa
64 et al. (2021). H₂O₂ has long been used as an oxidizing agent for degrading organic matter, originating in soil science nearly a
65 century ago and later adopted across disciplines for removing organic material prior to chemical or physical analyses (McLean,
66 1931; Mikutta et al., 2005; Robinson, 1922; Schultz et al., 1999; Sequi and Aringhieri, 1977). In the presence of UV light,
67 H₂O₂ photolyzes to form highly reactive hydroxyl radicals that can oxidize and structurally modify organic macromolecules,
68 diminishing or eliminating their ice-nucleating activity (DeMott et al., 2023; Gute and Abbatt, 2020). Within the INP
69 community, H₂O₂ treatments typically range from 10 to 35% (Beall et al., 2022; O’Sullivan et al., 2014; Perkins et al., 2020;

Deleted: (DI)

Deleted: DI

Deleted: slightly above

Formatted: Subscript

Formatted: Subscript

Formatted: Subscript

Formatted: Subscript

Formatted: Subscript

Formatted: Subscript

Roy et al., 2021; Teska et al., 2024; Tobo et al., 2019). We conducted recent tests showing minimal differences in ice-nucleating activity between 10% and 20% treatments; however, further validation is needed, and future community efforts should aim to establish a standardized protocol and concentration to ensure methodological consistency across studies.

The differences in freezing spectra before and after each treatment provide insights into INP composition—yielding total, heat-labile (biological), bio-organic, and inorganic (often mineral) INP concentrations. However, it is important to note that wet heating may lead to a slight decrease in ice nucleation activity in select mineral types (Daily et al., 2022). Blanks are included during peroxide digestion to monitor potential contamination from reagents. Treatments are typically applied to one-third of samples from each location. Due to the ongoing nature of this program, the numbers of treatments conducted on samples from any given site evolve continuously on a weekly basis. Exact sample sites, dates, identifiers, and other metadata regarding which samples undergo treatments can be accessed in real time via the publicly available field log on the INS website. In the data files available on the ARM Data Discovery portal, treatments are indicated by a flag: 0 for base/untreated data, 1 for heat-treated data, and 2 for peroxide-treated data.

3 From raw data to final product: processing and quality control

3.1 INP concentration and uncertainty calculations

INP concentrations are calculated at each temperature interval using the fraction of frozen droplets and the known total volume of air filtered, following Equation (1) (Vali, 1971):

$$K(\theta) \text{ (L}^{-1}\text{)} = \frac{\ln(1-f)}{V_{\text{drop}}} \times \frac{V_{\text{suspension}}}{V_{\text{air}}} \quad (1)$$

where f is the proportion of frozen droplets, V_{drop} is the volume of each droplet, $V_{\text{suspension}}$ is the volume of the suspension, and V_{air} is the volume of air sampled (liters at standard temperature and pressure (STP) of 0 °C and 101.32 kPa). Specifically, the $V_{\text{suspension}}$ is the volume of 0.1 μm-filtered deionized water used to resuspend the particles from the filter (7–10 mL). The primary output of the INS is the freezing temperature spectrum of cumulative immersion mode INP number concentration, $K(\theta)$, from aerosols re-suspended from individual filters. INS output includes freezing temperature (°C), INP number concentration (L⁻¹ STP), 95% confidence intervals, and a treatment flag. Binomial confidence intervals are calculated following Agresti and Coull (1998), varying with the proportion of wells frozen. For example, freezing in 1 of 32 wells yields a confidence interval range of ~ approximately 0.2–5.0 times the estimated concentration, while 16 of 32 yields approximately 0.7–1.3 times the estimated concentration. The treatment flag denotes whether the suspension was base/untreated (total INPs; a flag of 0), heat-treated (biological INPs deactivated; a flag of 1), or H₂O₂-treated (organic INPs removed; a flag of 2). These values are derived from preliminary data files that include the processing date and time, freezing temperatures, and number of wells frozen (typically out of 32, each containing a 50 μL aliquot) per 0.5 °C interval.

Deleted: =

Formatted: Font: Italic

Formatted: Font: Italic, Subscript

03 Beyond Agresti and Coull confidence intervals, additional systematic and volumetric uncertainties associated with the INP
04 measurements include instrumental and procedural sources that contribute to the overall error budget. The flow meter accuracy
05 is $\pm 4\%$, based on the TSI 5200 Series Gas Flow Meter Operation and Service Manual. Type T thermocouples have an estimated
06 absolute accuracy of ± 0.5 °C, though the relative uncertainty between data points is closer to ± 0.1 °C (Perkins et al., 2020),
07 and no measurable block temperature gradients have been observed in prior laboratory tests. Uncertainties in droplet and
08 suspension volumes arise from pipetting and dispensing variability: $\pm 1.3\%$ for the larger pipette used in dilutions, $\pm 2.5\%$ for
09 the smaller pipette, $\pm 1.8\%$ for the multipipetter, and ± 0.1 – 0.8% for syringe-dispensed suspension volumes determined via
10 gravimetric testing at CSU. Additionally, edge cases in freezing data (0/32 and 32/32) are not reported, and limits of detection
11 (LOD) vary by sample based on total air and suspension volumes, with values below detection reported as -9999 .

12 **3.2 Quality control and assessment**

13 To ensure the reliability and robustness of immersion freezing data from the INS, we implement a comprehensive quality
14 control and assessment pipeline (Figure 3). This includes field sampling protocols, lab procedures, data validation, and
15 instrument maintenance.

16 **3.2.1 Field sampling quality control**

17 Filter samples collected for offline INS processing are carefully monitored during field deployment. At both the start and end
18 of each sampling period, the in-line pressure (kPa) and flow rate (standard liters per minute; Slpm) are recorded. These values
19 are evaluated for anomalies such as significant pressure or flow changes, which may indicate issues like leaks in the filter unit,
20 tubing, or system connections. To ensure accurate total air volumes are recorded, a totalizing mass flow meter logs flow every
21 second during sample collection. This meter is annually sent to the manufacturer for recalibration. Units that deviate by more
22 than 5% are returned to the manufacturer for servicing and recalibration. Field blanks are also prepared by briefly exposing
23 unused filters to ambient air at the sampling site. All sites undergo field blank collection approximately once per month, with
24 the exception of OLI M1, which is detailed in a Data Quality Report (DQR) that can be viewed under the “Description” for
25 this site’s INP data on the ARM Data Discovery Portal. Moving forward, all existing and new sites in the program will include
26 routine monthly field blanks.

Deleted: (TSI Inc. 5200-1, $\pm 2\%$ accuracy)

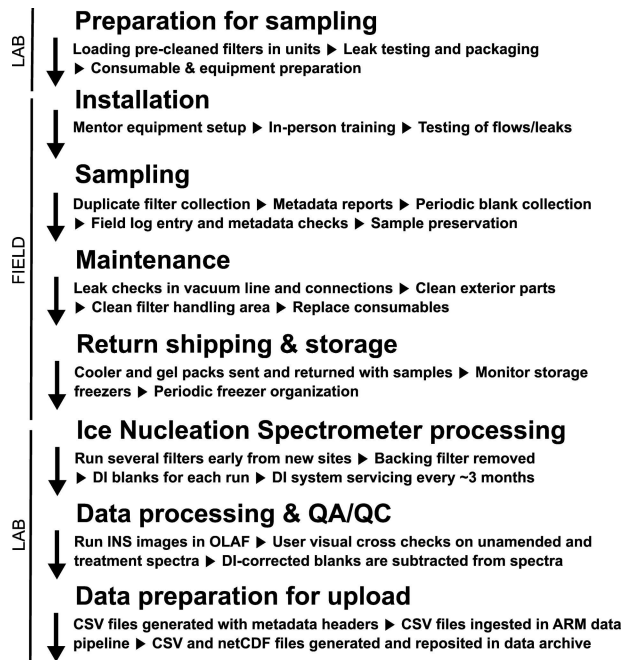


Figure 3: Flow diagram of quality assurance / quality control (QA/QC) protocols designed for DOE ARM INP data. Quality assurance ensures that data meet established standards for both ARM management and scientific end users, while quality control involves systematic inspection and testing to verify that performance characteristics align with predefined specifications. DI = deionized.

Routine maintenance for the field filter sampling system includes: (1) checking in-line temperature, pressure, and flow rate at the beginning and end of each sampling period, (2) inspecting precipitation shields and cleaning them as necessary, (3) ensuring single-use filter units are leak-free before deployment, (4) examining tubing and connection points for blockages or leaks, (5) verifying the performance of the vacuum pumps, which should sustain a 0.5 kPa vacuum, and (6) annual recalibration of the flow meters.

3.2.2 Laboratory protocols

To minimize contamination from lab surfaces or consumables (e.g., pipet tips, PCR plates, tubes), we follow a stringent sample preparation protocol (Barry et al., 2021). Pipets are calibrated annually, and a 0.1- μ m filtered deionized water blank is included with each INS run to correct for background INPs introduced during re-suspension or by the trays themselves. For peroxide

41 digestion experiments, blanks with deionized water are included to detect potential contamination from H₂O₂ or catalase
42 reagents. These are prepared using the same procedures as the actual samples to assess background INP levels and serve as a
43 quality control check to determine whether reprocessing is necessary.

44 **3.2.3 Instrument quality control and calibration**

45 INS temperature accuracy is critical and maintained within ± 0.2 °C, accounting for thermocouple uncertainty and ensuring no
46 block temperature gradients develop over time. Each PCR block contains one thermocouple inserted just below the wells, and
47 for each pair of blocks, the thermocouple readings are averaged. HEPA-filtered N₂ used to purge the PCR tray headspace is
48 pre-cooled to prevent condensation build-up on plexiglass lids and warming the 50 μ L aliquots during measurement. Camera
49 images are captured every 20 seconds (approximately every 0.1 °C) during analysis to verify automated freezing detection.
50 Each INS run is manually cross-checked against the recorded images to ensure proper identification of frozen wells. The
51 deionized water blanks run with every sample serve a dual purpose: they act as a positive control as well and help monitor
52 instrument drift, potential contamination, and the proper functioning of INS components (e.g., thermocouples, cameras).
53 Routine lab maintenance of the INS includes: (1) cleaning plexiglass lids biweekly with Windex and deionized water, (2)
54 monthly deep cleaning of the lab space, (3) monitoring copper piping for leaks of SYLTHERM™ XLT heat transfer fluid, and
55 (4) watching the nitrogen tank depletion rate to detect leaks. We have confirmed the repeatability and reliability of the INS
56 technique through replicate filter testing and campaign comparisons. Additionally, replicate filters have been analyzed to
57 ensure comparability (Creamean et al., 2024).

58 **3.3 Automated data processing algorithm**

59 Historically, data produced by the INS have been analyzed manually using Microsoft Excel. In 2024, a data scientist was hired
60 to develop the Open-source Library for Automating Freezing Data acQuisition from Ice Nucleation Spectrometer (OLAF DaQ
61 INS), which now has its Version 1 completed. More information and the software itself are available at:
62 <https://github.com/SiGran/OLAF>. Briefly, the OLAF DaQ INS software provides a graphical user interface that allows users
63 to manually cross-check camera images taken during each INS run against the recorded well freezing data. Once image
64 verification is complete, the program generates a CSV file with freezing data at every 0.5 °C interval, including the first
65 instance of observed freezing to the nearest 0.1 °C. PCR wells containing deionized water are automatically subtracted from
66 the sample wells for both the neat and serially diluted suspensions. These deionized water-corrected well freezing data are then
67 converted to INP concentrations (per liter of air at STP) at each temperature bin using Equation (1). Binomial confidence
68 intervals are calculated following Agresti and Coull (1998) and also converted to INP L⁻¹ using the same equation. For each
69 temperature bin, the program selects the INP concentration from the least dilute sample that remains statistically valid. When
70 a dilution reaches its statistically significant limit, the program moves to the next most dilute sample.

Deleted: is

Deleted: nearing completion

Deleted: can be found

Deleted: DI

Deleted: DI-

76 In cases where INP concentrations decrease with decreasing temperature (an artifact sometimes introduced by the stochastic
77 nature of serial dilution measurements) the program automatically adjusts the values to enforce monotonicity. We are currently
78 developing an additional QC flag for OLAF-generated data files to indicate which data points were adjusted due to
79 monotonicity-related corrections. This correction was not applied prior to OLAF when files were generated manually. Because
80 blank subtraction can also produce this artifact, the correction is applied after the blank subtraction step. Specifically, if a
81 blank-corrected value falls below the lower 95% confidence bound of the uncorrected value, the program replaces it with the
82 previous bin's value and propagates the upper confidence interval using the root mean square of the current and previous
83 intervals. This correction is applied only when the number of affected bins remains below a user-defined threshold (10% of
84 total temperature bins per sample); if exceeded, those bins are flagged with an error signal (−9999). At ground-based terrestrial
85 sites, corrections are almost entirely due to dilution stochasticity and rarely result from blank subtraction, whereas marine or
86 other low-aerosol loading environments tend to experience a higher frequency of corrections related to blank subtraction.
87 Finally, the software compiles all blank-corrected data across treatments (base/untreated, heat, and peroxide) into a single
88 output file, including treatment flags for each sample.

89 **3.4 Ingesting processed INP data into ARM Data Discovery**

90 The final step in making INS-derived INP data publicly available is ingestion into the ARM Data Discovery portal. This begins
91 with the CSV files generated during INS processing, which are passed through an automated pipeline that standardizes them
92 into a universal format used across all ARM datasets. This format includes all necessary metadata headers and timestamps.
93 During ingestion, the ARM Data Quality Office (DQO) evaluates the data by reviewing plots and statistical metrics of the INP
94 data. If any issues are identified, the DQO works with the mentor team to resolve them. This dual-level review, by both
95 scientific mentors and the DQO, ensures the robustness and reliability of the final data products. Once approved, the data are
96 published at the “a1” level, which denotes that calibration factors have been applied, values have been converted to geophysical
97 units, and the dataset is considered final. These files are available in NetCDF and/or ASCII-CSV formats and can be accessed
98 by placing a data order through the ARM Data Discovery portal. A free ARM account is required to request and download the
99 data.

00 **4 Applications of ARM INP data**

01 **4.1 Temporal trends in INP concentrations from long-term monitoring**

02 As the first established fixed site, SGP C1 hosts nearly five consecutive years of INP concentration data (Figure 4). Untreated
03 (i.e., base, or total INP) measurements, collected approximately every six days, are publicly available. Long-term datasets such
04 as this are invaluable for examining the annual cycle of INPs in detail. For instance, Figure 4 reveals a pronounced seasonal
05 pattern, with INP concentrations peaking during the fall/winter months (October–January), particularly at warmer freezing
06 temperatures (e.g., > −10 °C). At colder temperatures (e.g., ≤ −15 °C), the seasonal cycle is less distinct. Although INPs active

Deleted: ¶

Deleted: , such as field notes, contact information for the INP mentor team, time stamps, and details on sample processing...

Deleted: Unamended

Deleted: untreated

at the warmest temperatures (≥ -6 °C) were relatively rare, the few observed events tended to coincide with the fall/winter peak. This site is influenced by surrounding agricultural activities, which may contribute to the observed seasonal variability in INPs; however, a comprehensive source attribution is beyond the scope of this manuscript. Our intent here is to highlight the completeness and continuity of the SGP dataset and its utility. These measurements support both observational studies of INP variability and source characterization, and model evaluation efforts such as Knopf et al. (2021).

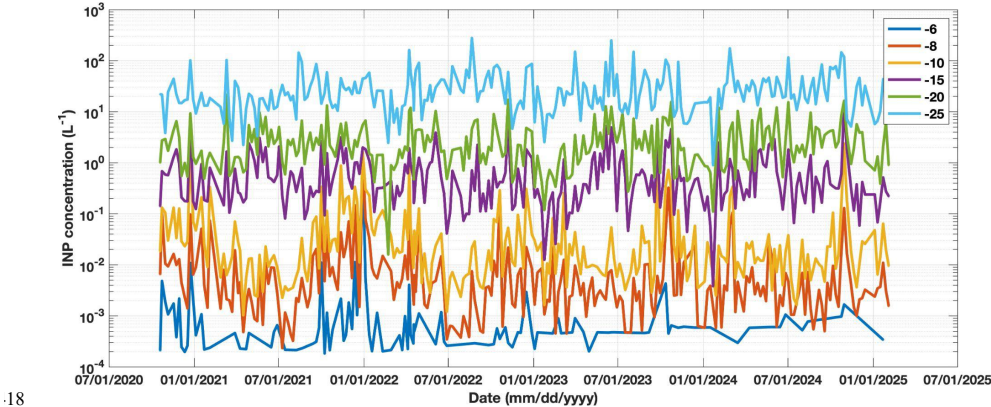


Figure 4: Complete time series of INP concentrations at select temperatures from the SGP C1 site that are currently publicly available on DOE ARM Data Discovery. Each line shows cumulative INP concentrations per liter of air (L^{-1}) at freezing temperatures designated in the legend (in °C). [Data are from 247 total processed filter samples.](#)

4.2 Characterizing INP types through heat and peroxide treatments

In addition to the time series of total INP concentrations, approximately one-third of the samples undergo specific heat and peroxide treatments to help identify broad classes of INP types. These treatments target: (1) heat-labile INPs, such as proteins commonly associated with biological particles; (2) heat-stable organics, isolated via hydrogen peroxide treatment; and (3) the remaining, largely inorganic fraction, which is often attributed to mineral dust (Barry et al., 2023a, 2025; Creamean et al., 2020; DeMott et al., 2025a; Hill et al., 2016; McCluskey et al., 2018c; Schiebel et al., 2016; Suski et al., 2018; Testa et al., 2021; Tobo et al., 2019). Figure 5 provides an example of the relative contributions of these INP types over time at SGP C1, shown as a percentage of total INPs at two temperatures. The fraction of “biological” INPs is derived by subtracting the heat-treated INP spectrum from the untreated spectrum. The “organic” component is isolated by subtracting the peroxide-treated spectrum from the heat-treated spectrum. The residual “inorganic” fraction is estimated by subtracting the peroxide-treated spectrum from the untreated spectrum.

Deleted: unamended

Deleted: unamended

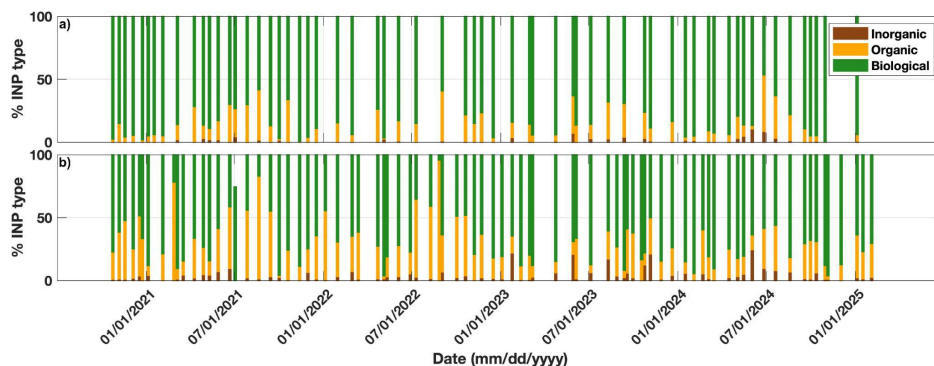


Figure 5: Relative abundance of INP type at the SGP C1 site at a) -15°C and b) -25°C that are currently available on DOE ARM Data Discovery. INP types are determined through heat and peroxide treatments. We assume that the reduction of INPs from heat are biological in nature (e.g., heat labile proteins) while the reduction of INPs from peroxide, UV, and heat are organic (e.g., heat labile organics). INPs remaining (unaffected) by both treatments are inorganic (e.g., mineral dust). Data are from 84 samples that have been heat- and peroxide-treated (34% of the processed SGP samples in Figure 4).

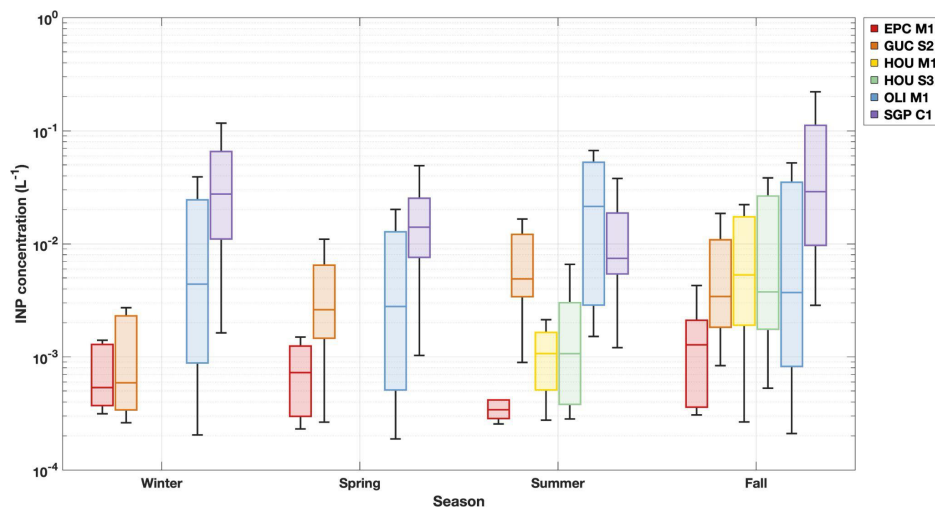
These unique long-term data offer insights into the seasonal variability and relative importance of different INP sources. For instance, at -15°C , biological INPs dominate at SGP, with smaller contributions from organics and inorganics. The inorganic component becomes more apparent during the summer months, likely associated with dry, dusty conditions on agricultural lands (Evans, 2025; Ginoux et al., 2012). At -25°C , the relative contributions of organic and inorganic INPs increase, yet biological INPs still remain the dominant type overall. Although the Great Plains region is periodically influenced by dust events, its agricultural soils are rich in biological material (Delgado-Baquerizo et al., 2018; Garcia et al., 2012; Hill et al., 2016; Kanji et al., 2017; O’Sullivan et al., 2014; Pereira et al., 2022; Steinke et al., 2016; Suski et al., 2018; Tobo et al., 2014), which distinguishes it from more arid, desert regions where mineral dust may dominate. These compositional insights are particularly valuable for users interested not only in INP abundance but also in potential sources. The treatment data can be used in combination with aerosol composition and meteorological observations at SGP C1 (and other ARM sites), and air mass trajectory analysis to further constrain the origins of INPs.

4.3 Seasonal INP variability across sites

INP data can be meaningfully compared across a diverse range of sites throughout the year, as illustrated in Figures 6 and 7 for -10°C and -20°C , respectively. The purpose of these figures is to highlight the diversity of INP concentrations across a range of environments and to demonstrate the value of consistent INP measurements at multiple sites. Each site shown represents a distinct setting: EPC M1 is a coastal marine site in California; GUC S2 is located at high elevation in the Colorado Rocky Mountains; the HOU sites include both urban and rural environments in Texas; OLI S3 is situated in a coastal oilfield region of northern Alaska within the Arctic; and SGP represents a high plains agricultural site in the central U.S. These are the

Deleted: ,

60 sites for which data are currently available through ARM Data Discovery, with additional datasets forthcoming for sites in
61 Tasmania, northern Alaska, and the northeastern and southeastern United States.



62
63 **Figure 6: Seasonal INP concentrations at -10 °C at all fixed and mobile facility sites currently available from the DOE ARM Data**
64 **Discovery.** Data are presented in box-and-whisker format, with the middle line being the median (50th percentile), box edges representing
65 the 25th and 75th percentiles, and the whiskers representing data within 1.5× the interquartile range. The numbers above each median line
66 indicate the number of data points that went into each bar.

67 Several noteworthy patterns emerge from these intercomparisons. At -10 °C, where INPs are likely dominated by biological
68 materials (Huang et al., 2021; Kanji et al., 2017), many sites exhibit clear seasonal cycles, though the timing and magnitude
69 of these cycles differ. For instance, SGP shows elevated INP concentrations in the winter and fall, consistent with agricultural
70 activity and associated emissions during that time. In contrast, GUC exhibits higher concentrations in summer, which aligns
71 with the seasonal exposure of vegetation and the wintertime snow cover typical of the Colorado Rocky Mountains. Similarly,
72 the Arctic coastal site OLI displays peak concentrations in summer, even exceeding those at the midlatitude SGP site. This is
73 consistent with findings highlighting the biological productivity of Alaskan Arctic waters and tundra in May through
74 September leading to increased airborne INPs (Barry et al., 2025; Creamean et al., 2018a, 2019; Eufemio et al., 2023; Fountain
75 and Ohtake, 1985; Nieto-Caballero et al., 2025; Perring et al., 2023; Rogers et al., 2001; Wex et al., 2019), despite the presence
76 of extensive oil and gas infrastructure near OLI that impacts the aerosol composition (Creamean et al., 2018b; Gunsch et al.,
77 2017). However, a few important considerations should be noted. Field blanks were not collected at OLI; instead, a laboratory
78 blank was used to subtract background INPs. This approach may lead to artificially elevated concentrations, as lab blanks

Deleted: —

typically have lower background levels than field blanks due to reduced handling and exposure. Additionally, the OLI data represent a single summer season, whereas the SGP data span four summers. If the OLI summer was anomalous, this could skew comparisons. These factors should be carefully considered when interpreting or using the OLI dataset.

Deleted: This limitation is noted in the Data Discovery metadata for the OLI dataset.

Conversely, EPC recorded the lowest INP concentrations among the sites, likely due to its exposure to clean marine air masses, which are generally associated with INP levels lower than terrestrial environments (e.g., DeMott et al., 2016; McCluskey et al., 2018b; Welti et al., 2020). Interestingly, both the urban and rural sites in HOU exhibited similar INP concentrations during the summer and fall, despite the common assumption that urban emissions are generally poor sources of INPs (Bi et al., 2019; Cabrera-Segoviano et al., 2022; Chen et al., 2018; Hasenkopf et al., 2016; Ren et al., 2023; Schrod et al., 2020; Tobo et al., 2020; Wagh et al., 2021; Yadav et al., 2019; Zhang et al., 2022; Zhao et al., 2019). The results from OLI and HOU collectively suggest that nearby regional marine sources can substantially influence INP concentrations, even in regions characterized by high levels of industrialization or urbanization.

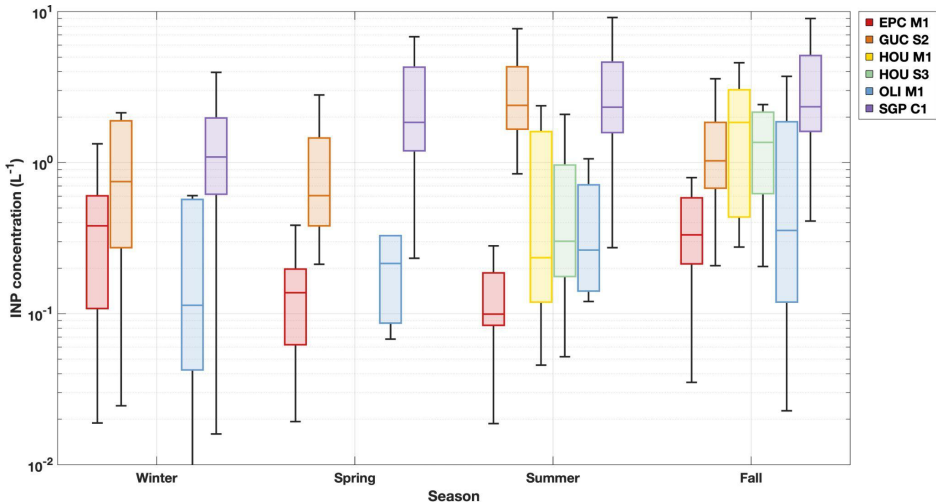


Figure 7: Same as Figure 6, but for seasonal INP concentrations at -20°C at all fixed and mobile facility sites currently available from the DOE ARM Data Discovery. Note the scale of the INP concentration axis is higher than Figure 6.

At -20°C , seasonal patterns in INP concentrations remain evident across most sites, but notable differences emerge compared to the -10°C data. INP concentrations at the two HOU sites remain comparable, consistent with the pattern observed at warmer temperatures. However, one of the most striking differences is that OLI, which had among the highest concentrations at $-$

10 °C, no longer stands out; instead, it shows significantly lower INP levels than SGP. This shift suggests that SGP may have a more prominent source of mineral dust or cold-temperature-active organic INPs than the Arctic coastal OLI site. This interpretation is consistent with known regional differences, as the U.S. midlatitudes, including the central plains where SGP is located, coexist with more prominent dust emissions compared to the North American Arctic (e.g., Ginoux et al., 2012; Rodriguez-Caballero et al., 2022; Song et al., 2021). Interestingly, INP concentrations at OLI are now more comparable to those at EPC, likely reflecting the marine influence at both locations, which generally has lower INP concentrations relative to continental sources.

These INP measurements are consistent with many principal investigator-led datasets collected at other ARM-supported locations, such as those that employ the Colorado State University Ice Spectrometer (see Table 3). The INS that is used to produce the ARM INP data is almost identical to the Ice Spectrometer. This opens opportunities for broader comparisons to campaigns such as the 2017–2018 MARCUS (Measurements of Aerosols, Radiation, and Clouds over the Southern Ocean; DeMott et al., 2018b; McCluskey et al., 2018c; McFarquhar et al., 2019, 2021; Niu et al., 2024; Raman et al., 2023) and 2016–2018 MICRE (Macquarie Island Cloud and Radiation Experiment; DeMott et al., 2018a; Marchand, 2020; McCluskey et al., 2023; Niu et al., 2024; Raman et al., 2023) campaigns in the Southern Ocean, 2019–2020 MOSAiC (Multidisciplinary drifting Observatory for the Study of Arctic Climate; Barry et al., 2025; Creamean et al., 2022; Shupe et al., 2021, 2022) campaign in the Arctic Ocean, 2019–2020 COMBLE (Cold-Air Outbreaks in the Marine Boundary Layer Experiment; DeMott and Hill, 2021; DeMott et al., 2025b; Geerts et al., 2021, 2022) campaign along the Norwegian Arctic coast, 2018–2019 CACTI (Cloud, Aerosol, and Complex Terrain Interactions; DeMott and Hill, 2020; Testa et al., 2021; Varble et al., 2019) campaign in agricultural regions of South America, the 2019 AEROICESTUDY (Aerosol-Ice Formation Closure Pilot Study; Knopf et al., 2020, 2021) and 2014 INCE (Ice Nuclei Characterization Experiment; DeMott et al., 2015) at SGP, and the 2015 ACAPEX (ARM Cloud Aerosol Precipitation Experiment; DeMott and Hill, 2016; Fan et al., 2014; Leung, 2016; Levin et al., 2019; Lin et al., 2022) study off the coast of California. These complementary datasets are also publicly available through ARM Data Discovery, but labeled as “icespec” (or “icespec-air” for aircraft measurements).

Table 3. List of previous PI-led DOE ARM field campaigns with comparable INP data to the INS. Includes measurement location, start and end dates, filter collection details, and DOI for the INP measurements. Data from earlier studies do not have available DOIs. Note all of these campaigns are AMF deployments. RV is abbreviated for Research Vessel.

Field campaign name	Location	INP filter start	INP filter end	Filter collection details	DOI (https://doi.org/)
Measurements of Aerosols, Radiation, and Clouds over the Southern Ocean (MARCUS)	Southern Ocean on the <i>Aurora Australis</i>	Oct 2017	Apr 2018	continuous; 24- to 48-h samples	10.5439/1638968
Macquarie Island Cloud and	Macquarie Island,	Mar	Mar 2018	continuous; 48- to	10.5439/1638330

Radiation Experiment (MICRE)	Australia	2016		72-h samples	
Multidisciplinary Drifting Observatory for the Study of Arctic Climate (MOSAIC)	Arctic Ocean on the <i>RV Polarstern</i>	Oct 2019	Oct 2020	continuous; 72-h samples	10.5439/1804484
Cold-Air Outbreaks in the Marine Boundary Layer Experiment (COMBLE)	Andenes, Norway	Dec 2019	Mar 2020	during CAOs; 6- to 74-h samples	10.5439/1755091
Cloud, Aerosol, and Complex Terrain Interactions (CACTI)	Villa Yacanto, Argentina	Oct 2018	Apr 2019	quasi-continuous; 8-h samples	10.5439/1607786
Cloud, Aerosol, and Complex Terrain Interactions (CACTI)	Sierras de Córdoba, Argentina	Nov 2018	Dec 2018	flight duration; various sample durations	10.5439/1607793
Aerosol-Ice Formation Closure Pilot Study (AEROICESTUDY)	SGP	Oct 2019	Oct 2019	continuous; 12- to 24-h samples	10.5439/1637710
Ice Nuclei Characterization Experiment (INCE)	SGP	Apr 2014	Jun 2014	continuous; 24-h samples	none
ARM Cloud Aerosol Precipitation Experiment (ACAPEX)	Pacific Ocean on the ARM G-1 aircraft	Jan 2015	Mar 2015	flight duration; 10-min to 3-h samples	none

Deleted: the

Deleted: access to

5 Community use and ~~limitations of~~ ARM INP data

We present a comprehensive dataset of immersion mode INP concentrations from multiple sites across the United States and beyond. Most of these data are publicly available through the DOE ARM Data Discovery portal (<https://adc.arm.gov/discovery/>). On the portal, data from fixed sites and AMF deployments can be found by searching for “INP,” while data collected via ARM tethered balloon systems can be found by searching for “TBSINP.” DOIs for INP and TBSINP are <https://doi.org/10.5439/1770816> and <https://doi.org/10.5439/2001041>, respectively. For sites with ongoing measurements, data are routinely uploaded as batches of samples are processed using the INS. Upcoming INP datasets from the CAPE-k (KCG S3), CoURAGE (CRG M1 and S2), BNF (M1), and NSA (C1) sites will also be made available in the near future. These ARM-based INP measurements are directly comparable to other principal investigator-led datasets collected in previous studies at a wider range of locations, allowing for meaningful cross-site comparisons.

Importantly, duplicate filters are collected at most sites and preserved frozen for potential future analyses. Researchers interested in obtaining additional INP data on unprocessed samples or conducting their own supplementary aerosol physicochemical analyses can request these archived samples by submitting an ARM Small Campaign Request

42 (<https://www.arm.gov/guidance/campaign-guidelines/small-campaigns>) with the option to contact the ARM INP mentor team
43 (co-authors on this manuscript) with questions. At many of the sites listed in Table 1, only a subset of collected filters has been
44 processed to date. Therefore, users with specific dates or time periods of interest are encouraged to reach out to the mentor
45 team to request new analyses, including specialized treatments. A detailed filter collection log is available on the ARM INS
46 homepage (<https://www.arm.gov/capabilities/instruments/ins>) to help guide these inquiries. INP data from future campaigns
47 requested by researchers will also be made accessible to the broader research community.

48
49 The DOE ARM baseline INP measurements provide valuable long-term and IOP-based observations but have several
50 limitations that users should be aware of. First, these measurements do not account for time dependence in freezing behavior,
51 which is generally less significant than temperature dependence (Ervens and Feingold, 2013). Second, sampling assumes
52 collection of the total aerosol size distribution; however, this has not been explicitly tested, so the exact size range collected is
53 uncertain. The 0.2- μ m filters we use have reduced transmission efficiency for particles around 150 nm (down to 65–78%), but
54 generally exhibit very high collection efficiency at most sizes (Spurny and Lodge, 1972). Third, because filters are collected
55 over 24 hours, typically every six days, short-term or episodic INP events may be missed, although higher-frequency sampling
56 can be requested. Fourth, these measurements are made at the surface and may not fully represent the INP population at cloud
57 level, though cloud-surface coupling analyses (e.g., Creamean et al., 2021; Griesche et al., 2021) and TBS INP data (Creamean
58 et al., 2025) can help bridge this gap. Lastly, not all samples are subjected to treatments unless requested, and, as noted by
59 Burrows et al. (2022), the absence of co-located baseline measurements of aerosol composition (particularly dust, sea spray,
60 and primary biological aerosol particles) limits the ability to fully constrain INP sources and improve parameterizations in
61 models.

62 Data availability

63 INP and TBSINP data are available from the DOE ARM Data Discovery portal (<https://adc.arm.gov/discovery/>) under DOIs
64 <https://doi.org/10.5439/1770816> (Creamean et al., 2024) and <https://doi.org/10.5439/2001041> (Creamean et al., 2025),
65 respectively.

66 Author contributions

67 JMC and AT conceptualised the INP mentor program. CCH and MV conducted the sample and data analysis for the INP data
68 that are publicly available for download from the DOE ARM Data Center. CCH and JMC were additionally responsible for
69 instrument installation and maintenance at the sites. All authors contributed to the writing of this manuscript.

Deleted: and TBS deployments

Deleted: s

Deleted: and 2

73 **Competing interests**

74 None of the authors has any competing interests

75 **Disclaimer**

76 Publisher's note: Copernicus Publications remains neutral with regard to jurisdictional claims made in the text, published maps,
77 institutional affiliations, or any other geographical representation in this paper. While Copernicus Publications makes every
78 effort to include appropriate place names, the final responsibility lies with the authors.

79 **Acknowledgements**

80 This work was supported by the Office of Biological and Environmental Research within the U.S. Department of Energy
81 (DOE) through the Atmospheric Radiation Measurement (ARM) user facility. JMC, CCH, and MV received support under
82 DOE contract no. DE-0F-60173. We gratefully acknowledge James Mather for his invaluable support in the development and
83 implementation of the INP program. We also extend our sincere thanks to the ARM site staff for their significant assistance
84 with instrument installation, sample collection, and logistics. We gratefully acknowledge Thomas C. J. Hill for his foundational
85 role as co-mentor alongside JMC during the inception of this program, and for his enduring guidance and expertise. He is now
86 enjoying a well-earned retirement in Australia. ChatGPT was used to assist in editing and improving the wording of this
87 manuscript.

88 **Financial support**

89 This research has been supported by Argonne National Laboratory for the DOE under contract DE-0F-60173.

90 **References**

- 91 Agresti, A. and Coull, B. A.: Approximate Is Better than “Exact” for Interval Estimation of Binomial Proportions, *Am. Stat.*,
92 52, 119, <https://doi.org/10.2307/2685469>, 1998.
- 93 Barry, K. R., Hill, T. C. J., Jentsch, C., Moffett, B. F., Stratmann, F., and DeMott, P. J.: Pragmatic protocols for working
94 cleanly when measuring ice nucleating particles, *Atmospheric Res.*, 250, 105419,
95 <https://doi.org/10.1016/j.atmosres.2020.105419>, 2021.
- 96 Barry, K. R., Hill, T. C. J., Nieto-Caballero, M., Douglas, T. A., Kreidenweis, S. M., DeMott, P. J., and Creamean, J. M.:
97 Active thermokarst regions contain rich sources of ice-nucleating particles, *Atmospheric Chem. Phys.*, 23, 15783–15793,
98 <https://doi.org/10.5194/acp-23-15783-2023>, 2023a.
- 99 Barry, K. R., Hill, T. C. J., Moore, K. A., Douglas, T. A., Kreidenweis, S. M., DeMott, P. J., and Creamean, J. M.: Persistence

and Potential Atmospheric Ramifications of Ice-Nucleating Particles Released from Thawing Permafrost, *Environ. Sci. Technol.*, 57, 3505–3515, <https://doi.org/10.1021/acs.est.2c06530>, 2023b.

Barry, K. R., Hill, T. C. J., Kreidenweis, S., DeMott, P. J., Tobo, Y., and Creamean, J. M.: Bioaerosols as indicators of central Arctic ice nucleating particle sources, *Atmos Chem Phys Lett*, 2025.

Beall, C. M., Lucero, D., Hill, T. C., DeMott, P. J., Stokes, M. D., and Prather, K. A.: Best practices for precipitation sample storage for offline studies of ice nucleation in marine and coastal environments, *Atmospheric Meas. Tech.*, 13, 6473–6486, <https://doi.org/10.5194/amt-13-6473-2020>, 2020.

Beall, C. M., Hill, T. C. J., DeMott, P. J., Köneman, T., Pikridas, M., Drewnick, F., Harder, H., Pöhlker, C., Lelieveld, J., Weber, B., Iakovides, M., Prokeš, R., Sciare, J., Andreae, M. O., Stokes, M. D., and Prather, K. A.: Ice-nucleating particles near two major dust source regions, *Atmospheric Chem. Phys.*, 22, 12607–12627, <https://doi.org/10.5194/acp-22-12607-2022>, 2022.

Bi, K., McMeeking, G. R., Ding, D. P., Levin, E. J. T., DeMott, P. J., Zhao, D. L., Wang, F., Liu, Q., Tian, P., Ma, X. C., Chen, Y. B., Huang, M. Y., Zhang, H. L., Gordon, T. D., and Chen, P.: Measurements of Ice Nucleating Particles in Beijing, China, *J. Geophys. Res. Atmospheres*, 124, 8065–8075, <https://doi.org/10.1029/2019JD030609>, 2019.

Burrows, S.: Agricultural Ice Nuclei at Southern Great Plains Supplemental Sampling (AGINSGP-SUPP) Field Campaign Report, 2023.

Burrows, S. M., McCluskey, C. S., Cornwell, G., Steinke, I., Zhang, K., Zhao, B., Zawadowicz, M., Raman, A., Kulkarni, G., Chen, S., Zelenyuk, A., and DeMott, P. J.: Ice-Nucleating Particles That Impact Clouds and Climate: Observational and Modeling Research Needs, *Rev. Geophys.*, 60, e2021RG000745, <https://doi.org/10.1029/2021RG000745>, 2022.

Cabrera-Segoviano, D., Pereira, D. L., Rodriguez, C., Raga, G. B., Miranda, J., Alvarez-Ospina, H., and Ladino, L. A.: Inter-annual variability of ice nucleating particles in Mexico city, *Atmos. Environ.*, 273, 118964, <https://doi.org/10.1016/j.atmosenv.2022.118964>, 2022.

Chen, J., Wu, Z., Augustin-Bauditz, S., Grawe, S., Hartmann, M., Pei, X., Liu, Z., Ji, D., and Wex, H.: Ice-nucleating particle concentrations unaffected by urban air pollution in Beijing, China, *Atmospheric Chem. Phys.*, 18, 3523–3539, <https://doi.org/10.5194/acp-18-3523-2018>, 2018.

Conen, F., Morris, C. E., Leifeld, J., Yakutin, M. V., and Alewell, C.: Biological residues define the ice nucleation properties of soil dust, *Atmospheric Chem. Phys.*, 11, 9643–9648, <https://doi.org/10.5194/acp-11-9643-2011>, 2011.

Creamean, J., Hill, T., Hume, C., and Devados, T.: Ice Nucleation Spectrometer (INS) Instrument Handbook, U.S. Department of Energy, Atmospheric Radiation Measurement user facility, Richland, Washington, 2024.

Creamean, J. M., Suski, K. J., Rosenfeld, D., Cazorla, A., DeMott, P. J., Sullivan, R. C., White, A. B., Ralph, F. M., Minnis, P., Comstock, J. M., Tomlinson, J. M., and Prather, K. A.: Dust and Biological Aerosols from the Sahara and Asia Influence Precipitation in the Western U.S., *Science*, 339, 1572–1578, <https://doi.org/10.1126/science.1227279>, 2013.

Creamean, J. M., Kirpes, R. M., Pratt, K. A., Spada, N. J., Maahn, M., de Boer, G., Schnell, R. C., and China, S.: Marine and terrestrial influences on ice nucleating particles during continuous springtime measurements in an Arctic oilfield location, *Atmospheric Chem. Phys.*, 18, 18023–18042, <https://doi.org/10.5194/acp-18-18023-2018>, 2018a.

Creamean, J. M., Maahn, M., de Boer, G., McComiskey, A., Sedlacek, A. J., and Feng, Y.: The influence of local oil exploration and regional wildfires on summer 2015 aerosol over the North Slope of Alaska, *Atmospheric Chem. Phys.*, 18,

555–570, <https://doi.org/10.5194/acp-18-555-2018>, 2018b.

Creamean, J. M., Cross, J. N., Pickart, R., McRaven, L., Lin, P., Pacini, A., Hanlon, R., Schmale, D. G., Ceniceros, J., Aydele, T., Colombi, N., Bolger, E., and DeMott, P. J.: Ice nucleating particles carried from below a phytoplankton bloom to the Arctic atmosphere, *Geophys. Res. Lett.*, 46, 8572–8581, <https://doi.org/10.1029/2019GL083039>, 2019.

Creamean, J. M., Hill, T. C. J., DeMott, P. J., Uetake, J., Kreidenweis, S., and Douglas, T. A.: Thawing permafrost: an overlooked source of seeds for Arctic cloud formation, *Environ. Res. Lett.*, 15, 084022, <https://doi.org/10.1088/1748-9326/ab87d3>, 2020.

Creamean, J. M., de Boer, G., Telg, H., Mei, F., Dexheimer, D., Shupe, M. D., Solomon, A., and McComiskey, A.: Assessing the vertical structure of Arctic aerosols using balloon-borne measurements, *Atmospheric Chem. Phys.*, 21, 1737–1757, <https://doi.org/10.5194/acp-21-1737-2021>, 2021.

Creamean, J. M., Barry, K., Hill, T. C. J., Hume, C., DeMott, P. J., Shupe, M. D., Dahlke, S., Willmes, S., Schmale, J., Beck, I., Hoppe, C. J. M., Fong, A., Chamberlain, E., Bowman, J., Scharien, R., and Persson, O.: Annual cycle observations of aerosols capable of ice formation in central Arctic clouds, *Nat. Commun.*, 13, 3537, <https://doi.org/10.1038/s41467-022-31182-x>, 2022.

Creamean, J. M., Dexheimer, D., Hume, C. C., Vazquez, M., Hess, B. T. M., Longbottom, C. M., Ruiz, C. A., and Theisen, A. K.: Reaching new heights: A vertically-resolved ice nucleating particle sampler operating on Atmospheric Radiation Measurement (ARM) tethered balloon systems, *Atmos Meas Tech Discuss*, submitted, 2025.

Cziczko, D. J., Ladino, L., Boose, Y., Kanji, Z. A., Kupiszewski, P., Lance, S., Mertes, S., and Wex, H.: Measurements of Ice Nucleating Particles and Ice Residuals, <https://doi.org/10.1175/AMSMONOGRAPHS-D-16-0008.1>, 2017.

Daily, M. I., Tarn, M. D., Whale, T. F., and Murray, B. J.: An evaluation of the heat test for the ice-nucleating ability of minerals and biological material, *Atmospheric Meas. Tech.*, 15, 2635–2665, <https://doi.org/10.5194/amt-15-2635-2022>, 2022.

Delgado-Baquerizo, M., Oliverio, A. M., Brewer, T. E., Benavent-González, A., Eldridge, D. J., Bardgett, R. D., Maestre, F. T., Singh, B. K., and Fierer, N.: A global atlas of the dominant bacteria found in soil, *Science*, 359, 320–325, <https://doi.org/10.1126/science.aap9516>, 2018.

DeMott, P. and Hill, T.: COMBLE ARM Mobile Facility (AMF) Measurements of Ice Nucleating Particles Field Campaign Report, <https://doi.org/10.2172/1767118>, 2021.

DeMott, P., Hill, T., Suski, K., and Levin, E.: Southern Great Plains Ice Nuclei Characterization Experiment Final Campaign Summary, 2015.

DeMott, P., Hill, T. C., Marchand, R., and Alexander, S.: Macquarie Island Cloud and Radiation Experiment (MICRE) Ice Nucleating Particle Measurements Field Campaign Report, 2018a.

DeMott, P., Hill, T., and McFarquhar, G.: Measurements of Aerosols, Radiation, and Clouds over the Southern Ocean (MARCUS) Ice Nucleating Particle Measurements Field Campaign Report, 2018b.

DeMott, P. J.: An Exploratory Study of Ice Nucleation by Soot Aerosols, *J. Appl. Meteorol. Climatol.*, 29, 1072–1079, [https://doi.org/10.1175/1520-0450\(1990\)029%253C:1072:AESOI%253E2.0.CO;2](https://doi.org/10.1175/1520-0450(1990)029%253C:1072:AESOI%253E2.0.CO;2), 1990.

DeMott, P. J. and Hill, T. C.: ACAPEX – Ship-Based Ice Nuclei Collections Field Campaign Report, <https://doi.org/10.2172/1253893>, 2016.

DeMott, P. J. and Hill, T. C. J.: Cloud, Aerosol, and Complex Terrain Interactions (CACTI) ARM Aerial Facility (AAF) Measurements of Ice Nucleating Particles Field Campaign Report, 2020.

DeMott, P. J., Hill, T. C. J., McCluskey, C. S., Prather, K. A., Collins, D. B., Sullivan, R. C., Ruppel, M. J., Mason, R. H., Irish, V. E., Lee, T., Hwang, C. Y., Rhee, T. S., Snider, J. R., McMeeking, G. R., Dhaniyala, S., Lewis, E. R., Wentzell, J. J. B., Abbatt, J., Lee, C., Sultana, C. M., Ault, A. P., Axson, J. L., Diaz Martinez, M., Venero, I., Santos-Figueroa, G., Stokes, M. D., Deane, G. B., Mayol-Bracero, O. L., Grassian, V. H., Bertram, T. H., Bertram, A. K., Moffett, B. F., and Franc, G. D.: Sea spray aerosol as a unique source of ice nucleating particles, *Proc. Natl. Acad. Sci.*, 113, 5797–5803, <https://doi.org/10.1073/pnas.1514034112>, 2016.

DeMott, P. J., Hill, T. C. J., Petters, M. D., Bertram, A. K., Tobo, Y., Mason, R. H., Suski, K. J., McCluskey, C. S., Levin, E. J. T., Schill, G. P., Boose, Y., Rauker, A. M., Miller, A. J., Zaragoza, J., Rocci, K., Rothfuss, N. E., Taylor, H. P., Hader, J. D., Chou, C., Huffman, J. A., Pöschl, U., Prenni, A. J., and Kreidenweis, S. M.: Comparative measurements of ambient atmospheric concentrations of ice nucleating particles using multiple immersion freezing methods and a continuous flow diffusion chamber, *Atmospheric Chem. Phys.*, 17, 11227–11245, <https://doi.org/10.5194/acp-17-11227-2017>, 2017.

DeMott, P. J., Mason, R. H., McCluskey, C. S., Hill, T. C. J., Perkins, R. J., Desyaterik, Y., Bertram, A. K., Trueblood, J. V., Grassian, V. H., Qiu, Y., Molinero, V., Tobo, Y., Sultana, C. M., Lee, C., and Prather, K. A.: Ice nucleation by particles containing long-chain fatty acids of relevance to freezing by sea spray aerosols, *Environ. Sci. Process. Impacts*, 20, 1559–1569, <https://doi.org/10.1039/C8EM00386F>, 2018c.

DeMott, P. J., Möhler, O., Cziczo, D. J., Hiranuma, N., Petters, M. D., Petters, S. S., Belosi, F., Bingemer, H. G., Brooks, S. D., Budke, C., Burkert-Kohn, M., Collier, K. N., Danielczok, A., Eppers, O., Felgitsch, L., Garimella, S., Grothe, H., Herenz, P., Hill, T. C. J., Höhler, K., Kanji, Z. A., Kiselev, A., Koop, T., Kristensen, T. B., Krüger, K., Kulkarni, G., Levin, E. J. T., Murray, B. J., Nicosia, A., O’Sullivan, D., Peckhaus, A., Polen, M. J., Price, H. C., Reicher, N., Rothenberg, D. A., Rudich, Y., Santachiara, G., Schiebel, T., Schrod, J., Seifried, T. M., Stratmann, F., Sullivan, R. C., Suski, K. J., Szakáll, M., Taylor, H. P., Ullrich, R., Vergara-Temprado, J., Wagner, R., Whale, T. F., Weber, D., Welti, A., Wilson, T. W., Wolf, M. J., and Zenker, J.: The Fifth International Workshop on Ice Nucleation phase 2 (FIN-02): laboratory intercomparison of ice nucleation measurements, *Atmospheric Meas. Tech.*, 11, 6231–6257, <https://doi.org/10.5194/amt-11-6231-2018>, 2018d.

DeMott, P. J., Hill, T. C. J., Moore, K. A., Perkins, R. J., Mael, L. E., Busse, H. L., Lee, H., Kaluarachchi, C. P., Mayer, K. J., Sauer, J. S., Mitts, B. A., Tivanski, A. V., Grassian, V. H., Cappa, C. D., Bertram, T. H., and Prather, K. A.: Atmospheric oxidation impact on sea spray produced ice nucleating particles, *Environ. Sci. Atmospheres*, 3, 1513–1532, <https://doi.org/10.1039/D3EA00060E>, 2023.

DeMott, P. J., Mirrieles, J. A., Petters, S. S., Cziczo, D. J., Petters, M. D., Bingemer, H. G., Hill, T. C. J., Froyd, K., Garimella, S., Hallar, A. G., Levin, E. J. T., McCubbin, I. B., Perring, A. E., Rapp, C. N., Schiebel, T., Schrod, J., Suski, K. J., Weber, D., Wolf, M. J., Zawadowicz, M., Zenker, J., Möhler, O., and Brooks, S. D.: Field intercomparison of ice nucleation measurements: the Fifth International Workshop on Ice Nucleation Phase 3 (FIN-03), *Atmospheric Meas. Tech.*, 18, 639–672, <https://doi.org/10.5194/amt-18-639-2025>, 2025a.

DeMott, P. J., Swanson, B. E., Creamean, J. M., Tobo, Y., Hill, T. C. J., Barry, K. R., Beck, I. F., Frietas, G. P., Heslin-Rees, D., Lackner, C. P., Schmale, J., Krejci, R., Zieger, P., Geerts, B., and Kreidenweis, S. M.: Ice nucleating particle sources and transports between the Central and Southern Arctic regions during winter cold air outbreaks, *Elem. Sci. Anthr.*, 13, 00063, <https://doi.org/10.1525/elementa.2024.00063>, 2025b.

Després, Viviane R., Huffman, J. A., Burrows, S. M., Hoose, C., Safatov, Aleksandr S., Buryak, G., Fröhlich-Nowoisky, J., Elbert, W., Andreae, Meinrat O., Pöschl, U., and Jaenicke, R.: Primary biological aerosol particles in the atmosphere: a review, *Tellus B Chem. Phys. Meteorol.*, 64, 15598, <https://doi.org/10.3402/tellusb.v64i0.15598>, 2012.

'14 Dexheimer, D., Whitson, G., Cheng, Z., Sammon, J., Gaustad, K., Mei, F., and Longbottom, C.: Tethered Balloon System
'15 (TBS) Instrument Handbook, <https://doi.org/10.2172/1415858>, 2024.

'16 Ervens, B. and Feingold, G.: Sensitivities of immersion freezing: Reconciling classical nucleation theory and deterministic
'17 expressions, *Geophys. Res. Lett.*, 40, 3320–3324, <https://doi.org/10.1002/grl.50580>, 2013.

'18 Eufemio, R. J., de Almeida Ribeiro, I., Sformo, T. L., Laursen, G. A., Molinero, V., Fröhlich-Nowoisky, J., Bonn, M., and
'19 Meister, K.: Lichen species across Alaska produce highly active and stable ice nucleators, *Biogeosciences*, 20, 2805–2812,
'20 <https://doi.org/10.5194/bg-20-2805-2023>, 2023.

'21 Evans, S.: Dust-producing weather patterns of the North American Great Plains, *Atmospheric Chem. Phys.*, 25, 4833–4845,
'22 <https://doi.org/10.5194/acp-25-4833-2025>, 2025.

'23 Fan, J., Leung, L. R., DeMott, P. J., Comstock, J. M., Singh, B., Rosenfeld, D., Tomlinson, J. M., White, A., Prather, K. A.,
'24 Minnis, P., Ayers, J. K., and Min, Q.: Aerosol impacts on California winter clouds and precipitation during CalWater 2011:
'25 local pollution versus long-range transported dust, *Atmospheric Chem. Phys.*, 14, 81–101, [https://doi.org/10.5194/acp-14-81-](https://doi.org/10.5194/acp-14-81-2014)
'26 2014, 2014.

'27 Feldman, D. R., Aiken, A. C., Boos, W. R., Carroll, R. W. H., Chandrasekar, V., Collis, S., Creamean, J. M., Boer, G. de,
'28 Deems, J., DeMott, P. J., Fan, J., Flores, A. N., Gochis, D., Grover, M., Hill, T. C. J., Hodshire, A., Hulm, E., Hume, C. C.,
'29 Jackson, R., Junyent, F., Kennedy, A., Kumjian, M., Levin, E. J. T., Lundquist, J. D., O'Brien, J., Raleigh, M. S., Reithel, J.,
'30 Rhoades, A., Rittger, K., Rudisill, W., Sherman, Z., Siirila-Woodburn, E., Skiles, S. M., Smith, J. N., Sullivan, R. C., Theisen,
'31 A., Tuftedal, M., Varble, A. C., Wiedlea, A., Wielandt, S., Williams, K., and Xu, Z.: The Surface Atmosphere Integrated Field
'32 Laboratory (SAIL) Campaign, <https://doi.org/10.1175/BAMS-D-22-0049.1>, 2023.

'33 Fountain, A. G. and Ohtake, T.: Concentrations and Source Areas of Ice Nuclei in the Alaskan Atmosphere, *J. Clim. Appl.*
'34 *Meteorol.*, 24, 377–382, [https://doi.org/10.1175/1520-0450\(1985\)024%024%253C0377:CASAOI%253E2.0.CO;2](https://doi.org/10.1175/1520-0450(1985)024%024%253C0377:CASAOI%253E2.0.CO;2), 1985.

'35 Garcia, E., Hill, T. C. J., Prenni, A. J., DeMott, P. J., Franc, G. D., and Kreidenweis, S. M.: Biogenic ice nuclei in boundary
'36 layer air over two U.S. High Plains agricultural regions, *J. Geophys. Res. Atmospheres*, 117,
'37 <https://doi.org/10.1029/2012JD018343>, 2012.

'38 Geerts, B., McFarquhar, G., Xue, L., Jensen, M., Kollias, P., Ovchinnikov, M., Shupe, M., DeMott, P., Wang, Y., Tjernstrom,
'39 M., Field, P., Abel, S., Spengler, T., Neggers, R., Crewell, S., Wendisch, M., and Lupkes, C.: Cold-Air Outbreaks in the Marine
'40 Boundary Layer Experiment (COMBLE) Field Campaign Report, <https://doi.org/10.2172/1763013>, 2021.

'41 Geerts, B., Giangrande, S. E., McFarquhar, G. M., Xue, L., Abel, S. J., Comstock, J. M., Crewell, S., DeMott, P. J., Ebell, K.,
'42 Field, P., Hill, T. C. J., Hunzinger, A., Jensen, M. P., Johnson, K. L., Juliano, T. W., Kollias, P., Kosovic, B., Lackner, C.,
'43 Luke, E., Lüpkes, C., Matthews, A. A., Neggers, R., Ovchinnikov, M., Powers, H., Shupe, M. D., Spengler, T., Swanson, B.
'44 E., Tjernström, M., Theisen, A. K., Wales, N. A., Wang, Y., Wendisch, M., and Wu, P.: The COMBLE Campaign: A Study
'45 of Marine Boundary Layer Clouds in Arctic Cold-Air Outbreaks, *Bull. Am. Meteorol. Soc.*, 103, E1371–E1389,
'46 <https://doi.org/10.1175/BAMS-D-21-0044.1>, 2022.

'47 Ginoux, P., Prospero, J. M., Gill, T. E., Hsu, N. C., and Zhao, M.: Global-scale attribution of anthropogenic and natural dust
'48 sources and their emission rates based on MODIS Deep Blue aerosol products, *Rev. Geophys.*, 50,
'49 <https://doi.org/10.1029/2012RG000388>, 2012.

'50 Gratzl, J., Böhmländer, A., Pätsi, S., Pogner, C.-E., Gorfer, M., Brus, D., Doulgeris, K. M., Wieland, F., Asmi, E., Saarto, A.,
'51 Möhler, O., Stolzenburg, D., and Grothe, H.: Locally emitted fungal spores serve as high temperature ice nucleating particles
'52 in the European sub-Arctic, *EGUsphere*, 1–38, <https://doi.org/10.5194/egusphere-2025-1599>, 2025.

'53 Griesche, H. J., Ohneiser, K., Seifert, P., Radenz, M., Engelmann, R., and Ansmann, A.: Contrasting ice formation in Arctic
'54 clouds: surface-coupled vs. surface-decoupled clouds, *Atmospheric Chem. Phys.*, 21, 10357–10374,
'55 <https://doi.org/10.5194/acp-21-10357-2021>, 2021.

'56 Gunsch, M. J., Kirpes, R. M., Kolesar, K. R., Barrett, T. E., China, S., Sheesley, R. J., Laskin, A., Wiedensohler, A., Tuch, T.,
'57 and Pratt, K. A.: Contributions of transported Prudhoe Bay oil field emissions to the aerosol population in Utqiagvik, Alaska,
'58 *Atmospheric Chem. Phys.*, 17, 10879–10892, <https://doi.org/10.5194/acp-17-10879-2017>, 2017.

'59 Gute, E. and Abbatt, J. P. D.: Ice nucleating behavior of different tree pollen in the immersion mode, *Atmos. Environ.*, 231,
'60 117488, <https://doi.org/10.1016/j.atmosenv.2020.117488>, 2020.

'61 Hasenkopf, C. A., Veghte, D. P., Schill, G. P., Lodoysamba, S., Freedman, M. A., and Tolbert, M. A.: Ice nucleation, shape,
'62 and composition of aerosol particles in one of the most polluted cities in the world: Ulaanbaatar, Mongolia, *Atmos. Environ.*,
'63 139, 222–229, <https://doi.org/10.1016/j.atmosenv.2016.05.037>, 2016.

'64 Hill, T. C. J., DeMott, P. J., Tobo, Y., Fröhlich-Nowoisky, J., Moffett, B. F., Franc, G. D., and Kreidenweis, S. M.: Sources
'65 of organic ice nucleating particles in soils, *Atmospheric Chem. Phys.*, 16, 7195–7211, [https://doi.org/10.5194/acp-16-7195-](https://doi.org/10.5194/acp-16-7195-2016)
'66 2016, 2016.

'67 Hill, T. C. J., Malfatti, F., McCluskey, C. S., Schill, G. P., Santander, M. V., Moore, K. A., Rauker, A. M., Perkins, R. J.,
'68 Celussi, M., Levin, E. J. T., Suski, K. J., Cornwell, G. C., Lee, C., Negro, P. D., Kreidenweis, S. M., Prather, K. A., and
'69 DeMott, P. J.: Resolving the controls over the production and emission of ice-nucleating particles in sea spray, *Environ. Sci.*
'70 *Atmospheres*, 3, 970–990, <https://doi.org/10.1039/D2EA00154C>, 2023.

'71 Hoose, C. and Möhler, O.: Heterogeneous ice nucleation on atmospheric aerosols: a review of results from laboratory
'72 experiments, *Atmospheric Chem. Phys.*, 12, 9817–9854, <https://doi.org/10.5194/acp-12-9817-2012>, 2012.

'73 Huang, S., Hu, W., Chen, J., Wu, Z., Zhang, D., and Fu, P.: Overview of biological ice nucleating particles in the atmosphere,
'74 *Environ. Int.*, 146, 106197, <https://doi.org/10.1016/j.envint.2020.106197>, 2021.

'75 Janine Fröhlich-Nowoisky, Kampf, C. J., Weber, B., Huffman, J. A., Pöhlker, C., Andreae, M. O., Lang-Yona, N., Burrows,
'76 S. M., Gunthe, S. S., Elbert, W., Su, H., Hoor, P., Thines, E., Hoffmann, T., Després, V. R., and Pöschl, U.: Bioaerosols in the
'77 Earth system: Climate, health, and ecosystem interactions, *Atmospheric Res.*, 182, 346–376,
'78 <https://doi.org/10.1016/j.atmosres.2016.07.018>, 2016.

'79 Jensen, M., Flynn, J., Kollias, P., Kuang, C., McFarquhar, G., Powers, H., Brooks, S., Bruning, E., Collins, D., Collis, S., Fan,
'80 J., Fridlind, A., Giangrande, S., Griffin, R., Hu, J., Jackson, R., Kumjian, M., Logan, T., Matsui, T., Nowotarski, C., Oue, M.,
'81 Rapp, A., Rosenfeld, D., Ryzhkov, A., Sheesley, R., Snyder, J., Stier, P., Usenko, S., Van Den Heever, S., Van Lier-Walqui,
'82 M., Varble, A., Wang, Y., Aiken, A., Deng, M., Dexheimer, D., Dubey, M., Feng, Y., Ghate, V., Johnson, K., Lamer, K.,
'83 Saleeby, S., Wang, D., Zawadowicz, M., and Zhou, A.: Tracking Aerosol Convection Interactions Experiment (TRACER)
'84 Field Campaign Report, <https://doi.org/10.2172/2202672>, 2023.

'85 Kanji, Z. A., Ladino, L. A., Wex, H., Boose, Y., Burkert-Kohn, M., Cziczo, D. J., and Krämer, M.: Overview of ice nucleating
'86 particles, *Meteor Monogr*, 58, 1.1–1.33, <https://doi.org/10.1175/AMSMONOGRAPH5-D-16-0006.1>, 2017.

'87 Kaufmann, L., Marcolli, C., Hofer, J., Pinti, V., Hoyle, C. R., and Peter, T.: Ice nucleation efficiency of natural dust samples
'88 in the immersion mode, *Atmospheric Chem. Phys.*, 16, 11177–11206, <https://doi.org/10.5194/acp-16-11177-2016>, 2016.

'89 Knopf, D. A. and Alpert, P. A.: Atmospheric ice nucleation, *Nat. Rev. Phys.*, 5, 203–217, [https://doi.org/10.1038/s42254-023-](https://doi.org/10.1038/s42254-023-00570-7)
'90 00570-7, 2023.

Knopf, D. A., DeMott, P., Creamean, J., Riemer, N., Hiranuma, N., Laskin, A., Sullivan, R., Fridlind, A., Liu, X., West, M., and Hill, T.: Aerosol-Ice Formation Closure Pilot Study (AEROICESTUDY) Field Campaign Report, 2020.

Knopf, D. A., Barry, K. R., Brubaker, T. A., Jahl, L. G., Jankowski, K. A., Li, J., Lu, Y., Monroe, L. W., Moore, K. A., Rivera-Adorno, F. A., Saucedo, K. A., Shi, Y., Tomlin, J. M., Vepuri, H. S. K., Wang, P., Lata, N. N., Levin, E. J. T., Creamean, J. M., Hill, T. C. J., China, S., Alpert, P. A., Moffet, R. C., Hiranuma, N., Sullivan, R. C., Fridlind, A. M., West, M., Riemer, N., Laskin, A., DeMott, P. J., and Liu, X.: Aerosol-Ice Formation Closure: A Southern Great Plains Field Campaign, *Bull. Am. Meteorol. Soc.*, 102, E1952–E1971, <https://doi.org/10.1175/BAMS-D-20-0151.1>, 2021.

Lacher, L., Adams, M. P., Barry, K., Bertozzi, B., Bingemer, H., Boffo, C., Bras, Y., Büttner, N., Castarede, D., Cziczó, D. J., DeMott, P. J., Fösig, R., Goodell, M., Höhler, K., Hill, T. C. J., Jentsch, C., Ladino, L. A., Levin, E. J. T., Mertes, S., Möhler, O., Moore, K. A., Murray, B. J., Nadolny, J., Pfeuffer, T., Picard, D., Ramírez-Romero, C., Ribeiro, M., Richter, S., Schrod, J., Sellegri, K., Stratmann, F., Swanson, B. E., Thomson, E. S., Wex, H., Wolf, M. J., and Freney, E.: The Puy de Dôme Ice Nucleation Intercomparison Campaign (PICNIC): comparison between online and offline methods in ambient air, *Atmospheric Chem. Phys.*, 24, 2651–2678, <https://doi.org/10.5194/acp-24-2651-2024>, 2024.

Leung, L. R.: ARM Cloud-Aerosol-Precipitation Experiment (ACAPEX) Field Campaign Report, U.S. Department of Energy, Office of Science, Office of Biological and Environmental Research, 2016.

Levin, E. J. T., McMeeking, G. R., Carrico, C. M., Mack, L. E., Kreidenweis, S. M., Wold, C. E., Moosmüller, H., Arnott, W. P., Hao, W. M., Collett Jr., J. L., and Malm, W. C.: Biomass burning smoke aerosol properties measured during Fire Laboratory at Missoula Experiments (FLAME), *J. Geophys. Res. Atmospheres*, 115, <https://doi.org/10.1029/2009JD013601>, 2010.

Levin, E. J. T., DeMott, P. J., Suski, K. J., Boose, Y., Hill, T. C. J., McCluskey, C. S., Schill, G. P., Rocci, K., Al-Mashat, H., Kristensen, L. J., Cornwell, G., Prather, K., Tomlinson, J., Mei, F., Hubbe, J., Pekour, M., Sullivan, R., Leung, L. R., and Kreidenweis, S. M.: Characteristics of Ice Nucleating Particles in and Around California Winter Storms, *J. Geophys. Res. Atmospheres*, 124, 11530–11551, <https://doi.org/10.1029/2019JD030831>, 2019.

Lin, Y., Fan, J., Li, P., Leung, L. R., DeMott, P. J., Goldberger, L., Comstock, J., Liu, Y., Jeong, J.-H., and Tomlinson, J.: Modeling impacts of ice-nucleating particles from marine aerosols on mixed-phase orographic clouds during 2015 ACAPEX field campaign, *Atmospheric Chem. Phys.*, 22, 6749–6771, <https://doi.org/10.5194/acp-22-6749-2022>, 2022.

Maki, L. R., Galyan, E. L., Chang-Chien, M.-M., and Caldwell, D. R.: Ice Nucleation Induced by *Pseudomonas syringae*1, *Appl. Microbiol.*, 28, 456–459, 1974.

Marchand, R.: Macquarie Island Cloud and Radiation Experiment (MICRE) Field Campaign Report, <https://doi.org/10.2172/1602536>, 2020.

McCluskey, C. S., Hill, T. C. J., Malfatti, F., Sultana, C. M., Lee, C., Santander, M. V., Beall, C. M., Moore, K. A., Cornwell, G. C., Collins, D. B., Prather, K. A., Jayarathne, T., Stone, E. A., Azam, F., Kreidenweis, S. M., and DeMott, P. J.: A Dynamic Link between Ice Nucleating Particles Released in Nascent Sea Spray Aerosol and Oceanic Biological Activity during Two Mesocosm Experiments, *J. Atmospheric Sci.*, 74, 151–166, <https://doi.org/10.1175/JAS-D-16-0087.1>, 2017.

McCluskey, C. S., Hill, T. C. J., Sultana, C. M., Laskina, O., Trueblood, J., Santander, M. V., Beall, C. M., Michaud, J. M., Kreidenweis, S. M., Prather, K. A., Grassian, V., and DeMott, P. J.: A mesocosm double feature: Insights into the chemical makeup of marine ice nucleating particles, *J. Atmospheric Sci.*, 75, 2405–2423, <https://doi.org/10.1175/JAS-D-17-0155.1>, 2018a.

McCluskey, C. S., Ovadnevaite, J., Rinaldi, M., Atkinson, J., Belosi, F., Ceburnis, D., Marullo, S., Hill, T. C. J., Lohmann, U., Kanji, Z. A., O'Dowd, C., Kreidenweis, S. M., and DeMott, P. J.: Marine and terrestrial organic ice-nucleating particles in

pristine marine to continentally influenced Northeast Atlantic air masses, *J. Geophys. Res. Atmospheres*, 123, 6196–6212, <https://doi.org/10.1029/2017JD028033>, 2018b.

McCluskey, C. S., Hill, T. C. J., Humphries, R. S., Rauker, A. M., Moreau, S., Strutton, P. G., Chambers, S. D., Williams, A. G., McRobert, I., Ward, J., Keywood, M. D., Harnwell, J., Ponsonby, W., Loh, Z. M., Krummel, P. B., Protat, A., Kreidenweis, S. M., and DeMott, P. J.: Observations of ice nucleating particles over Southern Ocean waters, *Geophys. Res. Lett.*, 45, 11,989–11,997, <https://doi.org/10.1029/2018GL079981>, 2018c.

McCluskey, C. S., Gettelman, A., Bardeen, C. G., DeMott, P. J., Moore, K. A., Kreidenweis, S. M., Hill, T. C. J., Barry, K. R., Toohey, C. H., Toohey, D. W., Rainwater, B., Jensen, J. B., Reeves, J. M., Alexander, S. P., and McFarquhar, G. M.: Simulating Southern Ocean aerosol and ice nucleating particles in the Community Earth System Model version 2, *J. Geophys. Res. Atmospheres*, 128, e2022JD036955, <https://doi.org/10.1029/2022JD036955>, 2023.

McFarquhar, G., Marchand, R., Bretherton, C., Alexander, S., Protat, A., Siems, S., Wood, R., and DeMott, P.: Measurements of Aerosols, Radiation, and Clouds Over the Southern Ocean (MARCUS) Field Campaign Report, 2019.

McFarquhar, G. M., Bretherton, C. S., Marchand, R., Protat, A., DeMott, P. J., Alexander, S. P., Roberts, G. C., Toohey, C. H., Toohey, D., Siems, S., Huang, Y., Wood, R., Rauber, R. M., Lasher-Trapp, S., Jensen, J., Stith, J. L., Mace, J., Um, J., Järvinen, E., Schnaiter, M., Gettelman, A., Sanchez, K. J., McCluskey, C. S., Russell, L. M., McCoy, I. L., Atlas, R. L., Bardeen, C. G., Moore, K. A., Hill, T. C. J., Humphries, R. S., Keywood, M. D., Ristovski, Z., Cravigan, L., Schofield, R., Fairall, C., Mallet, M. D., Kreidenweis, S. M., Rainwater, B., D’Alessandro, J., Wang, Y., Wu, W., Saliba, G., Levin, E. J. T., Ding, S., Lang, F., Truong, S. C. H., Wolff, C., Haggerty, J., Harvey, M. J., Klekociuk, A. R., and McDonald, A.: Observations of clouds, aerosols, precipitation, and surface radiation over the Southern Ocean: An overview of CAPRICORN, MARCUS, MICRE, and SOCRATES, *Bull. Am. Meteorol. Soc.*, 102, E894–E928, <https://doi.org/10.1175/BAMS-D-20-0132.1>, 2021.

McLean, W.: The Nature of Soil Organic Matter as Shown by the attack of Hydrogen Peroxide, *J. Agric. Sci.*, 21, 595–611, <https://doi.org/10.1017/S002185960009813>, 1931.

Mikutta, R., Kleber, M., Kaiser, K., and Jahn, R.: Review: Organic Matter Removal from Soils using Hydrogen Peroxide, Sodium Hypochlorite, and Disodium Peroxodisulfate, *Soil Sci. Soc. Am. J.*, 69, 120–135, <https://doi.org/10.2136/sssaj2005.0120>, 2005.

Moore, K. A., Hill, T. C. J., Madawala, C. K., Leibensperger III, R. J., Greeney, S., Cappa, C. D., Stokes, M. D., Deane, G. B., Lee, C., Tivanski, A. V., Prather, K. A., and DeMott, P. J.: Wind-driven emission of marine ice-nucleating particles in the Scripps Ocean-Atmosphere Research Simulator (SOARS), *Atmospheric Chem. Phys.*, 25, 3131–3159, <https://doi.org/10.5194/acp-25-3131-2025>, 2025.

Nieto-Caballero, M., Barry, K. R., Hill, T. C. J., Douglas, T. A., DeMott, P. J., Kreidenweis, S. M., and Creamean, J. M.: Airborne bacteria over thawing permafrost landscapes in the Arctic, *Environ. Sci. Technol.*, 59, 9027–9036, <https://doi.org/10.1021/acs.est.4c11774>, 2025.

Niu, Q., McFarquhar, G. M., Marchand, R., Theisen, A., Cavallo, S. M., Flynn, C., DeMott, P. J., McCluskey, C. S., Humphries, R. S., and Hill, T. C. J.: 62°S Witnesses the Transition of Boundary Layer Marine Aerosol Pattern Over the Southern Ocean (50°S–68°S, 63°E–150°E) During the Spring and Summer: Results From MARCUS (I), *J. Geophys. Res. Atmospheres*, 129, e2023JD040396, <https://doi.org/10.1029/2023JD040396>, 2024.

O’Sullivan, D., Murray, B. J., Malkin, T. L., Whale, T. F., Umo, N. S., Atkinson, J. D., Price, H. C., Baustian, K. J., Browne, J., and Webb, M. E.: Ice nucleation by fertile soil dusts: relative importance of mineral and biogenic components, *Atmospheric Chem. Phys.*, 14, 1853–1867, <https://doi.org/10.5194/acp-14-1853-2014>, 2014.

69 O'Sullivan, D., Murray, B. J., Ross, J. F., and Webb, M. E.: The adsorption of fungal ice-nucleating proteins on mineral dusts:
70 a terrestrial reservoir of atmospheric ice-nucleating particles, *Atmospheric Chem. Phys.*, 16, 7879–7887,
71 <https://doi.org/10.5194/acp-16-7879-2016>, 2016.

72 Pereira, D. L., Gavilán, I., Letechipia, C., Raga, G. B., Puig, T. P., Mugica-Álvarez, V., Alvarez-Ospina, H., Rosas, I.,
73 Martinez, L., Salinas, E., Quintana, E. T., Rosas, D., and Ladino, L. A.: Mexican agricultural soil dust as a source of ice
74 nucleating particles, *Atmospheric Chem. Phys.*, 22, 6435–6447, <https://doi.org/10.5194/acp-22-6435-2022>, 2022.

75 Pereira Freitas, G., Adachi, K., Conen, F., Heslin-Rees, D., Krejci, R., Tobo, Y., Yttri, K. E., and Zieger, P.: Regionally sourced
76 bioaerosols drive high-temperature ice nucleating particles in the Arctic, *Nat. Commun.*, 14, 5997,
77 <https://doi.org/10.1038/s41467-023-41696-7>, 2023.

78 Perkins, R. J., Gillette, S. M., Hill, T. C. J., and DeMott, P. J.: The Labile Nature of Ice Nucleation by Arizona Test Dust, *ACS*
79 *Earth Space Chem.*, 4, 133–141, <https://doi.org/10.1021/acsearthspacechem.9b00304>, 2020.

80 Perring, A. E., Mediavilla, B., Wilbanks, G. D., Churnside, J. H., Marchbanks, R., Lamb, K. D., and Gao, R.-S.: Airborne
81 bioaerosol observations imply a strong terrestrial source in the summertime Arctic, *J. Geophys. Res. Atmospheres*, 128,
82 e2023JD039165, <https://doi.org/10.1029/2023JD039165>, 2023.

83 Raman, A., Hill, T., DeMott, P. J., Singh, B., Zhang, K., Ma, P.-L., Wu, M., Wang, H., Alexander, S. P., and Burrows, S. M.:
84 Long-term variability in immersion-mode marine ice-nucleating particles from climate model simulations and observations,
85 *Atmospheric Chem. Phys.*, 23, 5735–5762, <https://doi.org/10.5194/acp-23-5735-2023>, 2023.

86 Ren, Y. Z., Bi, K., Fu, S. Z., Tian, P., Huang, M. Y., Zhu, R. H., and Xue, H. W.: The Relationship of Aerosol Properties and
87 Ice-Nucleating Particle Concentrations in Beijing, *J. Geophys. Res. Atmospheres*, 128, e2022JD037383,
88 <https://doi.org/10.1029/2022JD037383>, 2023.

89 Robinson, G. W.: Note on the mechanical analysis of humus soils, *J. Agric. Sci.*, 12, 287–291,
90 <https://doi.org/10.1017/S0021859600005347>, 1922.

91 Rodriguez-Caballero, E., Stanelle, T., Egerer, S., Cheng, Y., Su, H., Canton, Y., Belnap, J., Andreae, M. O., Tegen, I., Reick,
92 C. H., Pöschl, U., and Weber, B.: Global cycling and climate effects of aeolian dust controlled by biological soil crusts, *Nat.*
93 *Geosci.*, 15, 458–463, <https://doi.org/10.1038/s41561-022-00942-1>, 2022.

94 Rogers, D. C., DeMott, P. J., and Kreidenweis, S. M.: Airborne measurements of tropospheric ice-nucleating aerosol particles
95 in the Arctic spring, *J. Geophys. Res. Atmospheres*, 106, 15053–15063, <https://doi.org/10.1029/2000JD900790>, 2001.

96 Roy, P., Mael, L. E., Hill, T. C. J., Mehndiratta, L., Peiker, G., House, M. L., DeMott, P. J., Grassian, V. H., and Dutcher, C.
97 S.: Ice Nucleating Activity and Residual Particle Morphology of Bulk Seawater and Sea Surface Microlayer, *ACS Earth Space*
98 *Chem.*, 5, 1916–1928, <https://doi.org/10.1021/acsearthspacechem.1c00175>, 2021.

99 Russell, L. M., Lubin, D., Silber, I., Eloranta, E., Muelmenstaedt, J., Burrows, S., Aiken, A., Wang, D., Petters, M., Miller,
00 M., Ackerman, A., Fridlind, A., Witte, M., Lebsock, M., Painemal, D., Chang, R., Liggio, J., and Wheeler, M.: EPCAPE Field
01 Campaign Final Campaign Report April, 2024.

02 Schiebel, T., Höhler, K., Funk, R., Hill, T. C. J., Levin, E. J. T., Nadolny, J., Steinke, I., Suski, K. J., Ullrich, R., Wagner, R.,
03 Weber, I., DeMott, P. J., and Möhler, O.: Ice nucleation activity of various agricultural soil dust aerosol particles, *European*
04 *Geosciences Union General Assembly, Wien, A*, April 17-22, 2016. *Geophysical Research Abstracts*, 18(2016) EGU2016-
05 13422, 2016.

006 Schneider, J., Höhler, K., Heikkilä, P., Keskinen, J., Bertozzi, B., Bogert, P., Schorr, T., Umo, N. S., Vogel, F., Brasseur, Z.,
 007 Wu, Y., Hakala, S., Duplissy, J., Moiseev, D., Kulmala, M., Adams, M. P., Murray, B. J., Korhonen, K., Hao, L., Thomson,
 008 E. S., Castarède, D., Leisner, T., Petäjä, T., and Möhler, O.: The seasonal cycle of ice-nucleating particles linked to the
 009 abundance of biogenic aerosol in boreal forests, *Atmospheric Chem. Phys.*, 21, 3899–3918, [https://doi.org/10.5194/acp-21-](https://doi.org/10.5194/acp-21-3899-2021)
 010 3899-2021, 2021.

011 Schnell, R. C. and Vali, G.: Biogenic Ice Nuclei: Part I. Terrestrial and Marine Sources, *J. Atmospheric Sci.*, 33, 1554–1564,
 012 [https://doi.org/10.1175/1520-0469\(1976\)033%253C1554:BINPIT%253E2.0.CO;2](https://doi.org/10.1175/1520-0469(1976)033%253C1554:BINPIT%253E2.0.CO;2), 1976.

013 Schrod, J., Thomson, E. S., Weber, D., Kossmann, J., Pöhlker, C., Saturno, J., Ditas, F., Artaxo, P., Clouard, V., Saurel, J.-M.,
 014 Ebert, M., Curtius, J., and Bingemer, H. G.: Long-term deposition and condensation ice-nucleating particle measurements
 015 from four stations across the globe, *Atmospheric Chem. Phys.*, 20, 15983–16006, <https://doi.org/10.5194/acp-20-15983-2020>,
 016 2020.

017 Schultz, M. K., Biegalski, S. R., Inn, K. G. W., Yu, L., Burnett, W. C., Thomas, J. L. W., and Smith, G. E.: Optimizing the
 018 removal of carbon phases in soils and sediments for sequential chemical extractions by coulometry, *J. Environ. Monit.*, 1, 183–
 019 190, <https://doi.org/10.1039/A900534J>, 1999.

020 Sequi, P. and Aringhieri, R.: Destruction of Organic Matter by Hydrogen Peroxide in the Presence of Pyrophosphate and Its
 021 Effect on Soil Specific Surface Area, *Soil Sci. Soc. Am. J.*, 41, 340–342,
 022 <https://doi.org/10.2136/sssaj1977.03615995004100020033x>, 1977.

023 Shupe, M., Chu, D., Costa, D., Cox, C., Creamean, J., De Boer, G., Dethloff, K., Engelmann, R., Gallagher, M., Hunke, E.,
 024 Maslowski, W., McComiskey, A., Osborn, J., Persson, O., Powers, H., Pratt, K., Randall, D., Solomon, A., Tjernstrom, M.,
 025 Turner, D., Uin, J., Uttal, T., Verlinde, J., and Wagner, D.: Multidisciplinary drifting Observatory for the Study of Arctic
 026 Climate (MOSAIC) (Field Campaign Report), <https://doi.org/10.2172/1787856>, 2021.

027 Shupe, M. D., Rex, M., Blomquist, B., Persson, P. O. G., Schmale, J., Uttal, T., Althausen, D., Angot, H., Archer, S., Bariteau,
 028 L., Beck, I., Bilberry, J., Bucci, S., Buck, C., Boyer, M., Brasseur, Z., Brooks, I. M., Calmer, R., Cassano, J., Castro, V., Chu,
 029 D., Costa, D., Cox, C. J., Creamean, J., Crewell, S., Dahlke, S., Damm, E., de Boer, G., Deckelmann, H., Dethloff, K., Dütsch,
 030 M., Ebell, K., Ehrlich, A., Ellis, J., Engelmann, R., Fong, A. A., Frey, M. M., Gallagher, M. R., Ganzeveld, L., Gradinger, R.,
 031 Graeser, J., Greenamyre, V., Griesche, H., Griffiths, S., Hamilton, J., Heinemann, G., Helmig, D., Herber, A., Heuzé, C.,
 032 Hofer, J., Houchens, T., Howard, D., Inoue, J., Jacobi, H.-W., Jaiser, R., Jokinen, T., Jourdan, O., Jozef, G., King, W.,
 033 Kirchgaessner, A., Klingebiel, M., Krassovski, M., Krumpen, T., Lampert, A., Landing, W., Laurila, T., Lawrence, D.,
 034 Lonardi, M., Loose, B., Lüpkes, C., Maahn, M., Macke, A., Maslowski, W., Marsay, C., Maturilli, M., Mech, M., Morris, S.,
 035 Moser, M., Nicolaus, M., Ortega, P., Osborn, J., Pätzold, F., Perovich, D. K., Petäjä, T., Pilz, C., Pirazzini, R., Posman, K.,
 036 Powers, H., Pratt, K. A., Preußner, A., Quéléver, L., Radenz, M., Rabe, B., Rinke, A., Sachs, T., Schulz, A., Siebert, H., Silva,
 037 T., Solomon, A., et al.: Overview of the MOSAiC expedition: *Atmosphere, Elem. Sci. Anthr.*, 10, 00060,
 038 <https://doi.org/10.1525/elementa.2021.00060>, 2022.

039 Song, Q., Zhang, Z., Yu, H., Ginoux, P., and Shen, J.: Global dust optical depth climatology derived from CALIOP and
 040 MODIS aerosol retrievals on decadal timescales: regional and interannual variability, *Atmospheric Chem. Phys.*, 21, 13369–
 041 13395, <https://doi.org/10.5194/acp-21-13369-2021>, 2021.

042 Spurny, K. R. and Lodge, J. P.: Collection Efficiency Tables for Membrane Filters Used in the Sampling and Analysis of
 043 Aerosols and Hydrosols, Laboratory of Atmospheric Science, National Center for Atmospheric Research, 56 pp., 1972.

044 Steinke, I., Funk, R., Busse, J., Iturri, A., Kirchen, S., Leue, M., Möhler, O., Schwartz, T., Schnaiter, M., Sierau, B., Toprak,
 045 E., Ullrich, R., Ulrich, A., Hoose, C., and Leisner, T.: Ice nucleation activity of agricultural soil dust aerosols from Mongolia,
 046 Argentina, and Germany, *J. Geophys. Res. Atmospheres*, 121, 13,559–13,576, <https://doi.org/10.1002/2016JD025160>, 2016.

47 Suski, K. J., Hill, T. C. J., Levin, E. J. T., Miller, A., DeMott, P. J., and Kreidenweis, S. M.: Agricultural harvesting emissions
 48 of ice-nucleating particles, *Atmospheric Chem. Phys.*, 18, 13755–13771, <https://doi.org/10.5194/acp-18-13755-2018>, 2018.

49 Teska, C. J., Dieser, M., and Foreman, C. M.: Clothing Textiles as Carriers of Biological Ice Nucleation Active Particles,
 50 *Environ. Sci. Technol.*, 58, 6305–6312, <https://doi.org/10.1021/acs.est.3c09600>, 2024.

51 Testa, B., Hill, T. C. J., Marsden, N. A., Barry, K. R., Hume, C. C., Bian, Q., Uetake, J., Hare, H., Perkins, R. J., Möhler, O.,
 52 Kreidenweis, S. M., and DeMott, P. J.: Ice Nucleating Particle Connections to Regional Argentinian Land Surface Emissions
 53 and Weather During the Cloud, Aerosol, and Complex Terrain Interactions Experiment, *J. Geophys. Res. Atmospheres*, 126,
 54 <https://doi.org/10.1029/2021JD035186>, 2021.

55 Tobo, Y., DeMott, P. J., Hill, T. C. J., Prenni, A. J., Swoboda-Colberg, N. G., Franc, G. D., and Kreidenweis, S. M.: Organic
 56 matter matters for ice nuclei of agricultural soil origin, *Atmospheric Chem. Phys.*, 14, 8521–8531, <https://doi.org/10.5194/acp-14-8521-2014>, 2014.

58 Tobo, Y., Adachi, K., DeMott, P. J., Hill, T. C. J., Hamilton, D. S., Mahowald, N. M., Nagatsuka, N., Ohata, S., Uetake, J.,
 59 Kondo, Y., and Koike, M.: Glacially sourced dust as a potentially significant source of ice nucleating particles, *Nat. Geosci.*,
 60 12, 253–258, <https://doi.org/10.1038/s41561-019-0314-x>, 2019.

61 Tobo, Y., Uetake, J., Matsui, H., Moteki, N., Uji, Y., Iwamoto, Y., Miura, K., and Misumi, R.: Seasonal Trends of Atmospheric
 62 Ice Nucleating Particles Over Tokyo, *J. Geophys. Res. Atmospheres*, 125, e2020JD033658,
 63 <https://doi.org/10.1029/2020JD033658>, 2020.

64 Vali, G.: Quantitative Evaluation of Experimental Results an the Heterogeneous Freezing Nucleation of Supercooled Liquids,
 65 *J. Atmospheric Sci.*, 28, 402–409, [https://doi.org/10.1175/1520-0469\(1971\)028%253C0402:QEOERA%253E2.0.CO;2](https://doi.org/10.1175/1520-0469(1971)028%253C0402:QEOERA%253E2.0.CO;2), 1971.

66 Vali, G., Christensen, M., Fresh, R. W., Galyan, E. L., Maki, L. R., and Schnell, R. C.: Biogenic Ice Nuclei. Part II: Bacterial
 67 Sources, *J. Atmospheric Sci.*, 33, 1565–1570, [https://doi.org/10.1175/1520-0469\(1976\)033%253C1565:BINPIB%253E2.0.CO;2](https://doi.org/10.1175/1520-0469(1976)033%253C1565:BINPIB%253E2.0.CO;2), 1976.

69 Varble, A., Nesbitt, S., Salio, P., Avila, E., Borque, P., McFarquhar, G., Van Den Heever, S., Zipser, E., Gochis, D., Houze
 70 Jr., R., Jensen, M., Kollias, P., Kreidenweis, S., Leung, R., Rasmussen, K., Romps, D., Williams, C., and DeMott, P.: Cloud,
 71 Aerosol, and Complex Terrain Interactions (CACTI) Field Campaign Report, <https://doi.org/10.2172/1574024>, 2019.

72 Wagh, S., Singh, P., Ghude, S. D., Safai, P., Prabhakaran, T., and Kumar, P. P.: Study of ice nucleating particles in fog-haze
 73 weather at New Delhi, India: A case of polluted environment, *Atmospheric Res.*, 259, 105693,
 74 <https://doi.org/10.1016/j.atmosres.2021.105693>, 2021.

75 Welti, A., Bigg, E. K., DeMott, P. J., Gong, X., Hartmann, M., Harvey, M., Henning, S., Herenz, P., Hill, T. C. J., Hornblow,
 76 B., Leck, C., Löffler, M., McCluskey, C. S., Rauker, A. M., Schmale, J., Tatzelt, C., Van Pinxteren, M., and Stratmann, F.:
 77 Ship-based measurements of ice nuclei concentrations over the Arctic, Atlantic, Pacific and Southern oceans, *Atmospheric
 78 Chem. Phys.*, 20, 15191–15206, <https://doi.org/10.5194/acp-20-15191-2020>, 2020.

79 Wex, H., Augustin-Bauditz, S., Boose, Y., Budke, C., Curtius, J., Diehl, K., Dreyer, A., Frank, F., Hartmann, S., Hiranuma,
 80 N., Jantsch, E., Kanji, Z. A., Kiselev, A., Koop, T., Möhler, O., Niedermeier, D., Nillius, B., Rösch, M., Rose, D., Schmidt,
 81 C., Steinke, I., and Stratmann, F.: Intercomparing different devices for the investigation of ice nucleating particles using
 82 Snomax[®] as test substance, *Atmospheric Chem. Phys.*, 15, 1463–1485, <https://doi.org/10.5194/acp-15-1463-2015>, 2015.

84 Wex, H., Huang, L., Zhang, W., Hung, H., Traversi, R., Becagli, S., Sheesley, R. J., Moffett, C. E., Barrett, T. E., Bossi, R.,

'85 Skov, H., Hünnerbein, A., Lubitz, J., Löffler, M., Linke, O., Hartmann, M., Herenz, P., and Stratmann, F.: Annual variability
'86 of ice-nucleating particle concentrations at different Arctic locations, *Atmospheric Chem. Phys.*, 19, 5293–5311,
'87 <https://doi.org/10.5194/acp-19-5293-2019>, 2019.

'88 Yadav, S., Venezia, R. E., Paerl, R. W., and Petters, M. D.: Characterization of Ice-Nucleating Particles Over Northern India,
'89 *J. Geophys. Res. Atmospheres*, 124, 10467–10482, <https://doi.org/10.1029/2019JD030702>, 2019.

'90 Zhang, C., Wu, Z., Chen, J., Chen, J., Tang, L., Zhu, W., Pei, X., Chen, S., Tian, P., Guo, S., Zeng, L., Hu, M., and Kanji, Z.
'91 A.: Ice-nucleating particles from multiple aerosol sources in the urban environment of Beijing under mixed-phase cloud
'92 conditions, *Atmospheric Chem. Phys.*, 22, 7539–7556, <https://doi.org/10.5194/acp-22-7539-2022>, 2022.

'93 Zhao, B., Wang, Y., Gu, Y., Liou, K.-N., Jiang, J. H., Fan, J., Liu, X., Huang, L., and Yung, Y. L.: Ice nucleation by aerosols
'94 from anthropogenic pollution, *Nat. Geosci.*, 12, 602–607, <https://doi.org/10.1038/s41561-019-0389-4>, 2019.