

1 **Microbial necromass contribution to topsoil organic carbon storage of natural**
2 **and agricultural ecosystems**

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21 **Abstract**

22 Microbial necromass is an important component of soil carbon (C). Yet, the relative
23 contribution of microbial necromass in shaping the global C stocks in agricultural and
24 natural ecosystems worldwide remains virtually unknown. Here, we compiled data on
25 fungal and bacterial necromass along with soil organic carbon (SOC) from the 0–20
26 cm soil layer across 486 study sites (145 agricultural and 341 natural ecosystems) to
27 evaluate the relative contribution of fungal necromass C (FNC) and bacterial
28 necromass C (BNC) to SOC. Our results indicated that, on average, FNC is two times
29 more important than BNC in explaining SOC in both agricultural and natural
30 ecosystems. The contributions of FNC and BNC to SOC were markedly higher in
31 agricultural ecosystems compared with natural ecosystems, with a contrasting trend in
32 the FNC/BNC ratio. Soil physicochemical properties (soil C/N ratio and clay content)
33 were the most important predictors of the contributions of FNC and BNC to SOC in
34 both ecosystems, while geographical factor (elevation) was the most important
35 predictor of the FNC/BNC ratio. Our study enhances the current level of
36 understanding regarding microbially mediated biogeochemical cycling and SOC
37 dynamics, underscoring the critical role of microbial necromass in the global C cycle.
38

39 **Keywords:** agricultural ecosystems, bacterial necromass carbon, fungal necromass
40 carbon, microbial necromass carbon, natural ecosystems
41

42 **1 Introduction**

43 As the largest carbon (C) pool in the terrestrial biosphere, soil organic carbon (SOC)
44 plays a pivotal role in shaping the global C cycle and climate system (Bellamy et al.,
45 2005; Crowther et al., 2015). In brief, plant inputs provide the primary carbon source
46 to soils, and microbial processing transforms these inputs into microbial necromass
47 that can persist over long turnover times (Cotrufo et al., 2013; Angst et al., 2021).
48 Although the living soil microbial biomass typically constitutes only about 2% of
49 SOC (a ratio referred to as the microbial quotient; Anderson & Domsch, 1989; Liu et
50 al., 2023), microbial necromass has been shown to contribute more than half and up to
51 approximately 80% of SOC, depending on soil type and analytical methods (Liang &
52 Balsler, 2011; Kallenbach et al., 2016; Liang et al., 2019). In other words, microbial
53 necromass C (MNC) constitutes a substantial and critical component of stable SOC
54 (Ma et al., 2018), and its dynamics are increasingly recognized for their role in
55 regulating the terrestrial carbon cycle and climate feedbacks (Zhao et al., 2023). As
56 such, there is growing scientific attention on the forces driving the accumulation of
57 MNC and its contribution to SOC (Liang et al., 2017; Ni et al., 2020; Luo et al., 2022;
58 Zhou et al., 2023). To gain a comprehensive understanding of MNC in the global C
59 cycle, recent research has highlighted the distinct roles of fungal and bacterial
60 necromass, revealing their contrasting responses to environmental and anthropogenic
61 drivers. For instance, studies have shown that the accumulation and contribution of
62 MNC are sensitive to factors such as aridity, primary productivity, agricultural
63 management practices like tillage and fertilization, as well as key soil properties
64 including pH and clay content (Zhang et al., 2021; Zhou et al., 2023; Xu et al., 2024).

65 Despite these advances, it remains unclear whether these organism-specific
66 mechanisms translate into systematic differences in necromass contributions between
67 ecosystems under varying degrees of human interference, such as agricultural versus
68 natural systems.

69 With the distinct roles of fungi and bacteria in decomposing organic matter and
70 stabilizing organic carbon in soil, the relative contribution to SOC of fungal and
71 bacterial necromass C could be used to track the dynamics of SOC storage (Malik et
72 al., 2016). The cell walls of fungi primarily consist of chitin (a nitrogen-containing
73 polysaccharide) and β -glucans, whereas bacterial cell walls are mainly composed of
74 peptidoglycan—a complex of sugars and amino acids (Lenardon et al., 2007). [As
75 bacterial amino sugars are readily degradable, while fungal chitin and -glucans are
76 more recalcitrant, fungal necromass generally exhibits a longer turnover time in soil
77 compared to bacterial necromass](#) (Xu et al., 2022). Wang et al. (2021a) reported that
78 the contribution of fungal necromass carbon (FNC) to SOC exceeded 65%,
79 considerably higher than that of bacterial necromass carbon (BNC, 32–36%). This
80 pattern is likely attributed to the slower decomposition rate and stronger
81 mineral-associative capacity of fungal necromass. Furthermore, greater fungal
82 biomass and higher turnover rates may enhance the input flux of fungal necromass
83 (Klink et al., 2022). The contributions of FNC and BNC to SOC depended on the type
84 of ecosystems (Wang et al., 2021a; Cao et al., 2023; Xu et al., 2024). However, few
85 studies on fungal and bacterial necromass carbon and their contribution to SOC has
86 been reported for ecosystems under human interference (Zhou et al., 2023).

87 Terrestrial ecosystems can be broadly categorized into managed (agricultural) and
88 natural ecosystems (Hobbs et al., 2011; Keith et al., 2022). The agricultural
89 ecosystems are typical of plant litter derived from single crops under intensive human
90 management (Bohan et al., 2013), a context that typically leads to bacterial-dominated
91 soil communities (van Der Heijden et al., 2008). In contrast, natural ecosystems
92 display greater diversity in plant litter and root deposits (Wu et al., 2019). In such
93 ecosystems, fungal mycelial networks and stable soil aggregates are enhanced,
94 leading to higher FNC contributions to SOC (Sanallah et al., 2020; Sae-Tun et al.,
95 2022). While bacteria are undoubtedly vital decomposers, fungi play a distinct and
96 often dominant role in the initial breakdown of complex plant polymers such as
97 cellulose and lignin. This functional prominence stems from their potent enzymatic
98 machinery and hyphal growth form, which enable physical penetration and decay of
99 solid organic matter (de Boer et al., 2005). As key decomposers, fungi are thus critical
100 in processing cellulose and other complex organic compounds (Hättenschwiler et al.,
101 2005). Accordingly, as demonstrated by Choi et al. (2018), soil cellulose-degrading
102 genes are frequently linked to fungal activity and abundance. Rather than implying
103 higher cellulose concentration per se, diverse plant inputs increase the chemical
104 heterogeneity of plant-derived polymers (e.g., cellulose, hemicelluloses, and lignin),
105 which broadens decomposer niches and often favors fungal communities in litter
106 horizons (Hättenschwiler et al., 2005; Štursová et al., 2012). In contrast, agricultural
107 monocultures tend to reduce fungal diversity unless mitigated by management
108 practices (Chen et al., 2020). Reflecting this context dependence, cellulose-rich inputs

109 can enrich saprotrophic fungi in arable soils (Clocchiatti et al., 2021), whereas
110 bacteria may contribute substantially in mineral soils or under specific microhabitat
111 and land management conditions (Štursová et al., 2012; Choi et al., 2018). Due to
112 distinct chemical properties and organo-mineral stabilization pathways, fungal and
113 bacterial necromass exhibit differing turnover times, making the FNC/BNC ratio a
114 mechanistic tracer of SOC formation (Angst et al., 2021; Kleber et al., 2021).
115 Therefore, elucidating the global distribution and drivers of FNC, BNC, and their ratio
116 across agricultural and natural ecosystems is essential for predicting
117 management-induced shifts in SOC under varying climatic and soil conditions (Zhang
118 et al., 2021; Zhou et al., 2023; Xu et al., 2024).

119 In order to explore the global patterns and drivers of FNC, BNC and the
120 FNC/BNC ratio in agricultural and natural ecosystems, we compiled data from 486
121 study sites worldwide. The aims of this study were: (1) to quantify the contributions
122 of FNC and BNC to SOC and the FNC/BNC ratio in agricultural and natural
123 ecosystems; and (2) to investigate the primary driving factors influencing the
124 contributions of FNC and BNC to SOC and the FNC/BNC ratio.

125

126 **2 Materials and methods**

127 2.1 Data collection

128 We compiled a comprehensive dataset following the stepwise workflow. (1) We
129 collected peer-reviewed papers published from 1996 to 31 December 2022 from Web
130 of Science (<http://apps.webofknowledge.com>), Google Scholar
131 (<http://scholar.google.com>), and the China National Knowledge Infrastructure
132 (<http://cnki.net>), using the keywords: ‘amino sugars’, ‘microbial necromass’,
133 ‘microbial residue’, ‘fungal residue’, and ‘bacterial residue’. Records from different
134 databases were merged and deduplicated to form an initial compilation. (2) We then
135 filtered the compiled studies to include only those focusing on topsoil, defined as the
136 0–20 cm layer. Studies reporting deeper or unspecified sampling depths (e.g., 0–30
137 cm) were excluded to ensure spatial comparability. (3) Full texts were assessed to
138 confirm the presence of paired fungal and bacterial residue data from the same
139 sample—specifically, glucosamine (GluN) and muramic acid (MurA), or directly
140 reported FNC and BNC values—to enable consistent cross-study calculation of the
141 FNC/BNC ratio. Studies lacking either biomarker were excluded from ratio analyses,
142 though those directly reporting the FNC/BNC ratio were retained. (4) Eligible
143 observations were classified into agricultural ecosystems (including dry land, irrigated
144 cropland, and submerged paddy) or natural ecosystems (forest and grassland) based
145 on study metadata. (5) For natural ecosystems, data from fertilized, polluted,
146 experimentally treated, or otherwise anthropogenically disturbed sites were excluded.
147 In total, the final dataset consisted of 2094 observations from 486 sites worldwide
148 (145 agricultural and 341 natural sites; Figure 1) reported in 164 peer-reviewed papers.
149 Of these observations, 1001 were from agricultural ecosystems, and 1093 from natural
150 ecosystems. Among the 341 natural sites, 195 were forests and 146 were grasslands.
151 For agricultural sites, we used Google Earth Engine with the LGRIP30 V1 dataset to
152 classify agricultural ecosystem into dry land and irrigated cropland, and we overlaid

153 the JRC surface-water seasonality layer to extract submerged paddy from the irrigated
154 class (LGRIP30 irrigated value = 2 and JRC seasonality ≥ 1). We ultimately classified
155 145 samples of agricultural ecosystems into 32 dry land, 72 irrigated, and 41
156 submerged paddy sites.

157 We calculated the FNC and BNC based on amino sugar concentrations following
158 widely used conversion factors, correcting total GluN for its bacterial share using
159 MurA:

$$160 \quad FNC = \left(\frac{GluN}{179.17} - 2 \times \frac{MurA}{251.23} \right) \times 179.17 \times 9 \quad \text{Equation 1}$$

161 where 9 (unitless) is the conversion factor from GluN to FNC. To estimate
162 fungal-derived GluN, we subtracted the bacterial share of GluN assuming an
163 empirical GluN:MurA molar ratio of 2:1 for bacterial residues. 179.17 and 251.23 are
164 the molecular weights of GluN and MurA, respectively. And their units are all g/mol.
165 The unit of FNC is mg/kg.

$$166 \quad BNC = MurA \times 45 \quad \text{Equation 2}$$

167 where 45 (unitless) is the conversion factor from MurA to BNC. The unit of BNC
168 is mg/kg.

169 Additional information including site geographic location (latitude and longitude),
170 topographical condition (elevation), climatic factors (mean annual temperature [MAT],
171 and mean annual precipitation [MAP]), soil physicochemical properties (pH, SOC,
172 total nitrogen [TN], clay content, and soil temperature), and biotic (microbial and
173 plant) factors were recorded. Specifically, biotic factors included microbial biomass
174 carbon (MBC), microbial biomass nitrogen (MBN), MBC/MBN, net primary
175 production (NPP), and belowground biomass C density (BGBC). The data of
176 topographical condition (elevation) was classified as geographical factor in this study.
177 When MAT and MAP were unavailable in the original articles, we extracted them
178 from the global climate layers of WorldClim (<http://www.worldclim.org/>) with a grid
179 precision of 30×30 arc sec according to geographic location. Missing elevation data
180 were extracted using the *elevatr* package v.0.4.2 (Hollister, 2021) in the R
181 environment. We acquired the data on annual mean soil temperature from the study of
182 Lembrechts et al. (2022), while other absent soil physicochemical data were extracted
183 from the Harmonized World Soil Database
184 (<https://www.fao.org/soils-portal/data-hub/soil-maps-and-databases/harmonized-world-soil-database-v12/en/>)
185 and SoilGrids 2.0 (Poggio et al., 2021;
186 <https://www.soilgrids.org/>) using ArcGIS 10.3. We supplemented missing climatic and
187 soil variables using high-resolution, globally interpolated datasets to ensure consistent
188 spatial coverage across all sites. After retrieving missing value from gridded data, we
189 typically calibrate them against field-reported values via a field-anchored bias
190 correction (i.e., a site- or region-specific “delta” adjustment) to minimize errors
191 introduced by gridded data. While the use of such data introduces inherent
192 uncertainties, these databases are widely adopted in global-scale ecological analyses
193 and provide the most feasible approach for a unified assessment. In addition, the data
194 on NPP and BGBC were acquired from the studies of Zhao and Running (2010) and
195 Spawn et al. (2020), respectively. Missing MBC and MBN data were acquired using a

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198 global database with a high resolution of 30×30 arc sec (Wang et al., 2022).

199

200 2.2 Statistical analysis

201 All the statistical analyses were performed using R v4.1.3 (R Core Team, 2021).

202 Initially, the Shapiro–Wilk test was employed to assess the normality of our data,

203 followed by the application of Levene's test to evaluate the homogeneity of variances

204 across different groups. To detect the significant differences in the contributions of

205 FNC and BNC to SOC, and the FNC/BNC ratio between agricultural and natural

206 ecosystems, as well as between forest and grassland ecosystems, the Wilcoxon rank

207 sum test was conducted. Kruskal–Wallis and Dunn's post hoc tests were used to

208 assess the significant differences of the contributions of FNC and BNC to SOC, and

209 the FNC/BNC ratio among dry land, irrigated cropland, and submerged paddy. We

210 used Spearman's rank correlation coefficient to explore the connections between the

211 16 variables considered in this study, including geographical and climatic factors, soil

212 physicochemical properties, and biotic factors. Since there was a strong positive

213 correlation between MAT and soil temperature (Figure S1), soil temperature was

214 excluded from our subsequent analyses. Linear regressions between different factors

215 and the contributions of FNC and BNC to SOC and the FNC/BNC ratio were

216 performed. Dots and smoothing curves were drawn using the *geom_point* and

217 *geom_smooth* functions, respectively, in the *ggplot2* package v.3.4.0 (Wickham,

218 2016).

219 Variation partitioning analysis was conducted using the *vegan* package v.2.5.7

220 (Oksanen et al., 2020) to evaluate the effects of four types of factors on the

221 contributions of FNC and BNC to SOC and the FNC/BNC ratio in agricultural and

222 natural ecosystems at global scale. We used a variance inflation factor threshold of 3.3

223 to eliminate those variables that were strongly correlated and avoid multicollinearity

224 (Figure S2; Kock, 2015; Fanin et al., 2020). Following factor selection, boosted

225 regression trees (BRTs) were used to partition independent influences of geographical

226 (elevation) and climatic (MAT and MAP) factors, soil physicochemical properties (pH,

227 clay, C/N, and SOC), and biotic factors (NPP, BGBC, MBC, and MBC/MBN) on the

228 contributions of FNC and BNC to SOC and the FNC/BNC ratio with the *gbm* package

229 v.2.1.8.1 (Greenwell et al., 2022).

230 Utilizing the selected factors, we performed structural equation models (SEMs)

231 to quantify the effects (direct, indirect and both) of four types of factors on the

232 contributions of FNC and BNC to SOC and the FNC/BNC ratio using *lavaan* package

233 v.0.6.19 (Rosseel, 2012). According to the previously reported potential causal

234 relationships between explanatory and response variables (Wang et al., 2021a, 2021b;

235 Li et al., 2024), we established the *priori* structural equation models for agricultural

236 and natural ecosystems, respectively (Figure S3). The SEMs were fitted via maximum

237 likelihood estimation, with non-significant paths iteratively pruned through stepwise

238 exclusion, followed by model evaluation using modification indices and

239 goodness-of-fit criteria. The fit indices included degrees of freedom (df), chi-square

240 (χ^2 , $0 \leq \chi^2/df \leq 2$), comparative fit index (CFI > 0.9), and root mean square error of

241 approximation (RMSEA < 0.08), which were used to assess the adequacy of the SEM.

242 Map, box, bar, bubble, and lollipop charts were plotted with the *ggplot2* package
243 v.3.4.0 (Wickham, 2016). To enhance map visualization, the *ggnewscale* package
244 v.0.4.8 (Campitelli, 2022) was necessary alongside the *ggplot2* package v.3.4.0
245 (Wickham, 2016). Similarly, the *ggpubr* package v.0.5.0 (Kassambara, 2022) was an
246 additional necessity when creating lollipop charts.

247

248 **3 Results**

249 3.1 Fungal and bacterial necromass contribution to SOC in agricultural and natural 250 ecosystems

251 Our analysis revealed statistically significant disparities in the contributions of FNC
252 and BNC to SOC in agricultural and natural ecosystems at the global scale ($P < 0.05$;
253 Figure 2a, b). Notably, the average contributions of FNC and BNC to SOC were
254 substantially higher in agricultural ecosystems than in natural ecosystems ($P < 0.001$;
255 Figures 2a, b). For FNC, the average contribution was 34.39% in agricultural
256 ecosystems, versus 29.24% in natural ecosystems. BNC contributed an average of
257 15.65% to SOC in agricultural ecosystems, compared to 14.02% in natural
258 ecosystems (Table 1). Our results also indicated that the contributions of FNC to SOC
259 were approximately twice those of BNC in agricultural and natural ecosystems (Table
260 1).

261 The contributions of FNC and BNC to SOC indicated no significant difference
262 between dry land and irrigated cropland ($P > 0.05$), whereas both differed
263 significantly from submerged paddy ($P < 0.05$; Figure S4a, b). In detail, across dry
264 land, irrigated cropland, and submerged paddy, the mean contributions of FNC to
265 SOC were 37.77%, 35.35%, and 22.82%, respectively, whereas those of BNC were
266 17.34%, 15.95%, and 10.55% (Table 1). Moreover, there were no significant
267 differences in the contributions of FNC and BNC to SOC between forest and
268 grassland ecosystems ($P > 0.05$; Figure S5). Specifically, FNC contributed, on
269 average, 29.11% to SOC in forests and 26.75% in grasslands, while BNC contributed
270 13.48% in forests and 14.34% in grasslands (Table 1).

271

272 3.2 Ratios of fungal and bacterial necromass in agricultural and natural ecosystems

273 Our results indicated that, at the global scale, the soil FNC/BNC ratio differs
274 significantly between agricultural and natural ecosystems ($P < 0.05$; Fig. 2c), with a
275 higher ratio in natural ecosystems (3.22) than in agricultural ecosystems (2.61; Table
276 1). The FNC/BNC ratio did not differ significantly among dry land, irrigated cropland,
277 and submerged paddy ($P > 0.05$; Figure S4c), with average FNC/BNC ratios of 2.87,
278 2.51, and 2.62, respectively (Table 1). Similarly, there was no significant difference in
279 the FNC/BNC ratio between forest and grassland ecosystems ($P > 0.05$; Figure S5),
280 and the average FNC/BNC ratios for forests and grasslands were 2.80 and 3.58 (Table
281 1), respectively.

282

283 3.3 Associations of abiotic and biotic factors with microbial necromass parameters

284 Soil physicochemical factors were the most important influence on the contributions
285 of FNC and BNC to SOC across both ecosystem types (Figures 3a–d, 4a–d).

286 Specifically, they explained 16% and 17% of the variance in the contributions of FNC
287 and BNC to SOC in agricultural ecosystems, respectively (Figures 3a, c), and 20%
288 and 24% in natural ecosystems (Figures 3b, d). BRTs corroborated this pattern, with
289 soil physicochemical factors showing the highest relative influence (51% for FNC,
290 and 44% for BNC) in agricultural systems and 44% in natural systems (Figures 4a–d).
291 All BRT models were significant ($P < 0.001$), with explained variance 36–66%. While
292 soil factors dominated overall, responses to individual variables differed between
293 ecosystems. In detail, in agricultural systems, the C/N ratio ranked third for FNC after
294 clay and SOC (Figure 4a), whereas C/N was the top predictor for FNC in natural
295 systems and for BNC in both ecosystems (Figures 4b–d). Consistently, linear models
296 showed declines in the contributions of FNC and BNC with increasing C/N in both
297 ecosystems (Figures S6g, S7g). SEMs yielded convergent results, indicating both
298 direct and indirect pathways (Figures 5a–d, 6a–d). Notably, the direct and total effects
299 of soil physicochemical factors on FNC were negative in agricultural but positive in
300 natural ecosystems (Figures 5a, b, 6a, b), whereas the effects on BNC were negative
301 in both ecosystem types (Figures 5c, d, 6c, d).

302 Our results indicated that geographical factors were the most important
303 contributors to explain the FNC/BNC ratio in both agricultural and natural ecosystems,
304 accounting for 21% and 10% of the explained variance in the FNC/BNC ratio,
305 respectively (Figures 3e, f). The results of the BRTs suggested that geographical
306 factors played a similar role in explaining the FNC/BNC ratio (Figures 4e, f). In the
307 BRT models, geographical factors emerged as the primary influencers of the
308 FNC/BNC ratio in agricultural and natural ecosystems, accounting for 32% and 44%
309 of the variance in each case, respectively (Figures 4e, f). To be more specific,
310 elevation was the most significant geographical factors influencing the FNC/BNC
311 ratio in both ecosystems (Figures 4e, f). Moreover, the FNC/BNC ratio in agricultural
312 and natural ecosystems show significantly increased with an increase elevation
313 (Figure S8a). The results of SEMs also indicated that geographical factors were the
314 most influential factors for the FNC/BNC ratio in agricultural and natural ecosystems,
315 exerting both direct and indirect effects on this ratio (Figures 5e, 6e), with the
316 standardized total effect being positive (Figures 5f, 6f).

317

318 **4 Discussion**

319 **4.1 Fungal necromass contributes two times more to SOC than bacterial necromass**

320 Our results show that in agricultural ecosystems, FNC/SOC ranged from 0.09% to
321 97.53% (mean \pm SE: $34.39 \pm 0.67\%$), and BNC/SOC ranged from 0.81% to 65%
322 ($15.65 \pm 0.33\%$). In natural ecosystems, FNC/SOC ranged from 0.92% to 96.29%
323 ($29.24 \pm 0.51\%$), and BNC/SOC ranged from 0.25% to 89.45% ($14.02 \pm 0.36\%$)
324 (Table 1). The FNC/BNC ratio ranged from 0.02 to 12.74 (2.61 ± 0.06) in agricultural
325 ecosystems and from 0.12 to 44.24 (3.22 ± 0.11) in natural ecosystems (Table 1).
326 Despite substantial variability at the individual sample level, the mean contribution of
327 FNC was approximately twice that of BNC in both ecosystem types. Moreover, the
328 mean FNC/BNC ratio was significantly higher in natural ecosystems than in
329 agricultural ecosystems ($P < 0.05$; Figure 2). Although this general pattern has been

330 reported in previous studies (Liang et al., 2019; Wang et al., 2021a; Zhang et al., 2023;
331 Ding et al., 2024), the systematic differences in the magnitude of these contributions
332 between agricultural and natural ecosystems—and their underlying drivers—have
333 remained poorly understood. Our study not only confirms the broad pattern but also
334 elucidates these ecosystem-level disparities and their environmental determinants.
335 Consistent with our finding that the contribution of fungal necromass carbon (FNC) to
336 SOC exceeded that of bacterial necromass carbon (BNC) in both ecosystem types
337 (Table 1), the predominance of fungal necromass may be attributed to its more
338 recalcitrant cell wall composition (e.g., chitin) and slower decomposition rate (Wang
339 et al., 2021a). Our BRT and SEM analyses further identified soil clay content and C/N
340 ratio as key drivers of FNC accumulation (Figs. 4a, 5a), reinforcing the importance of
341 organo-mineral associations in the stabilization of fungal-derived carbon.

342 Additionally, our study reveals previously unreported disparities between
343 ecosystem types, for example, the contributions of both fungal and bacterial
344 necromass carbon (FNC and BNC) to SOC were significantly higher in agricultural
345 ecosystems, while the FNC/BNC ratio was substantially elevated in natural
346 ecosystems. The higher contributions of FNC and BNC to SOC in agricultural
347 ecosystems may be attributable to two main factors. First, natural ecosystems
348 typically receive larger and more heterogeneous plant-derived carbon inputs than
349 agricultural systems. These inputs expand the plant-derived SOC pool and can dilute
350 the relative contribution of microbial necromass to SOC, thereby resulting in a lower
351 perceived contribution of microbial necromass in natural ecosystems (Angst et al.,
352 2021; Kleber et al., 2021). Second, the significantly lower soil C/N ratio in
353 agricultural ecosystems (10.78) compared to natural ecosystems (27.44) reflects
354 relative nitrogen enrichment, largely resulting from anthropogenic fertilization
355 (Castellano et al., 2015; Chen et al., 2020). This nitrogen-rich environment can
356 enhance microbial carbon use efficiency and alleviate nutrient limitation, thereby
357 promoting the production and accumulation of microbial necromass (Liang et al.,
358 2017). Supporting this mechanism, we found that the contributions of both FNC and
359 BNC to SOC decreased significantly with increasing soil C/N ratio in both
360 agricultural ecosystems (FNC/SOC: $R = -0.27$, $P < 0.001$; BNC/SOC: $R = -0.29$, $P <$
361 0.001) and natural ecosystems (FNC/SOC: $R = -0.17$, $P < 0.001$; BNC/SOC: $R =$
362 -0.35 , $P < 0.001$; Figures S6g, S7g). These results further underscore that a lower soil
363 C/N ratio—often indicative of higher nitrogen availability—is a key driver of
364 microbial necromass accumulation. It should be noted that although in situ plant
365 residues in agricultural systems (e.g., cereal straw) may have high C/N ratios, the
366 overall soil C/N ratio is reduced by management practices such as mineral fertilization
367 and the incorporation of low C/N organic amendments.

368 Furthermore, nutrient-rich conditions prevalent in agricultural systems (e.g., due
369 to fertilization) often select for bacterial-dominated communities, as many bacteria
370 exhibit *r*-strategist traits that support rapid growth under high resource availability. In
371 contrast, natural ecosystems—characterized by lower nutrient availability and greater
372 resource heterogeneity—tend to favor fungal dominance, since fungi often function as
373 *K*-strategists with higher efficiency in decomposing complex organic matter under

374 resource-limited conditions (Strickland & Rousk, 2010; Yu et al., 2022). This shift in
375 microbial community composition is observed in our results, which show a
376 significantly higher FNC/BNC ratio in natural ecosystems across our global dataset
377 (Figure 2c, Table 1). A high FNC/BNC ratio signifies a fungal-dominated
378 decomposition pathway. Fungal necromass—rich in recalcitrant compounds such as
379 chitin—is more resistant to decay, and fungal hyphae play a key role in the formation
380 of stable soil aggregates that physically protect organic matter from degradation
381 (Lenardon et al., 2007). This pathway promotes the formation of stable, long-turnover
382 SOC pools essential for long-term carbon sequestration (Six et al., 2006; Lehmann et
383 al., 2020). Furthermore, fungi generally exhibit higher carbon use efficiency than
384 bacteria, meaning a larger proportion of assimilated carbon is allocated to biomass
385 production (and subsequently necromass) rather than being respired as CO₂ (Wang &
386 Kuzyakov, 2024). Thus, the fungal-driven pathway characteristic of natural
387 ecosystems represents a highly efficient conversion of plant litter into persistent soil
388 organic matter (Kallenbach et al., 2016; Malik et al., 2016). Conversely, the lower
389 FNC/BNC ratio observed in agricultural ecosystems reflects a bacterial-dominated
390 pathway, accelerated by practices such as tillage and nutrient amendments. This
391 pathway is associated with faster carbon cycling and greater carbon loss through
392 respiration. Although microbial necromass can accumulate under these
393 conditions—sometimes contributing more significantly to a reduced total SOC
394 pool—the resulting carbon is often less stabilized (Zhou et al., 2023). Therefore, the
395 FNC/BNC ratio serves not merely as a descriptive metric, but as a functional
396 biomarker that elucidates fundamental differences in the stability and persistence of
397 SOM between managed agricultural systems and natural ecosystems.

398 Notably, as major components of agricultural ecosystems, both dryland and
399 irrigated croplands exhibited significantly greater contributions of FNC and BNC to
400 SOC than submerged paddy soils, although the FNC/BNC ratio did not differ
401 significantly among these three systems (Figure S4). This pattern may reflect similar
402 aeration regimes in dryland and irrigated systems (predominantly oxygenated),
403 leading to comparable decomposition–transformation–mineral association pathways
404 and, thus, similar net contributions of fungal and bacterial residues to SOC
405 (Ghezzehei et al., 2019). By contrast, persistent or periodic flooding in paddy soils
406 induces anoxia, suppresses aerobic decomposition, and shifts metabolic pathways
407 (e.g., denitrification and methanogenesis), potentially suppressing fungal activity or
408 dominance and altering the relative accumulation and turnover of fungal and bacterial
409 necromass (Qiu et al., 2017), resulting in contributions that differ significantly
410 from—and are lower than—those in the other two systems. Flooding can suppress
411 fungi yet also enhance the joint retention of both fungal and bacterial necromass via
412 slower decomposition and mineral protection, yielding unchanged ratios but altered
413 totals or compositional pathways (Chen et al., 2021; Gao et al., 2024).

414
415 4.2 Driving factors of the change in fungal and bacterial necromass contribution to
416 SOC and their ratio

417 Deng and Liang (2022) suggested that the potential contribution of microbial

418 necromass to the SOC pool was governed by the C/N ratio. This finding was
419 confirmed by our results (Figures 4b–d). As elaborated in *Section 4.1*, high N
420 availability (i.e., low soil C/N ratio) promotes the production and accumulation of
421 microbial necromass (Wu et al., 2025). Consequently, the contributions of both FNC
422 and BNC to SOC decreased with increasing soil C/N ratio (Figures S6g, S7g). In
423 agricultural ecosystems, high soil N levels primarily result from fertilization (Chen et
424 al., 2020). In contrast, natural ecosystems experience minimal anthropogenic
425 disturbance, N often acts as the key limiting factor for microbial activity (Elser et al.,
426 2007). Under N-limited conditions, microbes (both fungi and bacteria) allocate more
427 energy and C resources to the synthesis of N-acquiring enzymes (e.g., proteases and
428 chitinases). This shift in metabolic strategy reduces the C allocated to biomass
429 synthesis, thereby diminishing the amount of C ultimately converted into microbial
430 necromass (Mooshammer et al., 2014; Liu et al., 2024). Thus, although microbial
431 community composition differs between natural and agricultural ecosystems, the
432 regulatory role of soil C/N ratio in shaping their structure and function remains
433 consistent (Han et al., 2024). In our study, soil clay content was identified as the
434 predominant factor governing the contribution of FNC to SOC in agricultural
435 ecosystems (Figure 4a), with this contribution increasing concomitantly with clay
436 content (Figure S6d). This suggests that soils with higher clay and silt contents
437 generally accumulate greater amounts of microbial residues, particularly those derived
438 from fungi, which can be attributed to the promotion of stable organo-mineral
439 complex formation by abundant fine soil particles (Six et al., 2006 and Liang et al.,
440 2017). Furthermore, although agricultural management practices often disturb soil
441 structure, they simultaneously enhance clay enrichment and aggregate formation,
442 thereby providing effective physical protection for the long-term stabilization of
443 fungal-derived C (Chen et al., 2020; Mou et al., 2021; Zhou et al., 2023).

444 On the contrary, geographical factor (elevation) was identified as the most
445 influential predictor of the FNC/BNC ratio in both agricultural and natural ecosystems
446 (Figures 4e, f, 5f, 6f), with the ratio increasing significantly with elevation (Figure
447 S8a). Increasing elevation typically leads to decreased temperature, and increased
448 precipitation (Körner, 2007), conditions favoring fungi over bacteria due to higher
449 enzymatic capabilities and resource-use efficiency of fungi under the environments
450 (Chen et al., 2020; Yu et al., 2022; Zhang et al., 2025). High elevation also results in
451 slower soil development, which can reduce the availability of soil nutrients
452 (Guerrero-Ramírez et al., 2020). This in turn increases environmental stress and
453 restricts bacterial activity, thereby favoring the accumulation and conversion of fungal
454 residues into necromass (Li et al., 2024). Our study further demonstrated that although
455 elevation had a direct effect on the FNC/BNC ratio, it also indirectly influenced the
456 ratio by modulating climatic factors, soil physicochemical properties, and biological
457 factors (Figures 5e, 6e). This may explain why elevation is always integrate other
458 environmental factor effects in the studies of MNC (Cui et al., 2023; Zhang et al.,
459 2025).

460
461 4.3 Limitations and uncertainties

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462 Although the present study provides important insights on global patterns and drivers
463 of soil microbial necromass in agricultural and natural ecosystems, we must clarify
464 three limitations. First, the limited data available on microbial characteristics, such as
465 microbial community composition, enzymatic activities, and the content of soil
466 aggregates and minerals hinder exploration of the drivers of soil microbial necromass.
467 Second, it is undeniable that our dataset is unevenly distributed, primarily
468 concentrating on the Northern Hemisphere, with sparse or nearly no data from other
469 regions such as Africa, South America, and Australia (Figure 1). Additionally, the
470 natural ecosystems in this study were limited to forests and grasslands, excluding
471 other natural habitats such as wetlands and deserts. The uneven distribution of data
472 may reduce the universality of MNC as a key driver of soil carbon pools in global
473 terrestrial ecosystems. Furthermore, the compiled studies employed varied
474 methodologies regarding sampling time, depth, and laboratory protocols. While such
475 heterogeneity is an inherent challenge in global meta-analyses, it likely introduces
476 additional variability and may constrain the direct comparability of certain data points.
477 Therefore, more standardized data from these important areas and biomes are clearly
478 required, and further investigation is warranted to fill the data gaps regarding the
479 contribution of MNC to SOC in terrestrial ecosystems.

480 **5 Data availability**

481 The data and R code for this manuscript are available at
482 <https://doi.org/10.6084/m9.figshare.28827383> (Lu, 2025).

484 **6 Conclusions**

485 Our results indicate that, on average, fungal necromass carbon (FNC) contributes
486 approximately twice as much to soil organic carbon (SOC) as bacterial necromass
487 carbon (BNC) in both agricultural and natural ecosystems. The relative contributions
488 of FNC and BNC to SOC were found to be higher in agricultural ecosystems—an
489 effect that is mediated by differences in soil physicochemical factors. The FNC/BNC
490 ratio was significantly higher in natural ecosystems than in agricultural ecosystems,
491 albeit with a modest effect size, and was primarily driven by geographical
492 factors—particularly elevation. Our findings demonstrate that, despite considerable
493 variability among individual sampling sites, statistically significant differences exist
494 between agricultural and natural ecosystems in the contributions of fungal and
495 bacterial necromass carbon (FNC and BNC) to soil organic carbon (SOC), as well as
496 in the FNC/BNC ratio, at a global scale. These results underscore a potential
497 fundamental divergence in the pathways and mechanisms of carbon turnover and
498 stabilization between these two broad ecosystem types. These insights provide novel
499 evidence that ecosystem management type (agricultural versus natural) is a key
500 determinant of the pathways through which microbial necromass contributes to the
501 global soil organic carbon (SOC) pool. Future studies that integrate microbial
502 community composition with necromass dynamics across a broader range of biomes
503 will be essential to predict ecosystem-specific responses of this critical carbon pool to
504 global change.
505

507

508 **Author contributions**

509 JL performed the data analysis and prepared the original draft. TC and MDB
510 contributed to manuscript review and editing. WL and HS contributed to data
511 collection. YJ contributed to data analysis. ZW supervised the project and contributed
512 to the original draft.

513

514 **Competing interests**

515 The contact author has declared that none of the authors has any competing interests.

516

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532

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832 **Table 1. Summary of the contributions of fungal necromass carbon (FNC) and**
833 **bacterial necromass carbon (BNC) to SOC and the FNC/BNC ratio in**
834 **agricultural and natural ecosystems at the global scale investigated in this study.**

Ecosystem	FNC/SOC (%)		BNC/SOC (%)		FNC/BNC	
	Range	Mean± SE	Range	Mean± SE	Range	Mean± SE
Natural ecosystem^{&} (<i>N</i> = 341)	0.92– 96.29	29.24 ± 0.51 b*	0.25– 89.45	14.02 ± 0.36 b	0.12– 44.24	3.22 ± 0.11 a
<i>Forest</i> (<i>N</i> = 195)	0.92– 96.29	29.11 ± 0.63 A [#]	0.94– 96.47	13.48 ± 0.43 A	0.22– 11.56	2.80 ± 0.07 A
<i>Grassland</i> (<i>N</i> = 146)	0.96– 93.89	26.75 ± 0.74 A	0.25– 89.45	14.34 ± 0.60 A	0.05– 44.24	3.58 ± 0.22 A
Agricultural ecosystem[%] (<i>N</i> = 145)	0.09– 97.53	34.39 ± 0.67 a	0.81– 65.00	15.65 ± 0.33 a	0.02– 12.74	2.61 ± 0.06 b
<i>Dry land</i> (<i>N</i> = 32)	3.01– 96.81	37.77 ± 1.15 A [#]	0.81– 65.00	17.34 ± 0.65 A	0.13– 9.12	2.87 ± 0.12 A
<i>Irrigated cropland</i> (<i>N</i> = 72)	0.09– 97.25	35.35 ± 0.73 A	1.18– 62.47	15.95 ± 0.38 A	0.02– 12.74	2.51 ± 0.06 A
<i>Submerged paddy</i> (<i>N</i> = 41)	4.96– 97.53	22.82 ± 1.55 B	1.48– 30.97	10.55 ± 0.66 B	0.31– 10.40	2.62 ± 0.16 A

835 Note: *N* refers to the number of study sites;

836 [&]Natural ecosystem includes forest and grassland;

837 [%]Agricultural ecosystem includes dry land, irrigated cropland, and submerged paddy;

838 ^{*}Within the same column, values with different lowercase letters indicate a significant
839 difference in the same variable between agricultural and natural ecosystems
840 (Wilcoxon rank sum test; *P* < 0.05);

841 [#]Within the same column, values with different capital letters indicate a significant
842 difference in the same variable between forests and grasslands (Wilcoxon rank sum
843 test), as well as between dry land, irrigated cropland, and submerged paddy (Kruskal–
844 Wallis and Dunn's post hoc tests; *P* < 0.05).

845 **Figure legends**

846 **Figure 1. Global distribution of the sites used in this study.** Ecosystem types are
847 distinguished by distinct shapes and colors, with the numbers in parentheses
848 indicating the number of study sites for each ecosystem type.

849 **Figure 2. Comparison of the contributions of MNC to SOC and their ratio in**
850 **agricultural and natural ecosystems.** Colors indicate different ecosystems types.
851 Significance levels: *** $P < 0.001$ and * $P < 0.05$.

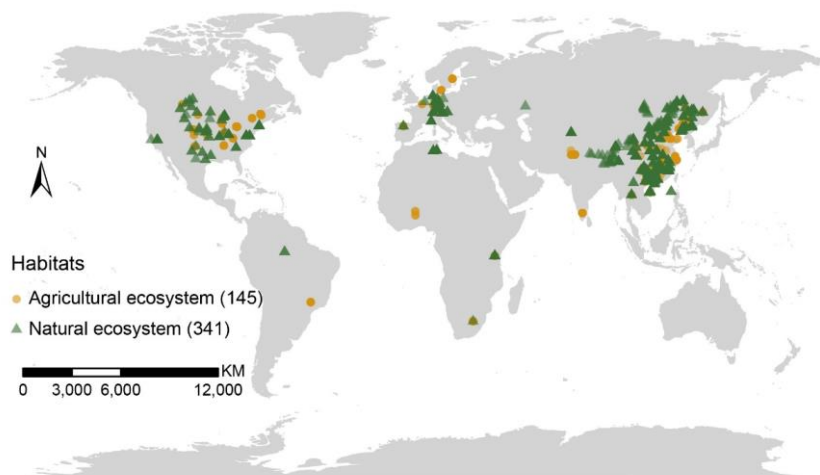
852 **Figure 3. Variations in the contributions of MNC to SOC and their ratio**
853 **explained by four types of factors in agricultural and natural ecosystems.** Colors
854 indicate different types of factors.

855 **Figure 4. Relative influence of different factors on the contributions of MNC to**
856 **SOC and their ratio in agricultural and natural ecosystems.** MAT, mean annual
857 temperature; MAP, mean annual precipitation; MBC, microbial biomass carbon; SOC,
858 soil organic carbon; C/N, the ratio of SOC to total nitrogen (TN); MBC/MBN, the
859 ratio of MBC to microbial biomass nitrogen (MBN); NPP, net primary production;
860 BGBC, belowground biomass carbon density. Colors indicate different types of
861 factors.

862 **Figure 5. The influence pathways of four types of factors on the contributions of**
863 **MNC to SOC and their ratio in agricultural ecosystems.** Direct and indirect effects
864 (a, c, e) and the standardized total effects (b, d, f) of different factors on the
865 contributions of MNC to SOC and their ratio of agricultural ecosystems are shown.
866 Standardized path coefficients representing the effect sizes of potential causal factors
867 are indicated by numbers adjacent to arrows. The width of arrows is proportional to
868 the potential causal effect between variables. The red arrows indicate positive effects,
869 and the blue arrows indicate negative effects. The numbers adjacent to boxes of
870 response variables denote the explained variance (R^2). Right-angled rectangles denote
871 single variables, whereas rounded rectangles represent composite variables. Colors
872 indicate different types of factors. Significance levels: *** $P < 0.001$, ** $P < 0.01$ and
873 * $P < 0.05$. The *priori* models are shown in Figure S3.

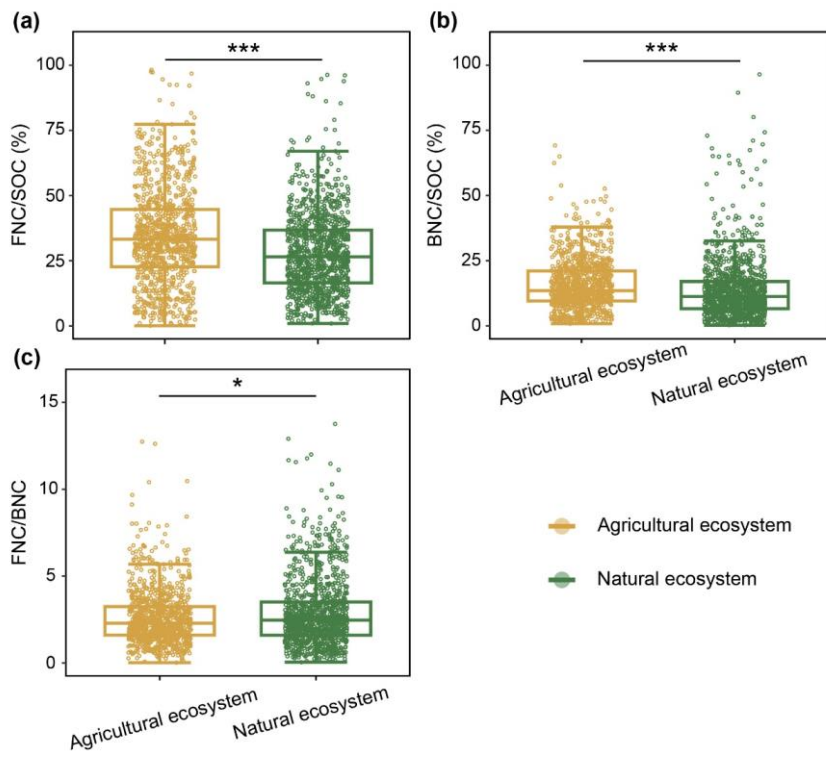
874 **Figure 6. The influence pathways of four types of factors on the contributions of**
875 **MNC to SOC and their ratio in natural ecosystems.** Direct and indirect effects (a, c,
876 e) and the standardized total effects (b, d, f) of different factors on the contributions of
877 MNC to SOC and their ratio of natural ecosystems are shown. Standardized path
878 coefficients representing the effect sizes of potential causal factors are indicated by
879 numbers adjacent to arrows. The width of arrows is proportional to the potential
880 causal effect between variables. The red arrows indicate positive effects, and the blue
881 arrows indicate negative effects. The numbers adjacent to boxes of response variables
882 denote the explained variance (R^2). Right-angled rectangles denote single variables,
883 whereas rounded rectangles represent composite variables. Colors indicate different
884 types of factors. Significance levels: *** $P < 0.001$, ** $P < 0.01$ and * $P < 0.05$. The
885 *priori* models are shown in Figure S3.

886 **Figure 1.**



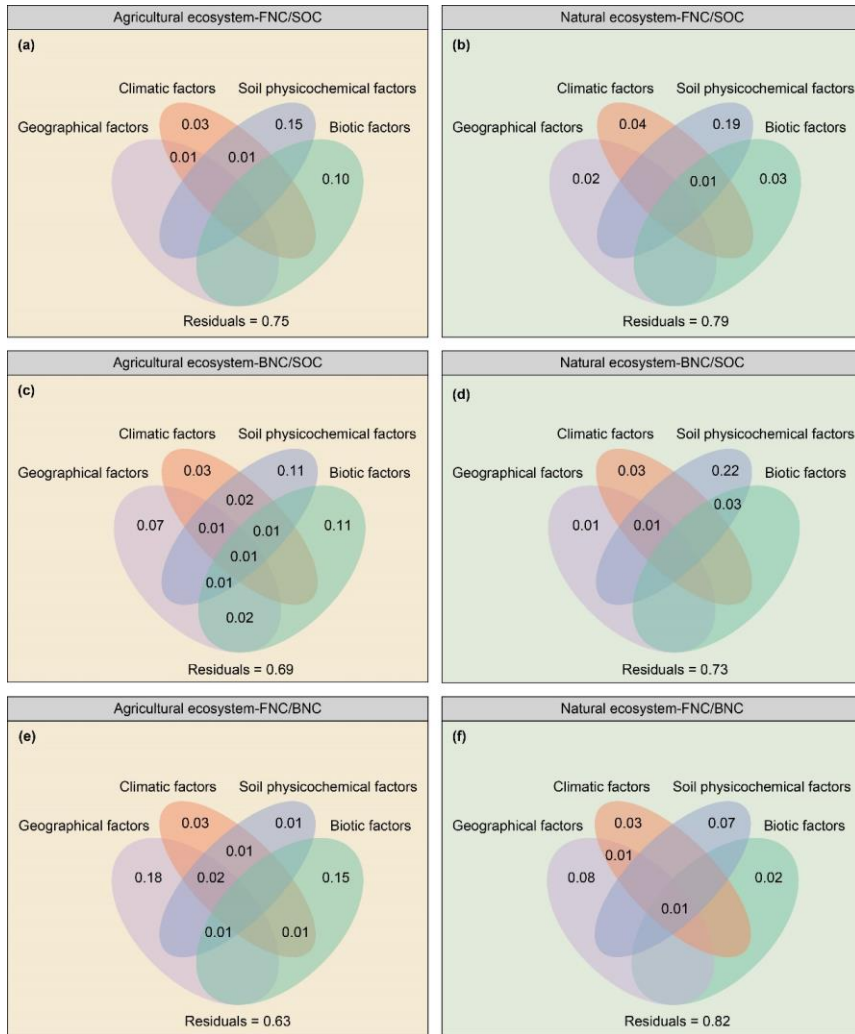
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888 **Figure 2.**



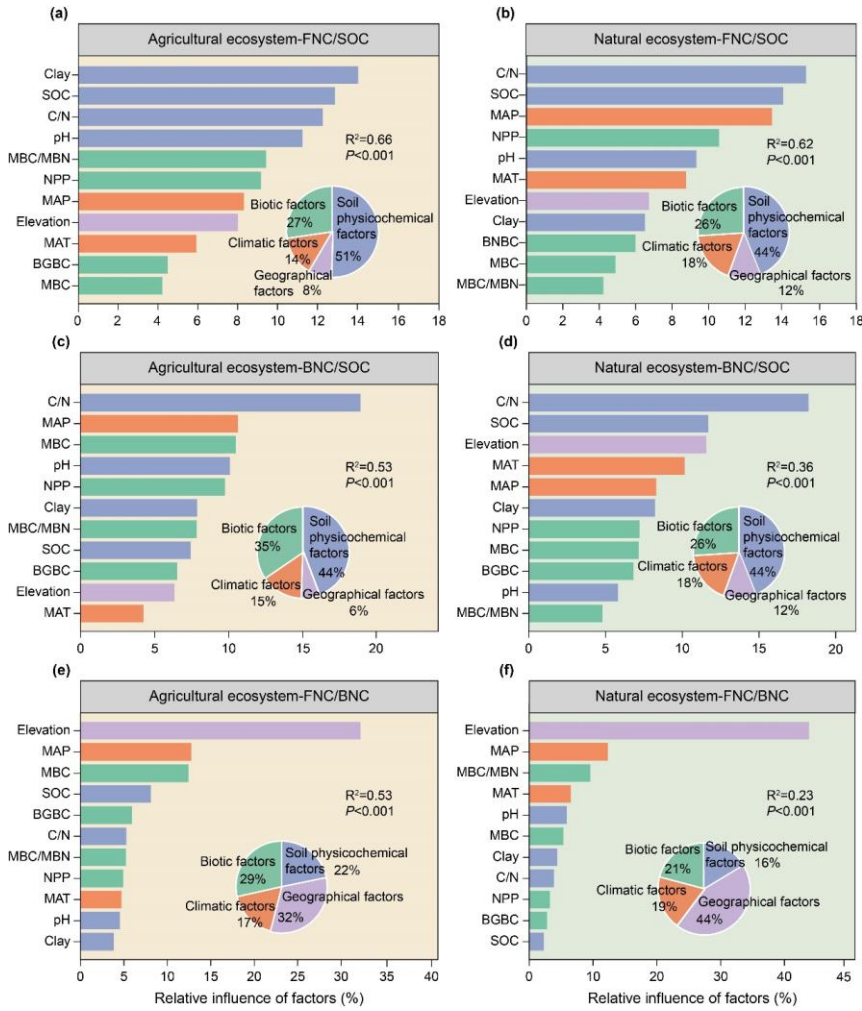
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890 **Figure 3.**



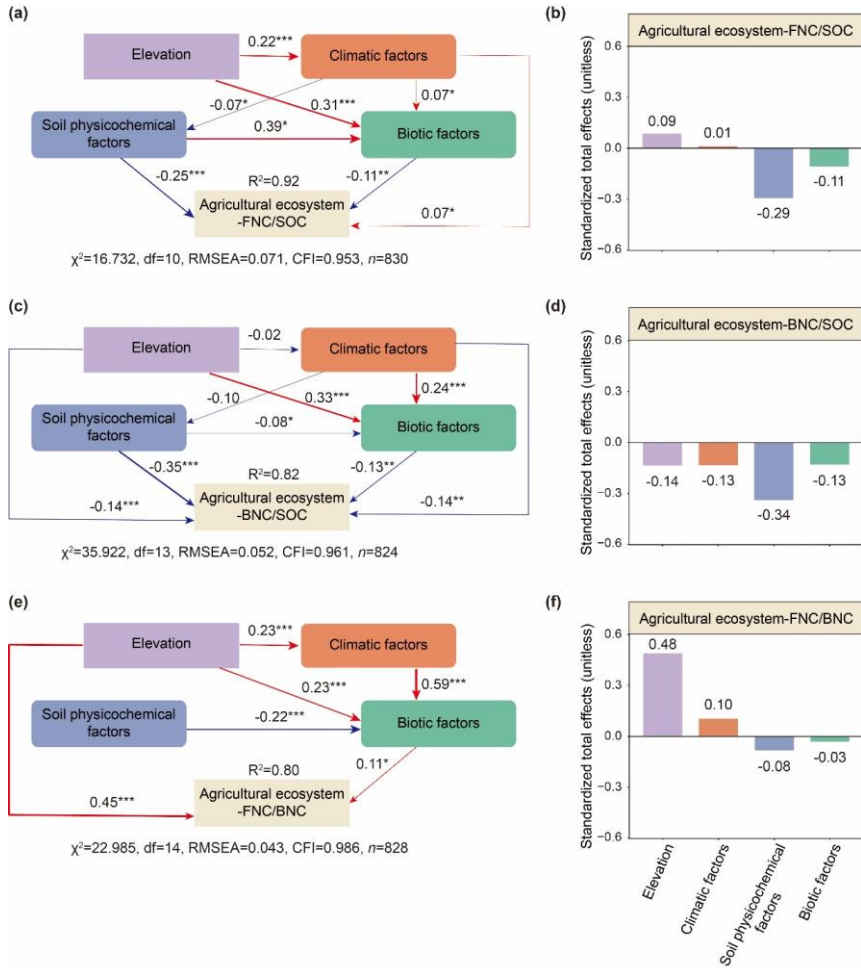
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892 **Figure 4.**



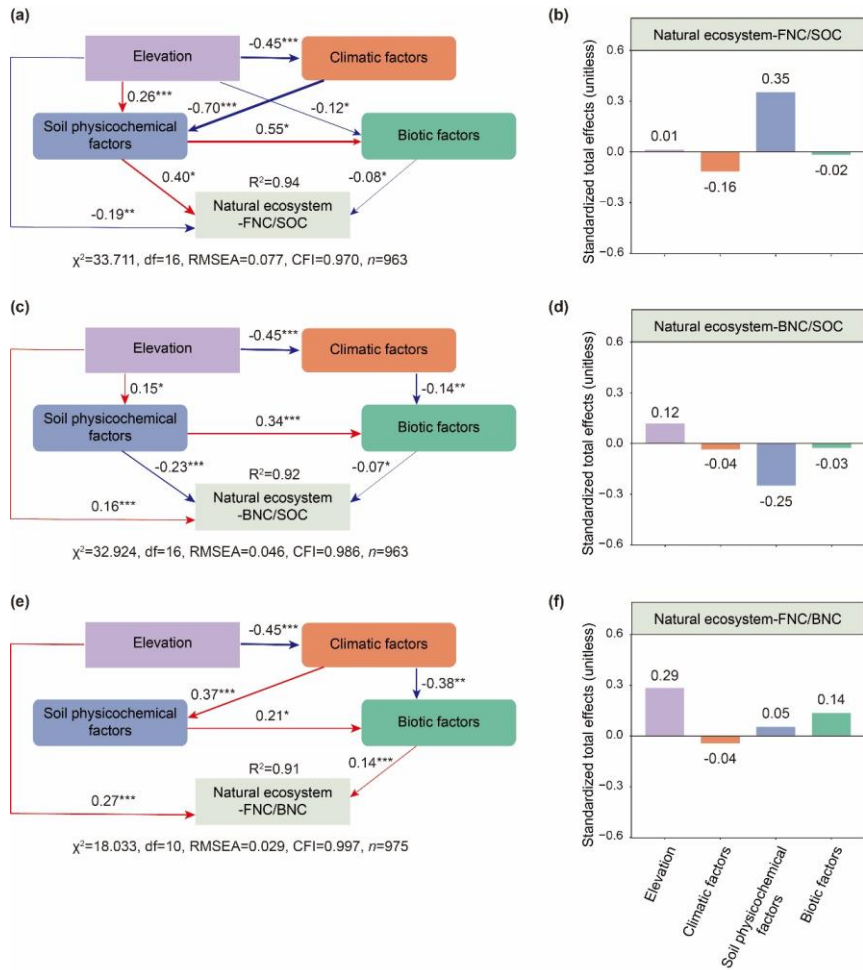
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894 **Figure 5.**



895

896 **Figure 6.**



897