



# Spatiotemporal variability of pH and carbonate parameters on the Canadian Atlantic Continental Shelf between 2014 and 2020

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**Abstract.** The Atlantic Zone Monitoring Program (AZMP) was established by Fisheries and Oceans Canada (DFO) in 1998 with the aim of monitoring physical and biological ocean conditions in Atlantic Canada in support of fisheries management. Since 2014, at least two of the carbonate parameters (pH, Total Alkalinity - TA, Dissolved Inorganic Carbon - DIC) have also been systematically measured as part of the AZMP, enabling the calculation of derived parameters (e.g., carbonate saturation states -  $\Omega$ , partial pressure of  $\text{CO}_2$  -  $p\text{CO}_2$ , etc.). The present study gives an overview of the spatiotemporal variability of these parameters between 2014 and 2020. Results show that the variability of carbonate parameters reflects changes in both physical (e.g., temperature, salinity) and biological (e.g., plankton photosynthesis and respiration) parameters. For example, most of the region undergoes a seasonal warming and freshening. While the former will tend to increase  $\Omega$ , the latter will decrease both TA and  $\Omega$ . Spring and summer plankton blooms decrease DIC near the surface and then remineralize and increase DIC at depth in the fall. The lowest  $p\text{CO}_2$  values are located in the cold Coastal Labrador Current and the highest in the fresh waters of the Gulf of St. Lawrence and the St. Lawrence Estuary. The latter is also the host of the lowest pH values of the zone. Finally, most of the bottom waters of the Gulf of St. Lawrence are undersaturated with respect to aragonite ( $\Omega_{\text{arg}} < 1$ ). In addition to providing a baseline of carbonate parameters of the Atlantic Zone as a whole, this comprehensive overview is a necessary and useful contribution for the modeling community and for more in-depth studies. The full data set of measured and derived parameters is available in the Federated Research Data Repository at <https://doi.org/10.20383/102.0673>.

## 1 Introduction

The Canadian Atlantic continental shelf and slope, termed the Canadian Atlantic Zone (or simply Atlantic Zone hereafter), is at the confluence of waters of contrasting origins (Figure 1). The northward flowing warm and saline Gulf Stream, the southward flowing cold less saline Labrador Current, and the fresh outflow from the St. Lawrence River and the Gulf of St. Lawrence provide a unique range in salinities and temperatures (Figure 2a) which form oceanic fronts along the shelf-slope edge surrounding the Atlantic Zone (Belkin et al., 2009; Cyr and Larouche, 2015). These fronts are often associated with vertical nutrient transport, high productivity and ecological hot spots with a rich diversity and abundance of organisms (e.g. Lévy et al., 2012). As



a result, the Atlantic Zone supports a variety of commercially important species which amounted to nearly 4 billion dollars (\$G) in commercial landings (\$3.2G) and finfish and shellfish aquaculture (\$0.5G) in Canada in 2019 (Fisheries and Oceans Canada, 2020). These ecosystems however are becoming stressed due to changes to the physical, chemical, and biological oceanographic environment as a result industrialization, e.g., burning of fossil fuels, agriculture, deforestation (Bernier et al., 2018; Doney, 2010). Increasing atmospheric carbon dioxide (CO<sub>2</sub>) concentrations not only increases atmospheric and sea surface temperatures but alters the ocean's carbonate cycle resulting in increased ocean acidity (i.e. decreased pH). Many aquatic species within the Atlantic Zone are vulnerable to the effects of ocean acidification, which may affect the entire ecosystem (Fabry et al., 2008; Doney, 2010). Calcifying shellfish are directly impacted by ocean acidification while other shellfish and finfish can be indirectly impacted (Kroeker et al., 2013). The socioeconomic impacts of ocean acidification and climate change on fisheries (e.g., resources decline, changes in species distribution, income and cultural losses, etc.) have been assessed for various ecosystems (Cooley and Doney, 2009; Mathis et al., 2015; Wilson et al., 2020). Monitoring the physical, chemical and biological oceanographic conditions and determining a baseline is required prior to an assessment of future aquaculture and fisheries in the Atlantic Zone.

Implemented in 1998 by Fisheries and Oceans Canada (DFO), the Atlantic Zone Monitoring Program (AZMP) was designed to evaluate the physical (temperature, salinity), chemical (nutrients and dissolved oxygen concentrations), and biological (chlorophyll-a, fluorescence, plankton species assemblage and abundance) oceanographic properties of the Canadian Northwest Atlantic (Therriault et al., 1998). The AZMP characterizes the spatial and temporal (seasonal to decadal) variability of these oceanic properties. It is carried out by the four Atlantic DFO administration centers (three geographic regions): Quebec/Gulf, Maritimes and Newfoundland and Labrador (Figure 3). The AZMP annually publishes a summary of oceanographic conditions that assesses the current state of the ecosystem. It is used for stock assessments and marine resource management by DFO, and supports fundamental oceanographic research. In fall 2014, the sampling and analysis of two of the carbonate parameters, including total alkalinity (TA), total dissolved inorganic carbon (DIC), and pH were added to the AZMP mandate. The resulting dataset is the focus of the present paper.

Ocean acidification is the decrease in ocean pH (increase in H<sup>+</sup>, acidity) and carbonate ion concentration (CO<sub>3</sub><sup>2-</sup>) due to the increased uptake of anthropogenic CO<sub>2</sub>. Seawater carbonate chemistry involves the dissolution of atmospheric CO<sub>2</sub> (Eq. 1), acid/base reactions forming the inorganic carbonate (DIC) species in equation (Eq. 2), and the formation and dissolution of solid calcium carbonate (CaCO<sub>3</sub>) (Eq. 3).





The photosynthesis and respiration of organic matter is another factor that contributes to changes in the carbonate system (Eq. 4). Spring phytoplankton blooms increase surface water pH by consuming CO<sub>2</sub> in water, while at depth the remineralization of that vertically exported organic matter to DIC lowers pH.



The degree in which seawater is saturated with calcium carbonate (CaCO<sub>3</sub>) is known as the CaCO<sub>3</sub> saturation state ( $\Omega$ ). The saturation state is a function of calcium and carbonate concentrations, and its apparent solubility product ( $K_{sp}$ ) which is a function of temperature, salinity and pressure (Eq. 5).

$$\Omega = \frac{[Ca^{2+}] + [CO_3^{2-}]}{K_{sp}^*} \quad (5)$$

Seawater with an  $\Omega > 1$  is considered oversaturated which induces carbonate precipitation, while water with  $\Omega < 1$  is considered undersaturated and corrosive which promotes carbonate dissolution. The carbonate saturation state customarily decreases with depth as a result of cold temperatures, increased pressure, and respired DIC. The depth at which the water becomes undersaturated is generally referred to as *saturation horizon*. Organisms such as phytoplankton, zooplankton and invertebrates (clams, oysters, corals) that form shells and skeletons of calcium carbonate (CaCO<sub>3</sub>) may have difficulty maintaining or forming hard structures in undersaturated waters. It should be noted, however, several studies have identified higher critical thresholds ( $\Omega \sim 1.3-2$ ) for marine organisms (e.g., Ekstrom et al., 2015; Waldbusser et al., 2015; Siedlecki et al., 2021). The two most common forms of CaCO<sub>3</sub> are calcite and aragonite. Because aragonite is more soluble than calcite, the aragonite saturation horizon ( $\Omega=1$ ) is shallower, and organisms that produce aragonite may be more vulnerable to a decreasing saturation state as atmospheric CO<sub>2</sub> increases.

Freshwater influx also plays an important role on the carbonate system. The freshwater effects on  $\Omega$  is due to both changes in DIC/TA ratio in source waters, and to the decrease in Ca<sup>+</sup> concentration as a function of salinity (e.g Azetsu-Scott et al., 2014; Hunt et al., 2021). For example, the dilution of sea water decreases all carbonate parameters, except for pCO<sub>2</sub>. TA is an indicator of seawater's ability to buffer (neutralize) acids. It is a measure of the excess total proton acceptors (anions) over proton donors (acids) formed by dissociation of carbonic, boric, and other weak acids in seawater. It is expressed as mole equivalents of hydrogen ions per kilogram of seawater. Therefore, TA will decrease with freshwater influx and during carbonate precipitation (decrease in DIC), which will subsequently decrease  $\Omega$ :



## 2 Physical Oceanographic setting

The physical properties of the Atlantic Zone are dominated by large-scale interactions of Arctic, sub-polar and sub-tropical waters and fresh water influx from the St. Lawrence river. (e.g., Loder et al., 1998; Han et al., 2008; Brickman et al., 2016).



The two major circulation features are the southward flowing Labrador Current, which brings cold and less saline subpolar waters along the Newfoundland and Labrador shelf, and the northeastward flowing North Atlantic Current (NAC; the northeast extension of the Gulf Stream) which carries warm and saline sub-tropical waters along the Scotian Shelf and the Grand Banks (Figure 1). An additional input of Arctic-origin cooler and fresher waters also flows on the inshore part of the Newfoundland and Labrador Shelf, a current generally referred to as the Inshore Labrador Current, or Labrador Coastal Current (Florindo-López et al., 2020). These contrasting water masses interact on the shelf and partially enter the Gulf of St. Lawrence (GSL), a semi-enclosed basin characterized by an estuarine circulation. The bottom waters of the GSL receive Slope Water through the Laurentian Channel.

On the Scotian Shelf, Slope Waters that lie along the shelf break are a combination of cold, less saline, oxygen-rich and nutrient-poor Labrador Slope Water (LSW) from the Labrador Current, and warm and saline, nutrient-rich and oxygen-poor Warm Slope Water (WSW) derived from the NAC. The properties of the Slope Water vary depending on which component (LSW vs WSW) is dominant (Petrie and Drinkwater, 1993). As the mixture flows landward into the GSL along the bottom of the Laurentian Channel toward the Lower St. Lawrence Estuary (LSLE), oxygen is consumed and the deep water becomes hypoxic (Gilbert et al., 2005). These low oxygen concentrations have been associated with increased composition ratio of WSW relative to LSW (Gilbert et al., 2005; Jutras et al., 2020), poor ventilation (>16 years, Mucci et al., 2011) and increased oxygen demand in bottom waters because of respiration and remineralization of organic matter supplied by increased primary productivity at surface (Thibodeau et al., 2006). The Scotian Shelf Water receives subpolar water primarily from the GSL through western Cabot Strait, and a smaller portion directly from the Newfoundland Shelf by the Labrador Current across the Laurentian Channel (Dever et al., 2016).

In the summer, the water column of the Atlantic Zone is generally characterised by a three-layer system. A seasonally warmed thin and relatively fresh surface layer overlies a cold intermediate layer (CIL), which is the remnant of the previous winter cold surface layer. A warmer, more saline slope-derived water lies below the CIL. The deeper waters (>200 m) are generally limited to channels and troughs, and there are areas (e.g. southwestern Scotian Shelf) where the CIL is absent due to mixing. In the late fall and winter, the surface water cools, deepens and combines with the CIL into a two-layer system. In places, this winter mixed layer reaches the freezing point and sea ice is formed. This winter layer is at the origin of the following summer CIL (Galbraith, 2006).

The Atlantic Zone is thus characterized by a large range in both temperature (freezing point to over 25°C during the summer) and salinity (freshwater to over 37), which influences the carbonate system as discussed later. The Atlantic Zone is divided here into three regions geographically and then further using bathymetrical considerations: shelf (fresher) versus slope (more saline), surface (warm in summer) versus subsurface (cool). The different water masses of this system are loosely highlighted in a temperature vs salinity (T-S) and depth plot of the entire dataset (Figure 2a).



### 3 Sampling and Methodology

The dataset presented here consist of carbonate parameters compiled from multiple surveys from all four administrative AZMP  
115 Regions: Newfoundland and Labrador (NL), Maritimes (MAR), Québec (QC) and Gulf. The latter two sample the GSL Region  
and will be combined under this name herein (e.g., Fig. 2). The map of the Atlantic Zone in Figure 3a shows the locations of the  
standard AZMP sections, as well as two sections through the GSL established here to plot depth profiles across (TCEN; from  
the Magdalen Shallows on the southwest to the strait of Belle Isle in the northeast) and along the Laurentian Channel (LC; the  
120 the data, the Cabot Strait Line (CSL) will be associated with the GSL. Unless explicitly stated, the waters of the Upper  
St. Lawrence Estuary (USLE) and the Saguenay Fjord (Figure 3b), are not discussed in this study because their properties,  
highly influenced by freshwater input, are very different than the rest of the Atlantic Zone. These data are however included  
in the dataset that accompanies this paper. All bathymetric maps were generated using the GEBCO 2014 Grid bathymetry  
product, version 20150318, <http://www.gebco.net> (consulted 10 April 2019).

#### 125 3.1 Sampling

Carbonate chemistry data have been sampled in the Atlantic Zone since the fall of 2014 (except for high frequency station  
Rimouski where the sampling started earlier, see below). Depending on the region, between two to three surveys usually occur  
every year since.

The NL Region, due to its extensive shelf/slope spatial extent, is scheduled to survey the NL Shelf and Slope three times a  
130 year, generally from section Southwest St. Pierre Bank (SWSPB) up to the northern sea ice extent, usually section Bonavista  
Bay (BB), during spring (April) surveys, from section Flemish Cap (FC) to Beachy Island (BI) during summer (July) surveys,  
and from section SWSPB to Seal Island (SI) during fall (November) surveys (Figure 3a). The surveys are most often incomplete  
due to operational constraints caused by sea ice, weather or research vessel downtime. The NL Region aims to sample at surface  
(~5 m), 10, 50, 75, 100, 150, 250, 500, 1000 m and bottom. Sampling down the shelf slope to depths greater than 1200 m was  
135 however only feasible during fall surveys (maximum 3677 m) because of limited vessel capacity during other seasons.

The Maritimes Region has a spring (April) and a fall (September/October, except November/December in 2017) survey  
which were consistently sampled between 2014 and spring 2020 (Figure 3a). The Scotian Shelf and Slope were sampled at  
surface (~2 m), 10, 50, 80, 100, 150, 250, 500, 1000, 1500 m and bottom.

The GSL is sampled during spring (June) and fall (October/November) surveys along the standard sections at surface (~2 m),  
140 15, 50, 100, 150, 200, 300, 400 m and bottom (Figure 3b). In 2015, the spring carbonate dataset (Trip\_Name 'iml2015160')  
contains only pH, while both pH and TA were sampled in the fall at the surface and bottom (Trip\_Name 'iml2015041').  
Samples are also acquired during groundfish surveys in the GSL (Figure 3b) during summer (August/September) 2017-2019  
(Trip\_Name: 'IML2017027', 'PER2017018', 'PER2017024', 'PER2017016', 'PER2017026', 'MLB2017001b', 'IML201841',  
'PER2018041', 'PER2018042', 'PER2018043', 'TEL2018196', 'IML2019036', 'PER2019140', 'PER2019008', 'TEL2019201',



145 'PER2020019'). Although these surveys are not within the regular AZMP program, they are staffed by the AZMP for oceanographic sampling and provide extensive spatial and added temporal (summer) coverage of carbonate parameters of the GSL.

The fall 2014 and spring 2017 AZMP surveys had the most complete spatial coverage of the Atlantic Zone among the entire time series. The other surveys are partial due to operational limitations specific to each region. Ship surveys identifier (*Trip\_Name*) and other bottle collection information are listed in Table 1.

150 In addition, two coastal stations are sampled at a greater temporal frequency. Rimouski Station (labeled "Rimouski" in Figure 1) located in the LSLE is sampled by the Quebec Region on a near-weekly basis. Station 27 (labeled "STN27" in the data set and in Figure 1), located on the path of the Labrador coastal current just outside St. John's harbour is sampled by the NL Region on a near-monthly basis. To illustrate their respective temporal coverage, the time series of pH at these stations (measured at Rimouski Station and derived from TA/DIC at Station 27) are presented in Figure 4. Note that Rimouski Station  
155 is the only time series starting in 2013 in this data set (all other sampling starts in the fall of 2014).

The collection of discrete water samples for carbonate analyses was achieved using a 12 or 24 Niskin bottles rosette equipped with a SeaBird Conductivity-Temperature-Depth (CTD) coupled with an oxygen sensor to continuously measure depth, temperature, salinity and dissolved oxygen concentration (DO). Aliquots of water were subsampled from each Niskin bottle for nutrients ( $\text{PO}_4$ ,  $\text{SiO}_3$ ,  $\text{NO}_3 + \text{NO}_2$ ) and DO, then processed and analysed according to established standard protocols (Mitchell  
160 et al., 2002). Although the DO concentration ( $\text{mLL}^{-1}$ ) is available with this dataset, it was sampled less frequently than the other oceanographic parameters because its purpose is to calibrate the DO sensor measurements. The GSL Region sampled DO at the surface and bottom, and the MAR region sampled DO at surface, bottom and mid-depth of the water column. The NL region sampled DO at all depths, but at a subsampled number of stations (roughly every 2 to 4 stations along a section). GSL and NL Regions used their subset of bottle DO values to calibrate their DO sensor on the CTD providing a DO value for each  
165 sample bottle (even when no Winkler titration was realized). For MAR data, only DO data obtained from Winkler titration are provided. Oxygen saturation ( $\text{O}_{2\text{sat}}$ ) in % was calculated as the ratio between the measured DO concentration and its solubility referenced to surface using the TEOS-10 toolbox (McDougall and Barker, 2011) and corresponding T-S observations.

Aliquots of water for the analysis of pH, total alkalinity (TA) and total dissolved inorganic carbon (DIC) (or combination thereof) were subsampled following protocols established by Dickson et al. (2007). Each aliquot was carefully drawn from  
170 the Niskin bottles, transferred into 500 mL gas-tight borosilicate reagent bottles (Corning, USA), and allowed to overflow by one volume while precluding air bubbles. A 5 mL volume was then removed to create a headspace for thermal expansion. The samples were preserved with 100  $\mu\text{L}$  of saturated  $\text{HgCl}_2$  to prevent further biological activity. Each bottle was sealed with a ground-glass stopper, high-vacuum (Apiezon M) grease, and a rubber band then stored in the dark between 4 and 18°C for up to 6 months prior to analysis.

### 175 3.2 Analytical methods – carbonate parameters

In 2014, the first year of the program, all three carbonate parameters (pH, TA, DIC) were analysed by the Maritimes Region at the Bedford Institute of Oceanography (BIO, Dartmouth NS). In subsequent years, the GSL region analysed TA and pH at the Maurice Lamontagne Institute (MLI, Mont-Joli QC), while the Maritimes (BIO) and NL (Northwest Atlantic Fisheries Centre



- NAFC, St. John's NL) Regions analysed TA and DIC within their respective facilities. Since fall 2019, IML also analyzes  
180 DIC in addition to pH and TA.

DIC (in  $\mu\text{mol kg}^{-1}$  of seawater) is extracted as  $\text{CO}_2$  by purging an acidified (1M, 8.5% phosphoric acid in excess) aliquot  
of water (warmed to  $25^\circ\text{C}$ ) with ultra-high purity (UHP) nitrogen gas using an automated sampling and gas extraction system.  
The dried gas, including the  $\text{CO}_2$ , is transferred and absorbed into a coulometric cell and analysed by titration and photometric  
detection (Johnson et al., 1993). Various makes and models of autosamplers and analyzers were used by the different regions.  
185 Duplicate or triplicate sample measurements provided an analytical precision of 0.03% at BIO and  $< 0.1\%$  at IML and NAFC.

TA (in  $\mu\text{mol kg}^{-1}$  of seawater) was measured by open - cell potentiometric titration (Dickson et al., 2007; Mintrop et al.,  
2000) with an automated sampling system. Each 50 mL sample, warmed to  $25^\circ\text{C}$ , is titrated with 0.1 M hydrochloric acid  
to the Gran equivalence point (non-linear curve fit method) using a computer controlled Dosimat (Metrohm AG) dispenser  
and combination glass electrode. In the NL Region, aliquots of 0.1M hydrochloric acid are added in a  $25^\circ\text{C}$  thermostated  
190 jacket open cell while gently mixing the sample, and pH measurements are allowed to stabilize between successive readings.  
The analytical precision of TA at BIO was calculated using repeat analyses of bulk seawater and reported as  $\pm 0.05\%$ , while  
duplicate sample analyses revealed a precision of  $< 0.1\%$  at IML and NAFC.

Seawater pH expressed in the total hydrogen ion scale ( $pH_T$ ) was determined by spectrophotometry (Clayton and Byrne,  
1993; Dickson et al., 2007). Purified m-cresol purple solution was added to seawater held at  $25 \pm 0.05^\circ\text{C}$  in a 10 cm path-  
195 length quartz cell and mixed thoroughly. The ratio of blank-corrected absorbances measured at 434, 578 and 730 nm with  
a spectrophotometer (Agilent Technologies, USA) was used to determine  $pH_T$ . Precision and accuracy, evaluated daily by  
repeat measurements of a tris(hydroxymethyl)aminomethane (TRIS) buffer solution (A.G. Dickson, Scripps Oceanographic  
Institution, San Diego, CA) prepared with a salinity of 30 (Millero, 1986), were typically  $\pm 0.002$  pH units at BIO and  $\pm 0.005$   
pH units or better at IML.

200 All analytical methods were calibrated with a series of seawater certified reference materials (CRM; A.G. Dickson, Scripps  
Institute of Oceanography, San Diego, CA), which allowed for performance evaluations of the various instruments and to  
normalize the TIC, TA and pH measurements. Since the analyses were conducted at three different regions, we will consider  
the largest analytical error provided for all measurements: TA and DIC values will be  $\pm 0.1\%$  or  $\pm 2.1 \mu\text{mol kg}^{-1}$ , and pH will  
be  $\pm 0.005$  units. These uncertainties are comparable to those suggested by Dickson (2010) for modern analytical techniques  
205 using reference materials. All data flagged as suspect because of analytical error by each laboratory have been removed from  
the dataset.

### 3.3 Calculation of carbonate parameters

The aragonite saturation state ( $\Omega_{\text{arg}}$ ), calcite saturation state ( $\Omega_{\text{cal}}$ ; in dataset but not discussed in this report),  $pH_{T,\text{is}}$  (total  
scale, in situ) and  $p\text{CO}_2$  ( $\mu\text{atm}$ ) were calculated using the CO2SYS program (Lewis and Wallace, 1998) modified for Python  
210 (Humphreys et al., 2020, <https://github.com/mvdh7PyCO2SYS/tree/v1.2.1>) as recommended for "best practices" by Orr et al.  
(2015). The dissociation constants ( $K_1$ ,  $K_2$ ) of Mehrbach et al. (1973) as refit by Dickson and Millero (1987) (Mdm), total  
boron constant from (Uppstrom, 1974), and  $K\text{HSO}_4$  constant from Dickson (1990) were also used as recommended for "best-



practices” (Chen et al., 2015; Dickson et al., 2007; Orr et al., 2015). Although it has been suggested that dissociation constants formulated for estuarine waters (Cai and Wang, 1998; Millero, 2010) should be used to avoid differences in the calculation of carbonate parameters at low salinity (Dinauer and Mucci, 2017), there is some evidence that formulations other than those for “best practices” may produce discrepancies (Orr et al., 2015; Dinauer and Mucci, 2018). The dissociation constants by Mdm were formulated for salinities  $> 20$ . Samples with salinities  $< 20$  constitute only 0.15% of this dataset (1% of the data from the GSL), and the differences in calculated carbonate parameters using the constants by Cai and Wang (1998) and Mdm are approximately double for samples with salinity  $\geq 20$  than those  $< 20$  (Table 2). Therefore, the dissociation constants by Mdm were used in order to avoid discrepancies to the other 99.85% of the dataset. The chosen combination of constants listed above was also suggested by Raimondi et al. (2019) because they generated the best internal consistency in their Labrador Sea (AR7W) dataset. However, their choice of constants provided less uncertainty than their choice of input parameters. The carbonate parameters in this dataset were calculated using the TA ( $\mu\text{mol kg}^{-1}$  seawater, or SW) provided by all regions and either DIC ( $\mu\text{mol kg}^{-1}$  SW) or  $pH_T$  (total scale), which is region dependent. According to the results by Raimondi et al. (2019), the TA-DIC based calculations of pH (e.g., NL and MAR Regions) produce low accuracy yet low uncertainty, and the TA-pH based calculations of DIC (e.g., GSL dataset) provide the best consistency. The TA-pH pair will thus be used for calculating  $p\text{CO}_2$  when both pH and DIC were collected in addition to TA (e.g. Trip\_Name ‘IML201437’, ‘18HU14030’, ‘IML2019049’, ‘PER2019140’, ‘PER2019008’, ‘TEL2019201’, ‘IML2019040’, ‘IML2020028’, ‘IML2019036’, ‘PER2020019’ and ‘IML2020040’). Since there are only a few surveys that include all three parameters (TA, pH and DIC), we have not produced the errors associated with the calculation of carbonate parameters because it would not be a representative calculation for the entire Atlantic Zone.

The spring 2015 GSL dataset (Trip\_Name ‘IML2015160’) consists solely of pH data (in vitro at  $25^\circ\text{C}$ ). In order to obtain in situ values and other carbonate parameters, TA was estimated using the TA-salinity linear relationship (Cai et al., 2010; Fassbender et al., 2017). Using spring data for all depths and remaining years in the GSL (excluding the high resolution Rimouski station data), this led to the fit:  $\text{TA} = 41.26 \times \text{S} + 870.05$  ( $r^2 = 0.93$ ,  $p < 0.001$ ; see Figure 5b). The estimated freshwater TA end-member ( $\text{S}=0$ ) is  $870.05 \mu\text{mol kg}^{-1}$ . This value is lower than the measured values of  $1204 \pm 99 \mu\text{mol kg}^{-1}$  or  $1081 \pm 30.2 \mu\text{mol kg}^{-1}$  by Dinauer and Mucci (2017, 2018) near Québec City, and the  $1000 \mu\text{mol kg}^{-1}$  measured by Mucci et al. (2017) west of the Saguenay Fjord. It is however higher than  $186 \mu\text{mol kg}^{-1}$  calculated by Dinauer and Mucci (2018) as a measure of the Saguenay Fjord and North Shore Rivers, and  $80 \mu\text{mol kg}^{-1}$  in the Saguenay River measured by Mucci et al. (2017). Using a localised dataset for this calculation accounted for the low salinity water exiting the St. Lawrence River. Considering the measurement error of up to 0.1%, there was no significant difference in the predicted TA between the spring dataset and all seasons in the GSL. To estimate the degree in which the predicted TA had an effect on the calculation of the other carbonate parameters, the parameters for the spring 2015 GSL dataset were calculated in CO2SYS (TA-pH) with the maximum and minimum values of  $\text{TA} \pm 2 \text{SD}$  ( $1.28 \mu\text{mol kg}^{-1}$ ), the TA-salinity regression uncertainty (Table 2).

The calculated carbonate parameters were calibrated to in situ conditions using temperature ( $^\circ\text{C}$ ), salinity (psu) and pressure (dbar). The nutrient alkalinity, which is represented by total soluble reactive phosphorus ( $\text{PO}_4$ ,  $\mu\text{mol kg}^{-1}$  SW) and total soluble reactive silicate ( $\text{SiO}_3$  -  $\mu\text{mol kg}^{-1}$  SW), were used as additional parameters when available since they contribute to the calculation of the total carbonate alkalinity (Orr et al., 2015). There can be a slight offset in calculated carbonate parameters



when the nutrient data are absent, especially in high concentration regions. This offset has been calculated for the Atlantic Zone by obtaining the difference in carbonate parameters calculated with and without nutrients (nutrients set to zero in CO2SYS; Fassbender et al., 2017). The calculation was performed on two datasets: the TA-DIC input pair from the NL and MAR regions and the TA-pH input pair from the GSL region. The results (Table 2) indicate that all calculated carbonate parameters except for pCO<sub>2</sub> will be slightly higher when nutrient data are lacking, with TA-DIC as input pair having the largest offset. The difference in calculated pH (0.002 pH units) and DIC (1.02 μmol kg<sup>-1</sup>) values are below the analytical uncertainty (±0.005 pH units, ±2.1 μmol kg<sup>-1</sup>, respectively). The mean saturation states and pCO<sub>2</sub> have changed by < 0.6% for the TA-DIC pair and < 0.1% for the TA-pH pair. The larger nutrient-no nutrient difference with the TA-DIC pair may be due to the higher nutrient concentrations in the GSL Region (> 17.3% PO<sub>4</sub>, > 49.1% SiO<sub>3</sub>), or reflecting the input parameter propagated error as suggested by Raimondi et al. (2019) as well as Dickson (2010) who estimated the uncertainty in calculating the saturation state with 3.7% error using the TA-pH pair relative to 1.7% with TA-DIC.

#### 4 Results

While a detailed description of the spatio-temporal variability of all physical and biogeochemical parameters over the entire Atlantic Zone remains virtually impossible as part of this study, the entire dataset used here is provided for follow up studies (see Data Availability section). This dataset includes the following physical, biogeochemical, and carbonate parameters: temperature (*T*), salinity (*S*), dissolved oxygen concentration (*DO*), oxygen saturation (O<sub>2,sat</sub>), nutrient data (PO<sub>4</sub>, SiO<sub>3</sub>, NO<sub>3</sub> + NO<sub>2</sub>), measured and calculated (same as measured when available), TA, DIC and pH, and calculated pCO<sub>2</sub>, Ω<sub>arg</sub>, and Ω<sub>cal</sub>.

In the following subsections, a summary of the physical and carbonate parameters is presented for different regions, seasons and depth range. Here surface is defined as the shallowest sample, but no deeper than 52 m (a depth chosen to encompass sampling variations around the targeted 50 m depth), and bottom as the deepest sample below 50 m. Due to some instances where near-bottom samples were not achieved (>1200m areas due to ship limitation, e.g. NL slope), the following descriptions are restricted to depth shallower than 600 m. All of the data are however in the dataset.

A subset of the data (excluding nutrients and O<sub>2</sub>) is presented in Appendix A on maps of the surface and bottom waters of the Atlantic Zone for each sampling season of 2017, the most extensively sampled year to date (Figures A1-A9). Each map represents one variable per depth per season in 2017. Seasons are defined as spring (beginning of March to end of June), summer (beginning of July to September 14<sup>th</sup>), and fall (September 15<sup>th</sup> to end of December). The fall 2017 maps are presented in Figures 6 to 8.

Depth profiles along a series of hydrographic sections from each region during the fall of 2017 are presented in Appendix B for the same set of variables. These sections include Seal Island (SI; Figure A10), Flemish Cap (FC; Figure A11), Halifax (HL; Figure A12), Cabot Strait (CSL; Figure A13), and Laurentian Channel (LC; Figure A14) which is the line of stations from the centre of the Lower St. Lawrence Estuary through the Laurentian Channel to Cabot Strait (Figure 3a).



#### 4.1 Oceanographic context ( $T$ , $S$ , $O_{2_{\text{sat}}}$ )

280 A series of temperature and salinity maps (Figures 6a-d, A1, A2) illustrate the physical oceanographic properties of the Atlantic Zone. The Arctic waters of the Labrador Coastal Current on the northern NL Shelf are characterized by low salinities and temperatures near freezing, while the Scotian Slope surface is characterized by warmer ( $>18^{\circ}\text{C}$ ) and more saline ( $>33$ ) Gulf Stream waters (Figure 2a). The lowest salinities ( $<27$ ) are found in the surface waters of the LSLE (ignoring the Saguenay Fjord and the USLE; see Figure 3).

285 The moderately cold and saline surface waters along the southeastern NL Shelf and within the GSL consist of Arctic waters that have been modified by the Gulf Stream, St. Lawrence River, or warmed in summer. The freshwater exiting the LSLE can be traced at surface along the coast of the Gaspé peninsula into the southern GSL and Northumberland Strait (mean  $S\sim 29$ ), where it warms significantly in summer (mean summer  $T\sim 16^{\circ}\text{C}$ ) (Figures A1, A2). The GSL encompasses the greatest sea surface temperature differences between spring and summer, yet the largest differences are from spring to fall and found on the  
290 Scotian Shelf ( $> 15^{\circ}\text{C}$ ). Since the AZMP does not include carbonate winter sampling, the coldest temperatures in this dataset, down to  $-1.7^{\circ}\text{C}$ , are located within the CIL, in the subsurface waters of the NL Shelf (hydrographic section BB, Figure 9a) and in the GSL (hydrographic section LC, Figure 9b). Warm, saline bottom waters influenced by the Gulf Stream can be traced along the southwest Scotian Shelf and Laurentian Channel into the GSL. Shallow bottom waters of the GSL can warm in summer and exceed  $17^{\circ}\text{C}$  (not shown).

295 The oxygen saturation ( $O_{2_{\text{sat}}}$ , see methodology) is greatest at the surface with mean regional values  $> 90\%$  and supersaturation occurring during spring plankton blooms (Figure 6e). Along the southern NL and Scotian outer shelf and slope, an oxygen minimum zone with  $O_{2_{\text{sat}}}$  down to 45% is centered around 250 m (e.g. hydrographic section SESP, Figure 9c). Mean bottom values on the NL Shelf range from nearly 100% off Labrador to  $\sim 55\%$  off southern NL. The  $O_{2_{\text{sat}}}$  reaches supersaturation in the shallow bottom waters of the southern GSL and undersaturation in deeper areas associated with the Laurentian Channel.  
300 Hypoxia, which occurs at the bottom of the LSLE and deeper areas of the GSL such as the Laurentian, Anticosti and Esquiman Channels (Figure 1; Gilbert et al. (2005)), is shown as red circles in Figure 6f.

#### 4.2 Carbonate parameters ( $\text{TA}$ , $\text{DIC}$ , $\text{pH}$ , $\Omega_{\text{arg}}$ )

This section comprises of a brief description of the spatial and temporal variations of carbonate parameters of the Atlantic Zone. Except for 2015, the values and seasonal cycles among annual surveys are relatively comparable, and the descriptions refer to  
305 maps and sections from 2017 (Figures 6-8, A1-A9). Comparing seasonal changes (spring, summer, fall) of the Atlantic Zone or changes by region is difficult due to the surveying protocols established by each region. The hydrographic sections FC and BB on the NL Shelf (Figure 3a) are the only ones planned for the three seasons every year (all were sampled except fall 2016 for FC and BB, and spring 2016 for BB), however the station separation is greater along BB, and thus the resolution poorer. Although the standard AZMP GSL sections are only surveyed in spring and fall, the summer groundfish surveys between 2017  
310 and 2019 provide enough data to evaluate some seasonal trends in the GSL (see below).



#### 4.2.1 Total Alkalinity

The linear relationship between TA and S for the Atlantic Zone provides an  $r^2 = 0.93$  ( $TA = 40.38 \times S + 892.58, p < 0.001$ ; Figure 5), and consequently TA values follow the same spatial pattern as S (Figures 6c,d, 7a,b, A2, A4). The lowest TA concentrations are located in surface waters of the LSLE and GSL due to freshwater outflow from the LSLE (TA value at S = 0 is  $< 1200 \mu\text{mol kg}^{-1}$ ; see section Calculation of carbonate parameters), which are also observed exiting the GSL along southwest Cabot Strait onto the Scotian Shelf (Figure 7a).

The surface TA is also low along the NL coast which is associated with coastal runoff and the fresh Labrador Coastal Current. Largest TA values, reaching nearly as high as reach  $2400 \mu\text{mol kg}^{-1}$ , are found within the upper 150 m of the southern NL and Scotian slope waters. Following the seasonal cycle of salinity, TA generally decreases from spring to summer/fall on the NL/Scotian Shelves, reflecting respectively the continued freshening of the NL coast (Cyr and Galbraith, 2021) and the freshwater export from the LSLE exiting the Gulf of St. Lawrence at Cabot Strait (Dever et al., 2016). Although the TA of the Atlantic Zone bottom waters follows a similar spatial pattern as the surface, the values are higher and the range is much narrower (Figure 7b). The mean shelf bottom ( $< 600\text{m}$ ) TA is  $2241 \mu\text{mol kg}^{-1}$ , with lowest values in the USLE and shallow southern GSL ( $\sim 2000 \mu\text{mol kg}^{-1}$ ). The highest TA values of the Atlantic Zone are in the arm Slope Water ( $\sim 2300 \mu\text{mol kg}^{-1}$ ) in deep waters of the Scotian Slope and GSL.

#### 4.2.2 Dissolved Inorganic Carbon

The highest concentrations of DIC are located in the deepest areas ( $> 275\text{ m}$ ) of the LSLE and GSL specifically the Laurentian, Anticosti and Esquiman channels, with maximum values just over  $2300 \mu\text{mol kg}^{-1}$  (Figures 7d, A5). Moderately high concentrations are observed deep offshore along the NL (up to  $2264 \mu\text{mol kg}^{-1}$ ) and Scotian (up to  $2244 \mu\text{mol kg}^{-1}$ ) shelves and slopes. These values decrease on the shelf toward the coasts, and the lowest values are in the shallow areas of the shelf under freshwater influence (down to  $\sim 2000 \mu\text{mol kg}^{-1}$  in the USLE). The accumulation of DIC within shelf bottom waters in fall is due to the remineralization of organic matter sinking to the bottom after spring and summer plankton blooms, with the highest values in the poorly ventilated deep GSL waters. The surface water has significantly lower DIC values, with the lowest concentrations occurring in the SLE, as well as the southern GSL and the coastal stations along the northern NL Shelf (Figure 7c).

In southern NL and the eastern GSL, the DIC decreases from spring to summer as it is consumed during the plankton bloom, then increases from summer to fall during the remineralization of that produced organic matter. The decrease in values from spring to fall along the Scotian Shelf reflects the transition from high winter DIC values due to remineralization, cold temperatures (increased solubility), and mixing with deep DIC enriched water during winter convection, to lower values in fall due to photosynthesis and stratification.



### 4.2.3 Partial pressure of CO<sub>2</sub>

The lowest pCO<sub>2</sub> values (down to ~200 μmol kg<sup>-1</sup>) are located on the Newfoundland and Labrador Shelf, particularly in the coldest northern surface waters (Figures 7e, A6). pCO<sub>2</sub> generally increase from surface to bottom and from spring to fall as plankton from the spring bloom is consumed (surface/spring) then remineralized (bottom/fall). Higher temperatures also influence the seasonal and southward increase in pCO<sub>2</sub>. Therefore, slightly higher values are located in the bottom waters of the southern NL and Scotian shelves and slopes, reaching a maximum (~600 μmol kg<sup>-1</sup>) at the O<sub>2</sub> minimum (Figure 9c-d). Coincident with DIC, the highest pCO<sub>2</sub> values are located in the deepest areas of the LSLE and GSL, with values up to ~1600 μmol kg<sup>-1</sup> (Figure 9f). This CO<sub>2</sub>, which accumulates at depth due to poor ventilation in the GSL, is produced by the remineralization of organic matter.

### 350 4.2.4 pH

The pH of the Atlantic Zone ranges from approximately 7.5 in the LSLE and GSL bottom waters (and lower in the Saguenay River) to 8.3 in the surface waters off Labrador, the highest values generally found along the shelf once the sea ice has melted in summer (Figures 8a-b, A7). The plankton bloom, which consumes CO<sub>2</sub>, is concurrent with the ice retreat, which increases the pH. Stations with high pH are often located away from shore with higher T and S/TA (Figure 6a-c, 8a), and less influenced by coastal processes. High pH is also observed during most annual spring surveys along the Scotian Shelf (particularly in 2017). These high pH values may be associated with the spring plankton bloom. The surface water pH is relatively uniform throughout the Atlantic Zone in the fall (~7.8 to 8.1), with the higher values along the NL and Scotian Shelves. The decreased GSL surface pH is influenced by the freshwater exiting the LSLE (area of lowest surface pH) and high concentrations of plankton which consumes CO<sub>2</sub> (increases pH) and is subsequently remineralized (decreases pH).

360 The bottom waters of the GSL, specifically the LSLE and northern and deeper areas of the GSL including the Laurentian, Anticosti and Esquiman Channels, contain the lowest pH (down to 7.5) of the Atlantic Zone (Figure 8b). These areas are also associated with the lowest O<sub>2sat</sub> (~16%; Figure 6f). Due to the estuarine nature of the GSL, its bottom waters have restricted ventilation allowing for the accumulation of CO<sub>2</sub> and reduction in pH produced by the respiration of organic matter in this highly productive environment. The highest bottom pH values are located within the cold shallow bottom waters of the GSL and in the coldest waters of the NL Shelf. The bottom pH of the Atlantic Zone generally decreases throughout the year, likely due to the remineralization of organic matter (Figure A7). The spatial and temporal distribution of pH appears to be inversely correlated with pCO<sub>2</sub>, which describes a pattern of the photosynthesis and subsequent remineralization of plankton.

### 4.2.5 Saturation state relative to Aragonite (Ω<sub>arg</sub>)

370 The Atlantic zone Ω<sub>arg</sub> ranges from 0.5 in the LSLE and GSL bottom waters (lower values in the Saguenay River) to 3.7 in the surface waters of the southwest Scotian Slope (Figures 8c,d, A8). These highest Ω<sub>arg</sub> surface waters are most influenced by the warm and saline Gulf Stream waters (Figure 6a,c and 8c, southernmost stations). Ω<sub>arg</sub> > 2.0 are also located in the surface waters of the outer NL shelf and slope. On average, these Scotian and NL waters increase in Ω<sub>arg</sub> from spring to fall



(Figure A8), corresponding to the seasonal temperature increase and decrease in S and TA (Figures A1, A2 and A4). Excluding the LSLE, the surface GSL  $\Omega_{\text{arg}}$  values are similar to those from the southern NL and northeastern Scotian Shelves ( $\sim 1.5 - 2.0$ ). The surface waters of the USLE are undersaturated ( $\Omega_{\text{arg}} < 1$ ) with respect to aragonite (and in some instances calcite) due to low TA and DIC (Figures A4, A5, A8 and A9), and can be traced into the GSL along the north coast of Gaspé. The bottom waters of the GSL, specifically the LSLE and northern and deeper areas of the GSL including the Laurentian, Anticosti and Esquiman channels, are undersaturated with respect to aragonite (Figure 8d).

Most of the bottom GSL samples (92%) are undersaturated, with the few saturated samples in shallow areas or along the Cabot Strait section. The poorly ventilated GSL allows for the accumulation DIC produced by the decomposition of organic matter in this highly productive environment. The high DIC, coupled with a low buffering capacity (TA), produced the undersaturated benthic conditions. Undersaturated waters also occur on the southeast Scotian Shelf where GSL bottom water exits through Cabot Strait, as well as the southern (colder) NL Shelf. These conditions are present during summer (NL) and fall (MAR, NL) surveys due to the remineralization of the spring/summer plankton bloom. Although undersaturation is present, the mean  $\Omega_{\text{arg}}$  on the NL and Scotian Shelves is  $\sim 1.3$ . The deep Scotian and NL Slopes are also undersaturated, and the saturation horizon ( $\Omega_{\text{arg}}=1$ ) is between 2200 m and 2300 m (Figure 10). Azetsu-Scott et al. (2010) has demonstrated that in the Labrador Sea, this horizon corresponds to the depth where the Labrador Sea Water overlies the North East Atlantic Deep Water. Therefore, the depth of the saturation horizon along the Labrador Slope can fluctuate with time depending on the deep winter convection in the Labrador Sea (Azetsu-Scott et al., 2010; Yashayaev and Loder, 2017).

## 390 5 Discussion and Conclusion

In the Atlantic Zone, the spatial and temporal variability of carbonate parameters reflects changes in both physical (temperature, salinity) and biological (plankton photosynthesis and respiration) parameters. While a complete description of the seasonal variability of the Atlantic zone is difficult due to the timing of the surveys and large geographic areas of all three regions, some general description can be drawn.

The surface is characterized by cold (Arctic) and warm (Atlantic) waters that undergo a seasonal warming and freshening (low TA), and decreased carbonate during spring/summer plankton blooms. The lowest  $\text{pCO}_2$  values are located in the cold Coastal Labrador Current and the highest in the fresh waters of the LSLE. These values generally increase from surface to bottom and from spring to fall as plankton from the spring bloom is consumed at surface and remineralized near the bottom in the fall. The lowest pH values are found in the GSL and the LSLE. This is especially true for the southern portion of the Gulf and in the deep (> 300m) layers of the Laurentian Channel. The most acidic waters of the Atlantic Zone are found in the Lower St. Lawrence Estuary, where pH has decreased by 0.2 to 0.3 between the 1930's and the early 2000's, a region affected by severe hypoxia (Mucci et al., 2011). These low and undersaturated conditions, which cover a large part of the Atlantic Zone, are concerning and not favourable for calcifying organisms.

$\Omega_{\text{arg}}$  increases at surface and in shallow areas of the shelf from spring to fall in most MAR and NL stations as a consequence of the seasonal temperature increase and decrease in S and TA coupled with high DIC of respired organic matter (see also



Shadwick et al., 2011a, b, for a description of the CO<sub>2</sub> system seasonal cycle on the SS). Most of the bottom GSL is also undersaturated with respect to aragonite ( $\Omega_{\text{arg}} < 1$ ), except for a few saturated samples in shallow areas and along the Cabot Strait section. Consistent with Azetsu-Scott et al. (2010), low  $\Omega_{\text{arg}}$  are found in the cold and fresh (thus lower TA) Arctic-origin waters of the Coastal Labrador Current. These cold acidic waters are a prominent feature of the Atlantic Zone as they contribute  
410 to the LSW formation southwest of the Grand Banks. The LSW, which has a  $\Omega_{\text{arg}} < 2$ , partly forms the deep waters of the Laurentian Channel in the GSL (Gilbert et al., 2005) then continues southward on the Scotian Slope and along the Northeast US Coast. These low  $\Omega_{\text{arg}}$  values can be traced to the deep Gulf of Maine. As the water temperature increases during their journey towards the southern US,  $\Omega_{\text{arg}}$  increases reaching  $\sim 4.5$  in the Gulf of Mexico (Wanninkhof et al., 2015).

As a result of the increasing atmospheric CO<sub>2</sub> uptake, the ocean has undergone acidification during the 20<sup>th</sup> century (Pörtner  
415 et al., 2019). Ocean acidification is also projected to continue through the 21<sup>st</sup> century (Pörtner et al., 2019). Observation programs such as the AZMP are essential to monitor these changes and their consequences on ecosystems (e.g., Tilbrook et al., 2019). Because *in situ* observations are often scattered in time and space, ocean biogeochemical models are another important tool available to improve our understanding of seasonal, inter-annual and climatic variations of ocean acidification. As the field of biogeochemical modeling is rapidly developing, the availability of biogeochemical data at meaningful spatial and temporal  
420 scales however appears as a considerable challenge that hinders the implementation or validation of such models at regional scales (Fennel et al., 2019; Capotondi et al., 2019; Pilcher et al., 2019; Lavoie et al., 2021). In addition to providing a baseline of carbonate parameters, a comprehensive overview of carbonate parameters such as the one provided here is thus a necessary and useful contribution to the modeling community.

Using the decrease in  $\Omega_{\text{arg}}$  of  $0.5 \pm 0.2$  in the surface ocean by the year 2100 suggested by Bates et al. (2009), it appears that  
425 ocean acidification may impact parts of the ecosystem within the next century. As climate changes, however, the Atlantic Zone will be influenced by changes in temperature, salinity, ocean currents, nutrients and productivity, all of which will contribute to regional changes in pH and saturation state. While this study provides a useful baseline for the Atlantic Zone, incorporating these data into biogeochemical models is a necessary step in order to examine the regional response to ocean acidification and the future of the Atlantic Canada aquaculture and fishing industries (Lavoie et al., 2020; Siedlecki et al., 2021).

## 430 6 Data availability

The full data set of measured and derived parameters is available in the Federated Research Data Repository at <https://doi.org/10.20383/102.0673> (Cyr et al., 2022). It consists of a single comma-separated values (CSV) file named *AZMP\_carbon.csv*. This file contains 12923 lines, each corresponding to a discrete sample. Out of this number, 11890 contains the full suite of parameters derived using *CO2sys*, the remaining 1033 samples have only one of the three carbonate parameters (TA, TIC or  
435 pH) available. A brief description of the columns in this file is provided here (see methods for a detailed description of the quantities):

- *Timestamp*: Date and Time (UTC) that the sample was collected in format 'yyyy-mm-dd HH:MM:SS'



- *Region*: Geographical region where the sample was collected ('GSL', 'MAR' or 'NL').
- *Trip\_Name*: Name of the sea going mission (convention varies between regions).
- 440 – *Station\_Name*: Hydrographic station names (e.g., 'TESL3', 'STN27', etc.).
- *Latitude\_(degNorth)*: Latitude of the sampling in decimal format (in °N).
- *Longitude\_(degEast)*: Longitude of the sampling in decimal format (in °E).
- *Depth\_(dbar)*: Depth of the sampling (in decibars). In some rare circumstances, the nominal depth of the sample was used when the CTD depth was missing.
- 445 – *Temperature\_(degC)*: Temperature (in °C) using the International Temperature Scale of 1990 (ITS-90)
- *Salinity\_(psu)*: Salinity expressed on the practical salinity scale (psu, unitless)
- *Dissolved\_Oxygen\_(mL/L)*: Dissolved oxygen concentration (in mL L<sup>-1</sup>) obtained either by Winkler titration or DO sensor calibrated *in situ* with Winkler titration.
- *Nitrate\_Concentration\_(mmol/m3)*: Nitrate (and Nitrite) concentration (in mmol m<sup>-3</sup>) determined in laboratory with  
450 auto-analyzer.
- *Phosphate\_Concentration\_(mmol/m3)*: Phosphate concentration (in mmol m<sup>-3</sup>) determined in laboratory with auto-analyzer.
- *Silicate\_Concentration\_(mmol/m3)*: Silicate concentration (in mmol m<sup>-3</sup>) determined in laboratory with auto-analyzer.
- *Total\_Alkalinity\_(umol/kg)*: Total Alkalinity (TA in μmol kg<sup>-1</sup>), measured or derived using CO2SYS (no missing values).  
455
- *Inorganic\_Carbon\_(umol/kg)*: Inorganic Carbon (DIC in μmol kg<sup>-1</sup>), measured or derived using CO2SYS (no missing values).
- *pH\_tot\_(total\_scale)*: pH on the total scale (unitless), measured or derived using CO2SYS (no missing values).
- *Omega\_Aragonite\_(unitless)*: Saturation state relative to Aragonite ( $\Omega_{\text{arg}}$ , unitless).
- 460 – *Omega\_Calcite\_(unitless)*: Saturation state relative to Calcite ( $\Omega_{\text{cal}}$ , unitless).
- *pCO2\_(uatm)*: Partial Pressure of CO<sub>2</sub> derived using CO2SYS (pCO<sub>2</sub>, in μatmosphere).
- *Oxygen\_Saturation\_(%)*: Dissolved Oxygen Saturation (in %).
- *pH\_lab\_(seawater\_scale)*: pH measured in the laboratory (total scale, unitless). The absence of value signifies that this parameter was not measured.



- 465 – *pH\_lab\_temp\_(degC)*: Temperature at which the pH was measured in the laboratory (in °C).
- *Total\_Alkalinity\_Measured\_(umol/kg)*: Total Alkalinity (TA in  $\mu\text{mol kg}^{-1}$ ) measured in the laboratory. The absence of value signifies that this parameter was not measured.
- *Inorganic\_Carbon\_measured\_(umol/kg)*: Inorganic Carbon (DIC in  $\mu\text{mol kg}^{-1}$ ) measured in the laboratory. The absence of value signifies that this parameter was not measured.

470 Note that the majority of the carbonate parameters data presented here have also been archived with extended metadata on the Ocean Carbon Data System (OCADS) accessible at the address <https://www.ncei.noaa.gov/access/ocean-carbon-data-system/> (last access: 13 July, 2021). Users are invited to retrieve these data for more in depth analysis of the Atlantic zone carbonate system.

475 *Author contributions.* OG and FC led the study, the data processing and archiving, and the writing. GM, KAS and SP, and MS provided the quality-controlled data for Newfoundland and Labrador shelf, the Scotian Shelf and the Gulf of St. Lawrence, respectively. PP and KAS initiated the collection of carbonate parameters as part of the AZMP in 2014. All authors reviewed and commented the manuscript.

*Competing interests.* The authors declare that no competing interests are present

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**Table 1.** Summary of all surveys conducted in the Atlantic Zone that collected carbonate chemistry data from fall 2014 to 2020 that are included in this dataset.

| Year | Season | Region | Trip Name   | Stations | Bottles    | Max Depth  | Start Date | End Date   |
|------|--------|--------|-------------|----------|------------|------------|------------|------------|
| 2014 | spring | GSL    | M14010      | 1        | 141        | 330        | 2014-04-25 | 2014-12-16 |
|      | fall   | GSL    | IML201437   | 57       | 316        | 465        | 2014-10-27 | 2014-11-09 |
|      |        | MAR    | 18HU14030   | 37       | 241        | 2905       | 2014-09-19 | 2014-10-08 |
|      |        | NL     | HUD114      | 37       | 254        | 1071       | 2014-11-16 | 2014-12-07 |
| 2015 | spring | GSL    | IML201516   | 9        | 51         | 432        | 2015-06-01 | 2015-06-12 |
|      |        |        | IML2015010  | 1        | 174        | 331        | 2015-01-18 | 2015-12-18 |
|      |        | MAR    | 18HU15004   | 21       | 142        | 2941       | 2015-04-18 | 2015-04-25 |
|      |        | NL     | TEL144      | 26       | 137        | 533        | 2015-04-12 | 2015-04-27 |
|      | summer | NL     | TEL148      | 43       | 255        | 1009       | 2015-07-09 | 2015-07-26 |
|      | fall   | GSL    | IML2015041  | 14       | 85         | 464        | 2015-10-19 | 2015-11-05 |
|      |        | MAR    | 18HU15030   | 51       | 386        | 5044       | 2015-09-20 | 2015-10-08 |
|      |        | NL     | HUD115      | 43       | 255        | 3676       | 2015-11-15 | 2015-12-06 |
| 2016 | spring | GSL    | IML2016010  | 1        | 201        | 333        | 2016-01-23 | 2016-12-12 |
|      |        |        | IML2016-015 | 19       | 110        | 459        | 2016-06-01 | 2016-06-26 |
|      |        | MAR    | 18HU16005   | 15       | 191        | 4717       | 2016-05-02 | 2016-05-24 |
|      |        |        | 18HU16003   | 15       | 106        | 1254       | 2016-04-10 | 2016-04-25 |
|      |        | NL     | TEL157      | 7        | 32         | 446        | 2016-04-01 | 2016-04-06 |
|      | TEL159 | 15     | 104         | 1210     | 2016-05-11 | 2016-05-17 |            |            |
|      | summer | NL     | TEL160      | 32       | 215        | 1074       | 2016-07-08 | 2016-07-28 |
|      | fall   | GSL    | IML2016050  | 55       | 202        | 464        | 2016-10-16 | 2016-11-02 |
|      |        | MAR    | 18HU16027   | 60       | 377        | 3947       | 2016-09-16 | 2016-10-04 |
| NL   |        | HUD116 | 23          | 145      | 3303       | 2016-11-13 | 2016-11-20 |            |



**Table 1.** [continued]

| Year       | Season | Region | Trip Name   | Stations   | Bottles | Max Depth  | Start Date | End Date   |
|------------|--------|--------|-------------|------------|---------|------------|------------|------------|
| 2017       | spring | GSL    | PER2017016  | 6          | 18      | 65         | 2017-06-15 | 2017-06-16 |
|            |        |        | PER2017018  | 4          | 8       | 20         | 2017-06-23 | 2017-06-28 |
|            |        |        | IML2017080  | 40         | 224     | 466        | 2017-05-30 | 2017-06-19 |
|            |        |        | IML2017050  | 1          | 170     | 333        | 2017-02-07 | 2017-12-04 |
|            |        | MAR    | 18OL17001   | 46         | 265     | 3000       | 2017-04-19 | 2017-05-01 |
|            |        | NL     | TEL173      | 51         | 363     | 1208       | 2017-04-06 | 2017-04-23 |
|            | summer | GSL    | IML2017027  | 28         | 148     | 524        | 2017-08-03 | 2017-09-01 |
|            |        |        | MLB2017001B | 9          | 30      | 454        | 2017-08-25 | 2017-08-30 |
|            |        |        | PER2017024  | 40         | 80      | 49         | 2017-07-12 | 2017-07-30 |
|            |        |        | PER2017026  | 27         | 80      | 136        | 2017-08-16 | 2017-08-21 |
|            |        | NL     | TEL176      | 48         | 352     | 1252       | 2017-07-08 | 2017-07-28 |
|            | fall   | GSL    | IML2017048  | 46         | 223     | 455        | 2017-11-04 | 2017-11-23 |
|            |        | MAR    | 32EV17605   | 56         | 324     | 3757       | 2017-11-24 | 2017-12-15 |
|            |        | NL     | DIS009      | 27         | 195     | 1201       | 2017-11-11 | 2017-12-16 |
|            | 2018   | spring | GSL         | IML2018040 | 1       | 364        | 331        | 2018-03-12 |
| IML2018014 |        |        |             | 70         | 275     | 458        | 2018-06-05 | 2018-06-24 |
| MAR        |        |        | 18HU18004   | 43         | 320     | 4713       | 2018-04-08 | 2018-04-23 |
| NL         |        |        | TEL185      | 33         | 213     | 1202       | 2018-04-06 | 2018-04-24 |
| summer     |        | GSL    | IML201841   | 32         | 157     | 483        | 2018-08-03 | 2018-08-31 |
|            |        |        | PER2018042  | 32         | 96      | 136        | 2018-08-13 | 2018-08-21 |
|            |        |        | PER2018041  | 40         | 80      | 45         | 2018-07-11 | 2018-07-30 |
| NL         |        | COR011 | 80          | 492        | 1202    | 2018-07-15 | 2018-08-02 |            |
| fall       |        | GSL    | IML2018028  | 31         | 154     | 459        | 2018-10-23 | 2018-11-01 |
|            |        |        | TEL2018195  | 36         | 120     | 388        | 2018-09-07 | 2018-10-01 |
|            |        |        | PER2018043  | 6          | 14      | 89         | 2018-09-27 | 2018-10-07 |
|            |        | MAR    | 18HU18030   | 49         | 361     | 4144       | 2018-09-15 | 2018-10-05 |
|            | NL     | HUD118 | 46          | 318        | 3677    | 2018-11-11 | 2018-12-02 |            |



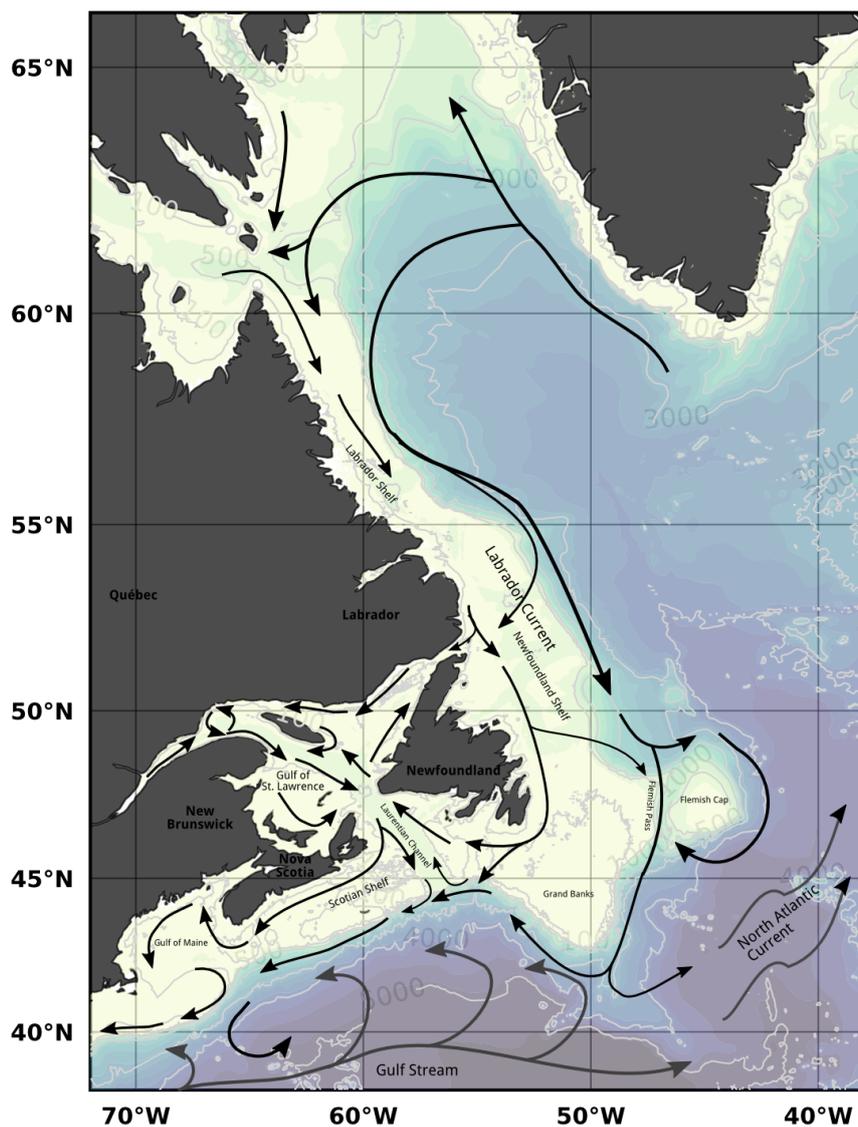
**Table 1.** [continued]

| Year | Season | Region | Trip Name  | Stations | Bottles | Max Depth  | Start Date | End Date   |
|------|--------|--------|------------|----------|---------|------------|------------|------------|
| 2019 | spring | GSL    | IML2019040 | 1        | 188     | 332        | 2019-04-12 | 2019-12-13 |
|      |        |        | IML201909  | 71       | 276     | 460        | 2019-05-26 | 2019-06-15 |
|      |        | MAR    | COR2019001 | 42       | 295     | 3770       | 2019-04-09 | 2019-04-25 |
|      |        | NL     | TEL199     | 1        | 7       | 100        | 2019-04-20 | 2019-04-20 |
|      |        |        | TEL200     | 55       | 432     | 1207       | 2019-06-27 | 2019-07-13 |
|      |        |        | TEL197     | 15       | 105     | 1197       | 2019-04-12 | 2019-04-18 |
|      | summer | GSL    | PER2019140 | 30       | 60      | 46         | 2019-07-23 | 2019-08-05 |
|      |        |        | PER2019008 | 20       | 60      | 134        | 2019-08-15 | 2019-08-18 |
|      |        |        | IML2019036 | 20       | 101     | 516        | 2019-08-15 | 2019-09-03 |
|      | fall   | GSL    | TEL2019201 | 35       | 126     | 372        | 2019-09-09 | 2019-10-01 |
|      |        |        | PER2020019 | 5        | 14      | 89         | 2019-09-22 | 2019-10-07 |
|      |        |        | IML2019049 | 45       | 228     | 470        | 2019-10-22 | 2019-11-06 |
| NL   |        | COO001 | 43         | 306      | 1220    | 2019-11-17 | 2019-12-10 |            |
| 2020 | spring | GSL    | IML2020040 | 1        | 120     | 332        | 2020-01-06 | 2020-12-09 |
|      | summer | NL     | TEL210     | 43       | 313     | 1209       | 2020-07-14 | 2020-07-31 |
|      |        |        | AMU014     | 1        | 5       | 150        | 2020-08-11 | 2020-08-11 |
|      | fall   | GSL    | IML2020028 | 39       | 206     | 470        | 2020-10-19 | 2020-10-30 |
|      |        |        | HUD2020063 | 33       | 243     | 2000       | 2020-10-04 | 2020-10-14 |
|      |        | MAR    | HUD2020063 | 34       | 255     | 2000       | 2020-10-04 | 2020-10-14 |
|      |        |        | NL         | HUD120   | 36      | 261        | 3668       | 2020-11-10 |

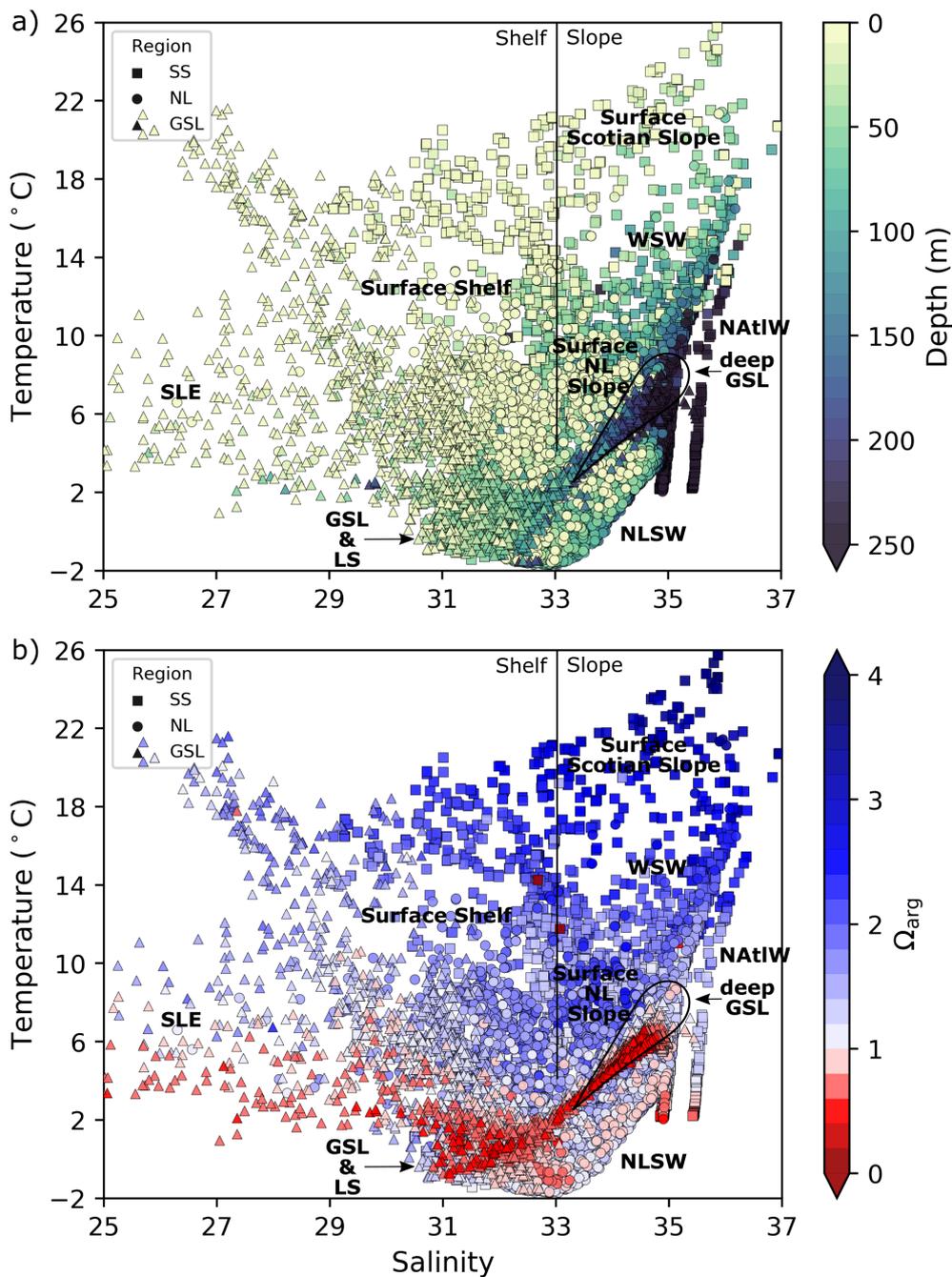


**Table 2.** Error in calculated carbonate parameters due to different sets of dataset variables: a) using dissociation constants by Mehrbach et al. (1973) as refit by Dickson and Millero (1987) (Mdm) rather than Cai and Wang (1998) at low salinities, b) presence/absence of nutrients for the two sets of input pairs, c) calculated TA from linear relationship with salinity. The absence of pH data at salinities < 20 is due to the fact that these low salinities are located in the GSL which is the region that measures pH.

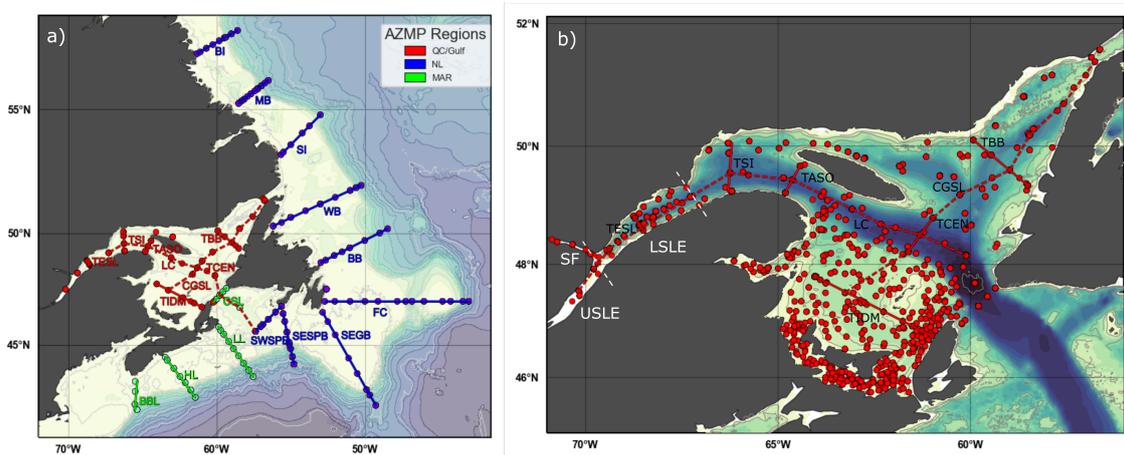
| Difference (%)  | pH     | $\Omega_{\text{arg}}$ | $\Omega_{\text{cal}}$ | pCO <sub>2</sub> | DIC        | TA         |
|---|--------|-----------------------|-----------------------|------------------|------------|------------|
| <b>a) Choice of dissociation constant</b>               |        |                       |                       |                  |            |            |
| $S_A < 20$  | 0.2    | 0.8                   | 0.8                   | 4                | 0.2        | 0          |
| $S_A \geq 20$   | -0.006 | 0.8                   | 0.8                   | -0.5             | -0.02      | <0.001     |
| <b>b) Using nutrients in CO<sub>2</sub>sys</b>          |        |                       |                       |                  |            |            |
| TA-DIC (nutrients - no nutrients)                       | -0.03  | -0.5                  | -0.5                  | -0.5             | 0          | 0          |
| TA-pH (nutrients - no nutrients)                        | 0.003  | 0.05                  | 0.05                  | -0.05            | 0          | 0.08       |
| <b>c) Error when inferring TA from <math>S_A</math></b> |        |                       |                       |                  |            |            |
| TA-pH ( $\pm 2\sigma$ on predicted TA)                  | 0      | $\pm 0.03$            | $\pm 0.03$            | $\pm 0.03$       | $\pm 0.03$ | $\pm 0.03$ |



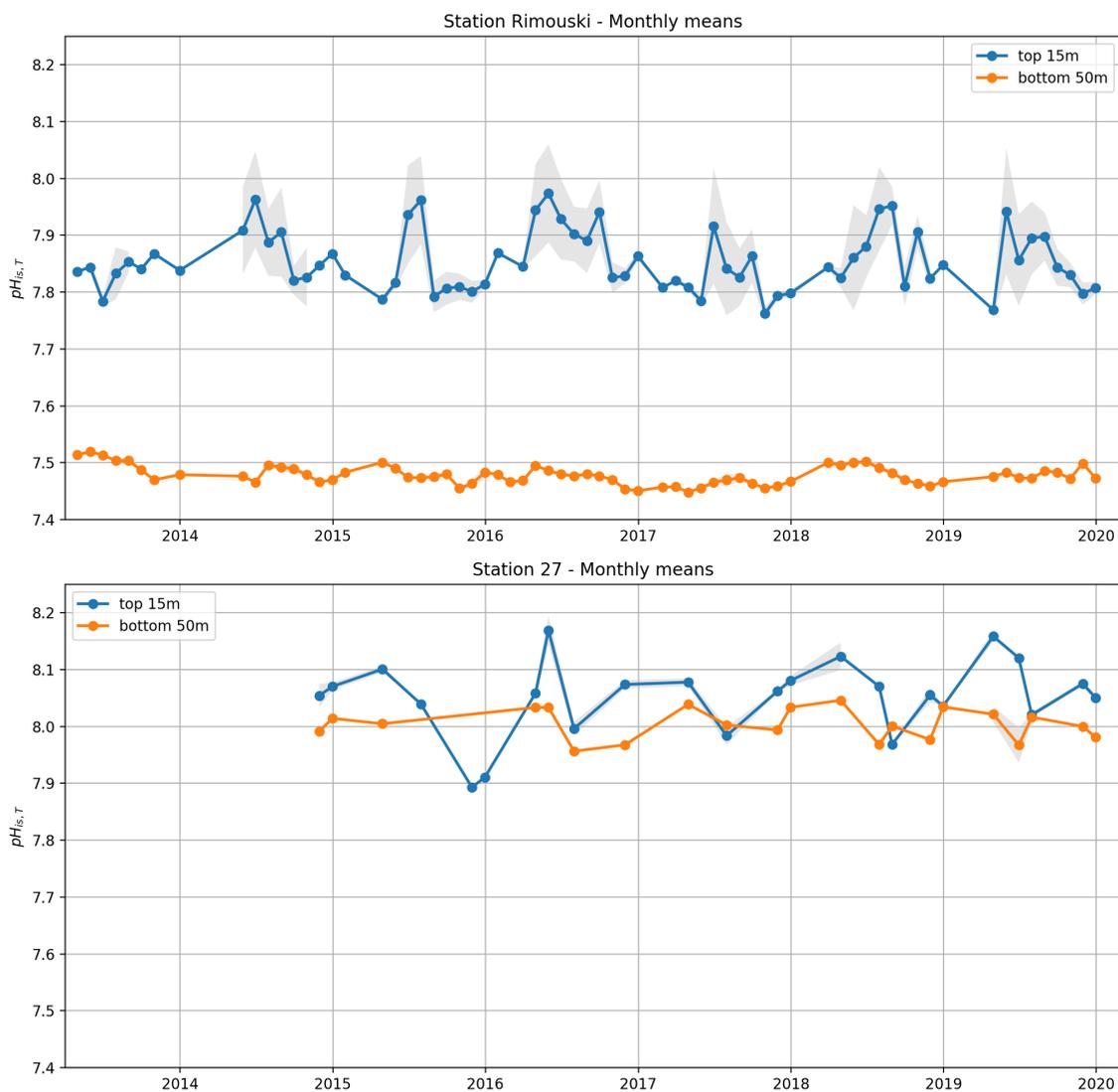
**Figure 1.** Bathymetric map of the Northwest Atlantic which includes the Newfoundland and Labrador Shelves and Slopes, Scotian Shelf and Slope, Gulf of St. Lawrence, and the St. Lawrence Estuary as the Atlantic Zone. Also identified are the deep channels within the Gulf of St. Lawrence: the Laurentian, Anticosti and Esquiman Channels. The approximate position and direction of the major current systems and their shelf and slope components are illustrated as arrows: Gulf Stream (gray) and Labrador Current (black).



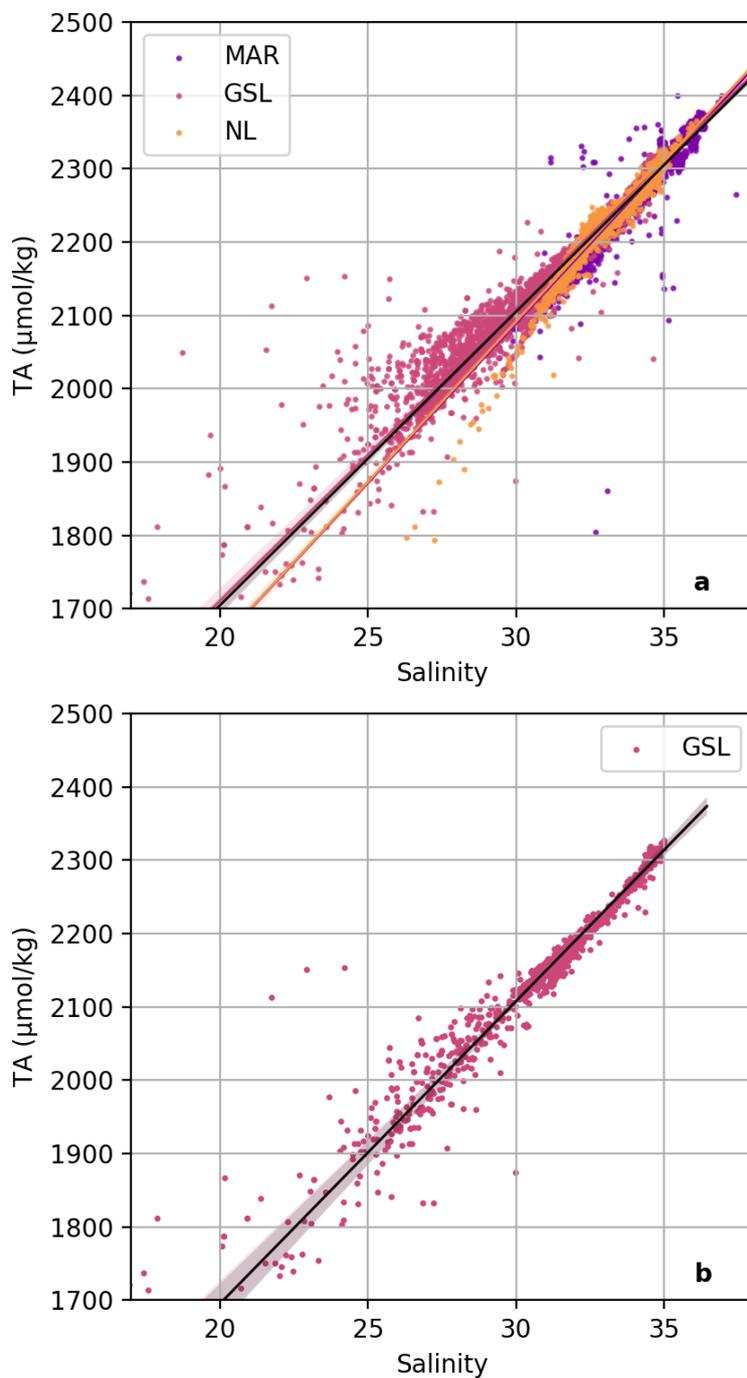
**Figure 2.** Diagrams of temperature - salinity versus depth (left) and temperature - salinity versus saturation state relative to aragonite for all bottle samples in the dataset. The three Regions of the Atlantic Zone are shown with different markers. An approximate division between the shelf and slope surface waters is indicated by the vertical black line at salinity 33. Various water masses are labeled (WSW = Warm Slope Water, NATIW = North Atlantic Water, NLSW = Newfoundland and Labrador Slope Water). In the absence of clear water mass definition, the origin of the water is labeled (LSLE = Lower St. Lawrence Estuary, GSL = Gulf of St. Lawrence, LS = Labrador Shelf).



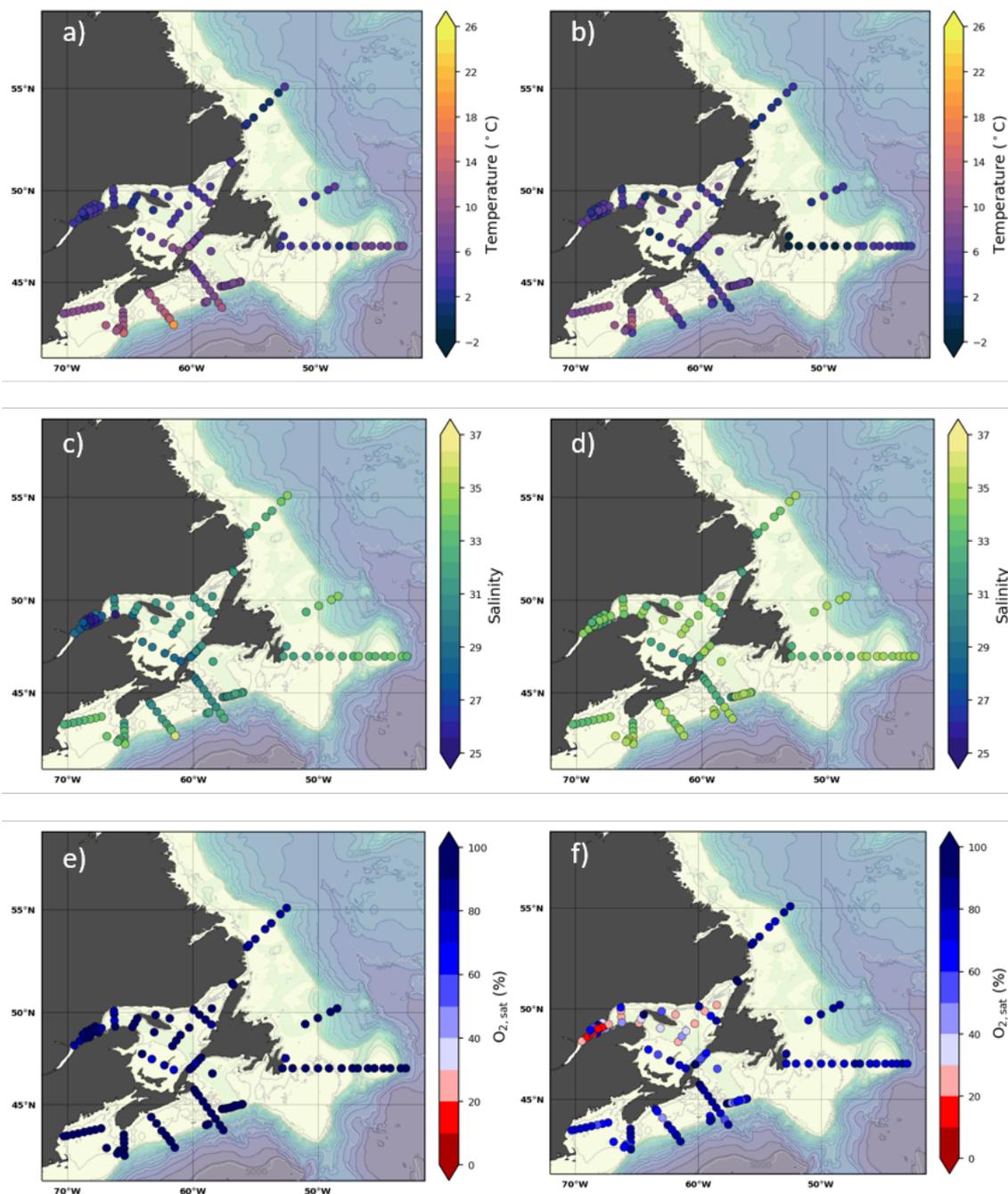
**Figure 3.** Bathymetric maps of the Atlantic Zone. AZMP surveys of the four Regions which include Maritimes (MAR), Newfoundland and Labrador (NL), and Québec (QC) and Gulf which have been combined as the Gulf of St. Lawrence (GSL) Region. The map of the Atlantic Zone in panel *a* shows the locations of the standard AZMP sections (from north to south and in the GSL): *Beachy Island* (BI), *Makkovik Bank* (MB), *Seal Island* (SI), *White Bay* (WB), *Bonavista Bay* (BB), *Flemish Cap* (FC), *Southeast Grand Bank* (SEGB), *Southeast St. Pierre Bank* (SESPB), *Southwest St. Pierre Bank* (SWSPB), *Cabot Strait Line* (CSL), *Louisbourg Line* (LL), *Halifax Line* (HL), *Browns Bank Line* (BBL), *Transect Bonne Bay* (TBB), *Transect Central* (TCEN), *Transect Iles de la Madeleine* (TIDM), *Transect Anticosti South* (TASO), *Transect Sept-Iles* (TSI), *Transect Estuaire du Saint-Laurent* (TESL) and *Laurentian Channel* (LC) from west to east across the entire GSL (e.g., Fig. A14). Panel *b* zooms in on the GSL Region and includes other sampling locations from in the dataset, such as the summer groundfish surveys. Here the Lower St. Lawrence Estuary (LSLE) correspond to the upstream portion of the GSL. The Upper St. Lawrence Estuary (USLE) and the Saguenay Fjord (SF) are also annotated.



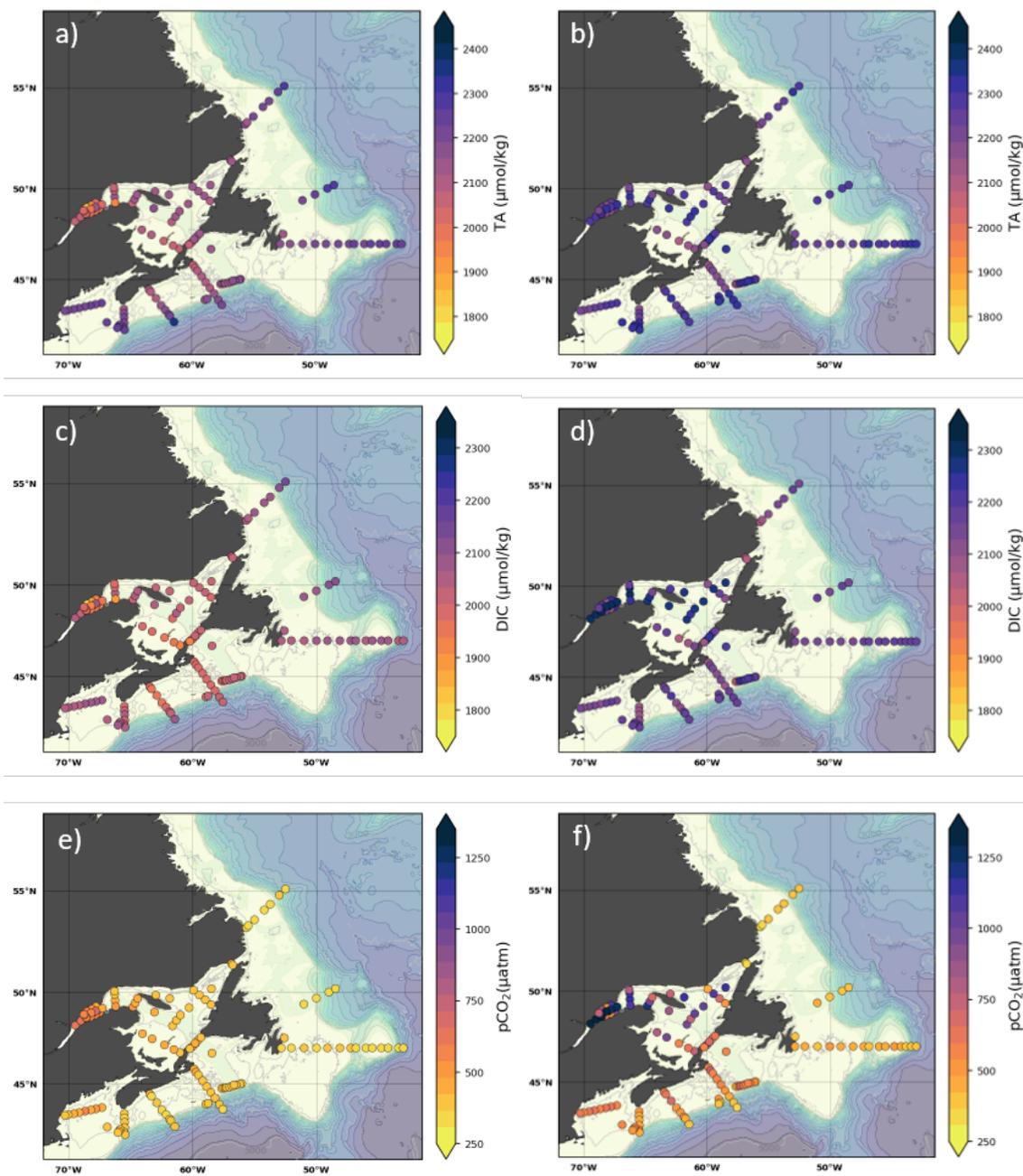
**Figure 4.** Monthly averages of pH data at Station Rimouski (top) and Station 27 (bottom). Surface observations (blue curves) are considered the average of the top 15m and bottom observations (orange curves) are the average of the bottom 50m. The shaded area correspond to  $\pm 0.5$  SD of the monthly distribution (when more than one visit per month is made). The tick marks on the horizontal axis correspond to the beginning of the year.



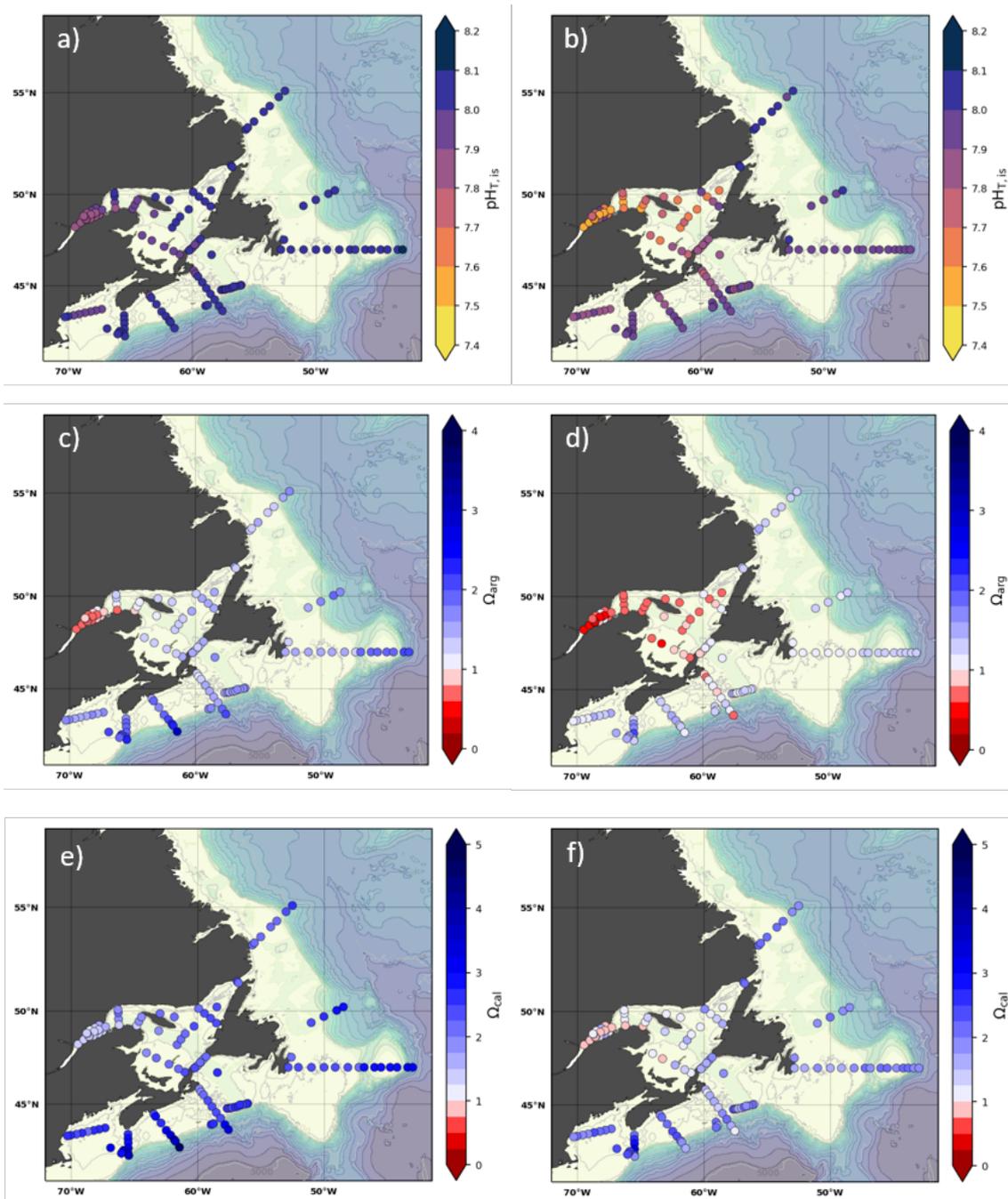
**Figure 5.** Scatter plot of total alkalinity (TA;  $\mu\text{mol/kg}$ ) versus salinity. a) all samples (2014 – 2020) have been established independently for each region (colored lines according to the legend) and for the entire dataset (black line). The latter TA and salinity relationship is  $\text{TA} = 40.05 \times \text{S} + 872.05$ ,  $r^2 = 0.94$ ,  $p < 0.001$ . b) A linear relationship is also established for the GSL spring data only. This relationship between TA and salinity ( $\text{TA} = 41.26 \times \text{S} + 870.05$ ,  $r^2 = 0.93$ ,  $p < 0.001$ ) is used in Section 3.3 to correct for missing TA data.



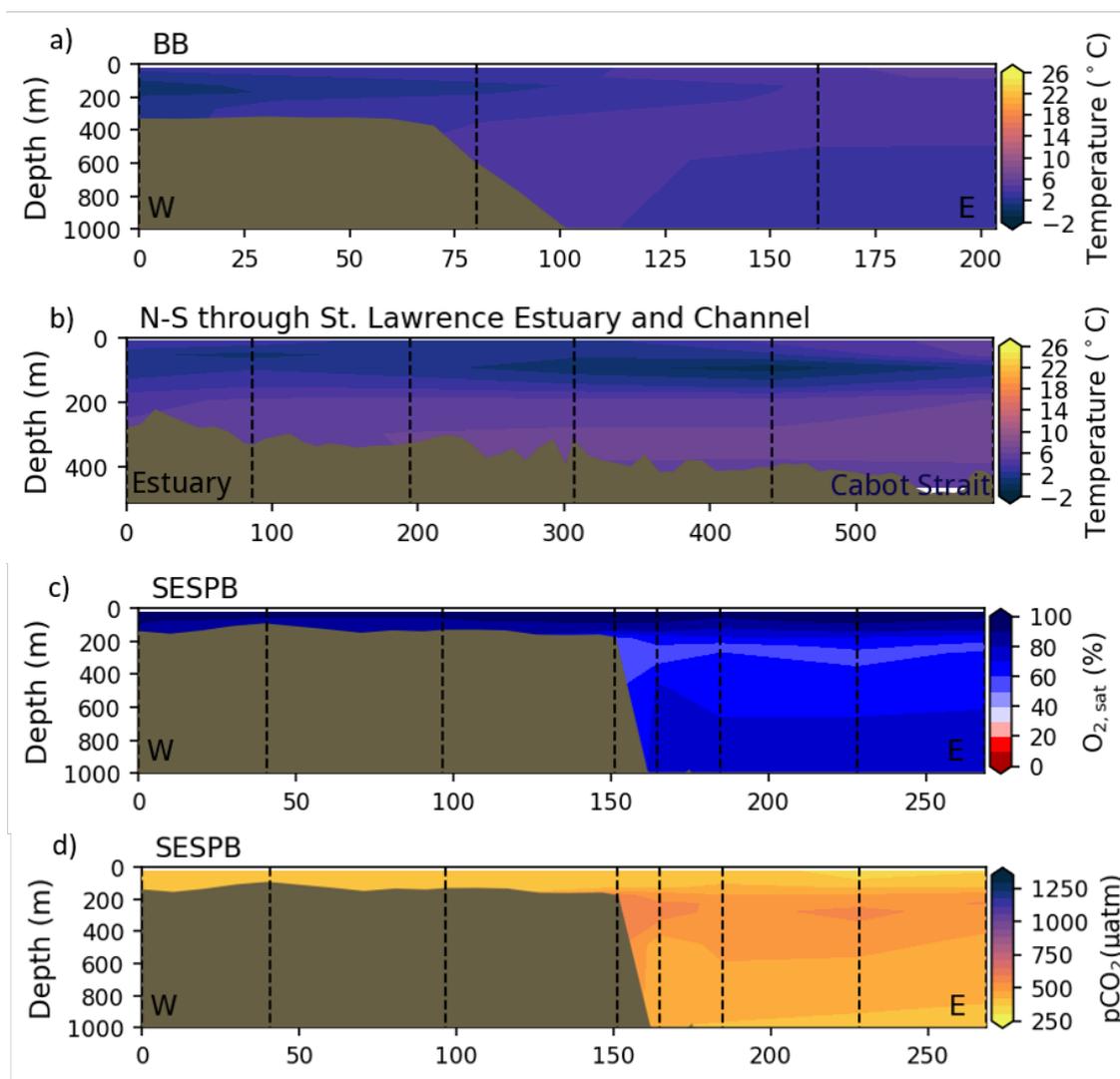
**Figure 6.** Maps of surface (upper most sample <52m; left side) and bottom (deepest sample >50m; right side) temperature (a, b), salinity (c, d) and oxygen saturation ( $O_{2,sat}$ ) (e, f) for fall 2017. The GSL surface  $O_{2,sat}$  is represented by samples at 50 m.



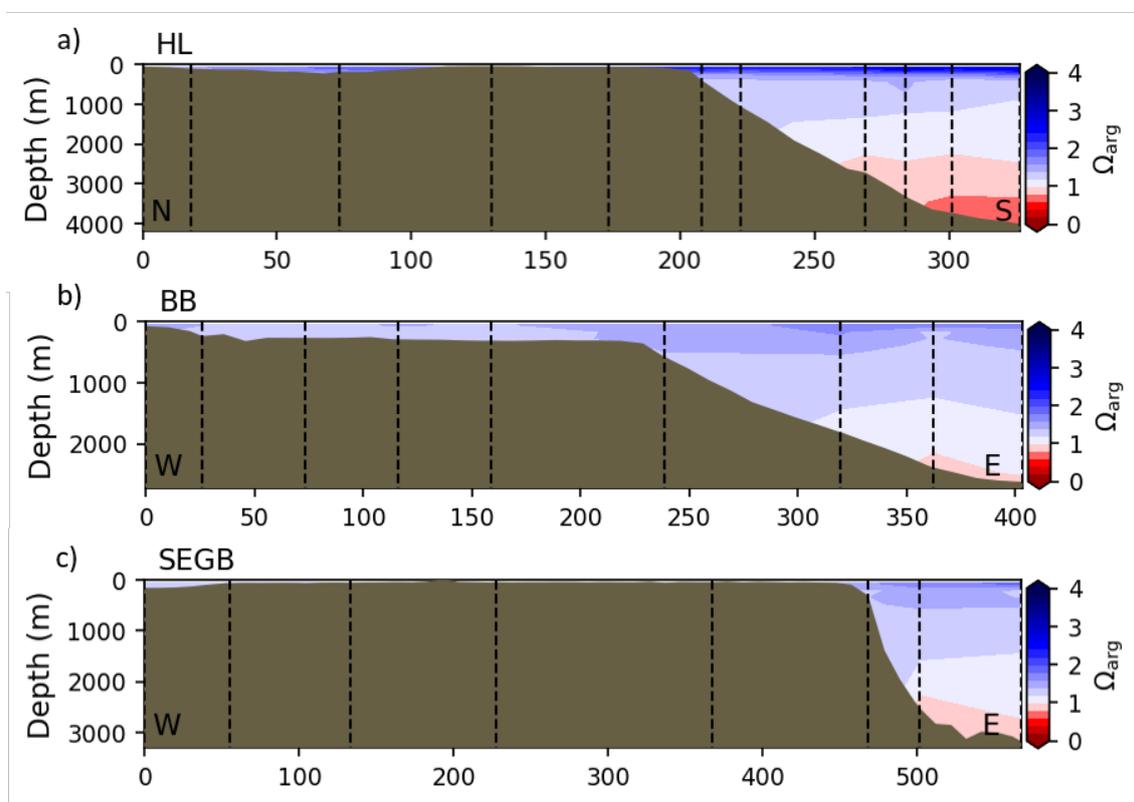
**Figure 7.** Maps of surface (upper most sample <52m; left side) and bottom (deepest sample <50m; right side) TA (a, b), DIC (c, d) and pCO<sub>2</sub> (e, f) for fall 2017.



**Figure 8.** Maps of surface (upper most sample <52m; left side) and bottom (deepest sample >50m; right side) pH (a, b),  $\Omega_{arg}$  (c, d) and  $\Omega_{cal}$  (e, f) for fall 2017.



**Figure 9.** Depth-distance (x-axis is the distance in km from first station) contours along standard sections in the Atlantic Zone. Sections a) and b) show the Cold Intermediate Layer (CIL) in fall 2017 with temperature ( $^{\circ}\text{C}$ ) along BB from west to east (a) and through the GSL north to south along the St. Lawrence Estuary and St. Lawrence Channel (b). Sections c) and d) present the oxygen minimum and  $\text{pCO}_2$  maximum off the southern Newfoundland Slope along the SESP section in spring 2017. The dashed-lines represent to positions of the casts where the samples are collected.



**Figure 10.** Depth profiles along standard sections in the Atlantic Zone for the fall of 2017. The aragonite saturation horizon ( $\Omega_{arg} = 1$ ) is located between  $\sim 2200$  and  $2300$  m along sections HL (a), BB (b), SEGB (c) above the Scotian and NL Slopes.

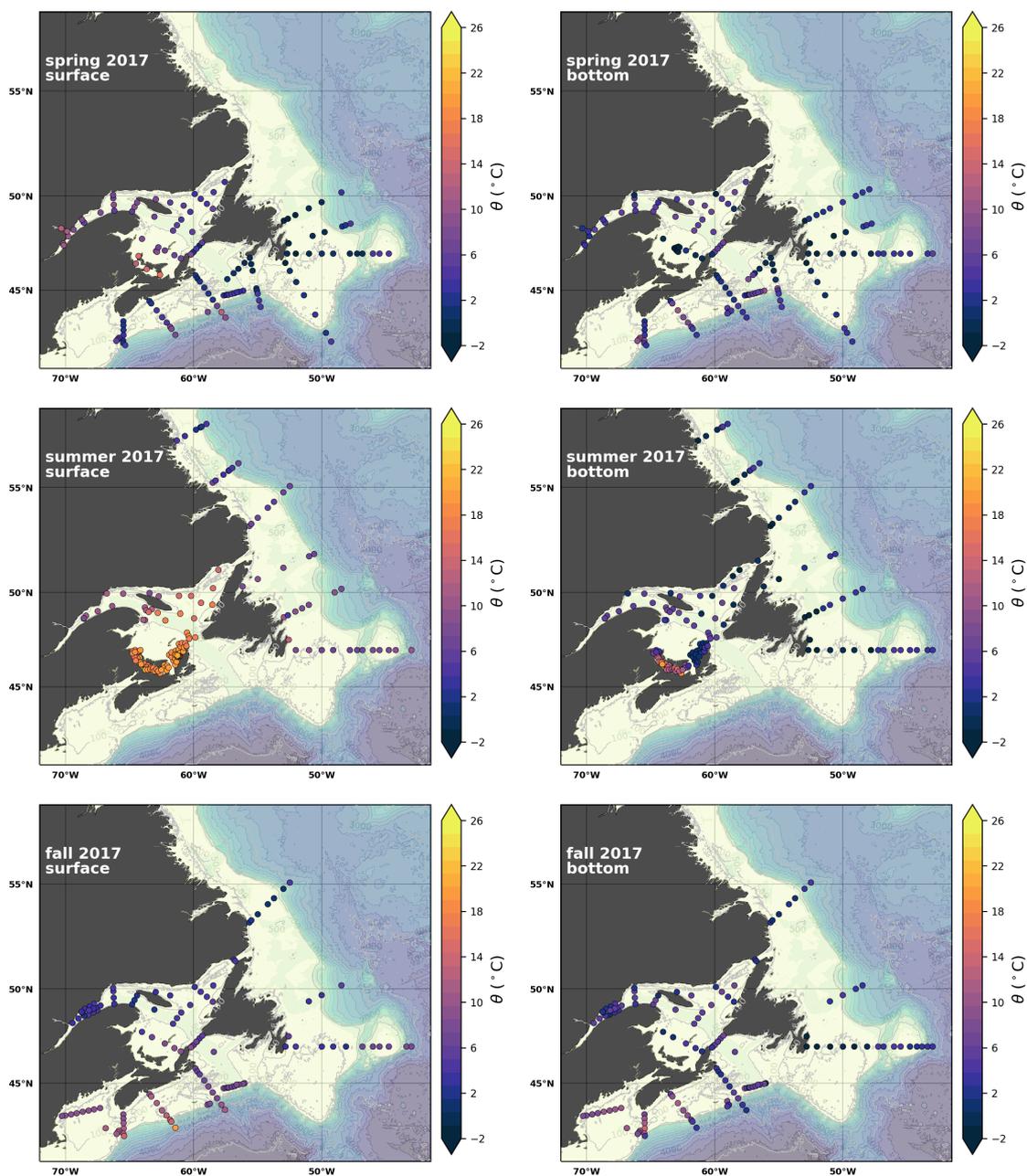


### **Appendix A: Seasonal maps of various parameters during 2017.**

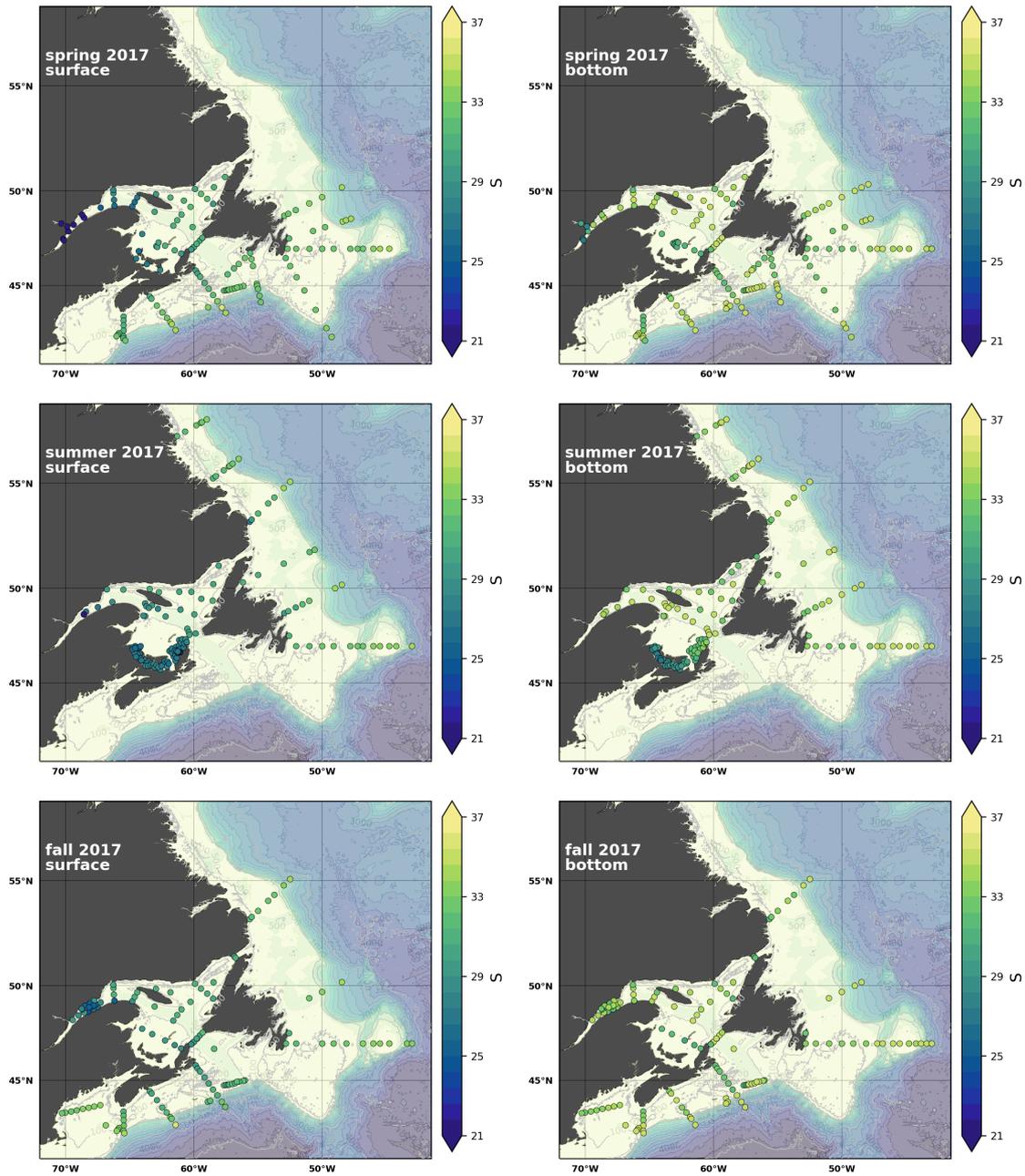
This Appendix presents a series of seasonal maps (spring, summer and fall) of various physico-chemical parameters collected as part of the AZMP during 2017, one of the most complete year of sampling. These parameters are: Temperature, Salinity,  $O_{2sat}$ , TA, TIC,  $pCO_2$ , pH,  $\Omega_{arg}$  and  $\Omega_{cal}$ . They are presented in Figures A1 to A9, respectively. Each figure has 6 panels where the top row is the surface (upper most sample <52m) and the bottom row the near bottom (deepest sample >50m) conditions. The spring, summer and fall seasons are presented from left to right.

### **Appendix B: Hydrographic Sections**

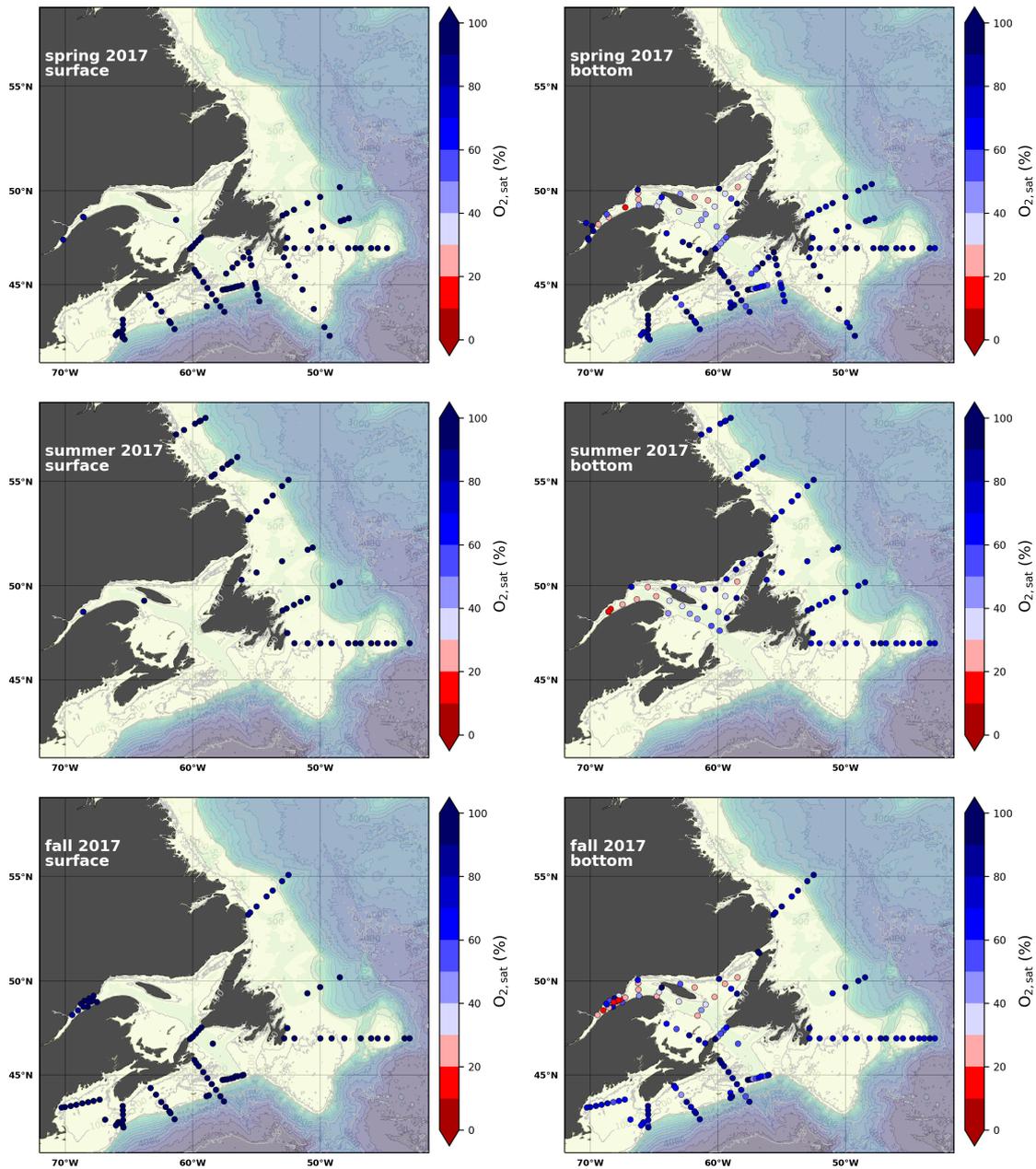
Figures A10 to A14 show contour plots of various physico-chemical parameters collected as part of the AZMP (the same as in Appendix A) along selected hydrographic sections of the Atlantic Zone during the fall of 2017 (see Figure 3 for location). These sections are Seal Island (SI), Flemish cap (FC), Halifax (HL), Cabot Strait (CSL) and Laurentian Channel (LC) and are presented in Figures A10 to A14, respectively. Each figure shows, from top to bottom panels, the Temperature, Salinity,  $O_{2sat}$ , TA, TIC,  $pCO_2$ , pH,  $\Omega_{arg}$  and  $\Omega_{cal}$ , respectively.



**Figure A1.** Surface (upper most sample <52m; top row), and near-bottom (deepest sample >50m; bottom row) temperature sampled during the spring (left), summer (middle) and fall (right) AZMP surveys.



**Figure A2.** Same as in Figure A1, but for salinity.



**Figure A3.** Same as in Figure A1, but for  $O_{2,sat}$ .

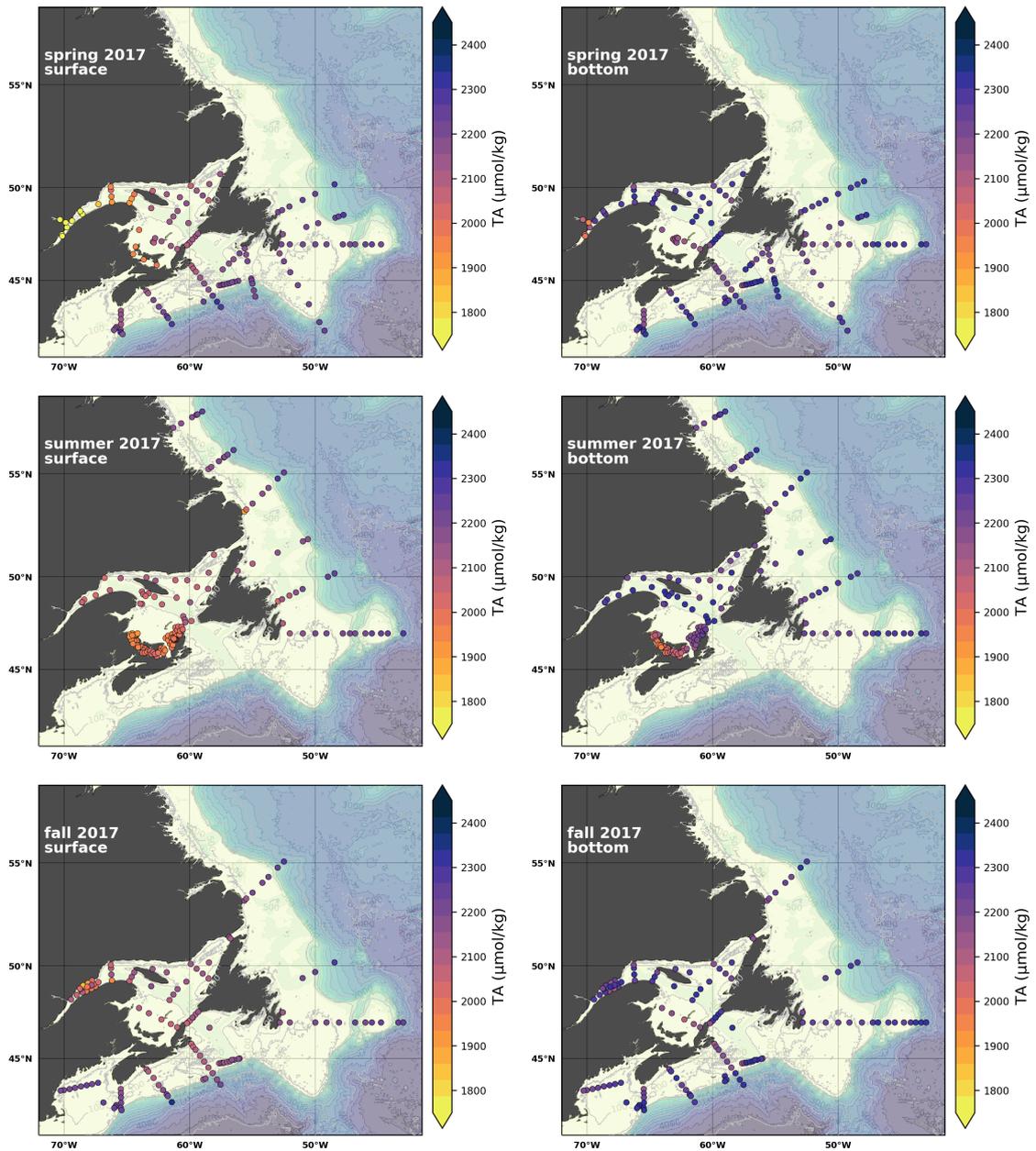


Figure A4. Same as in Figure A1, but for TA.

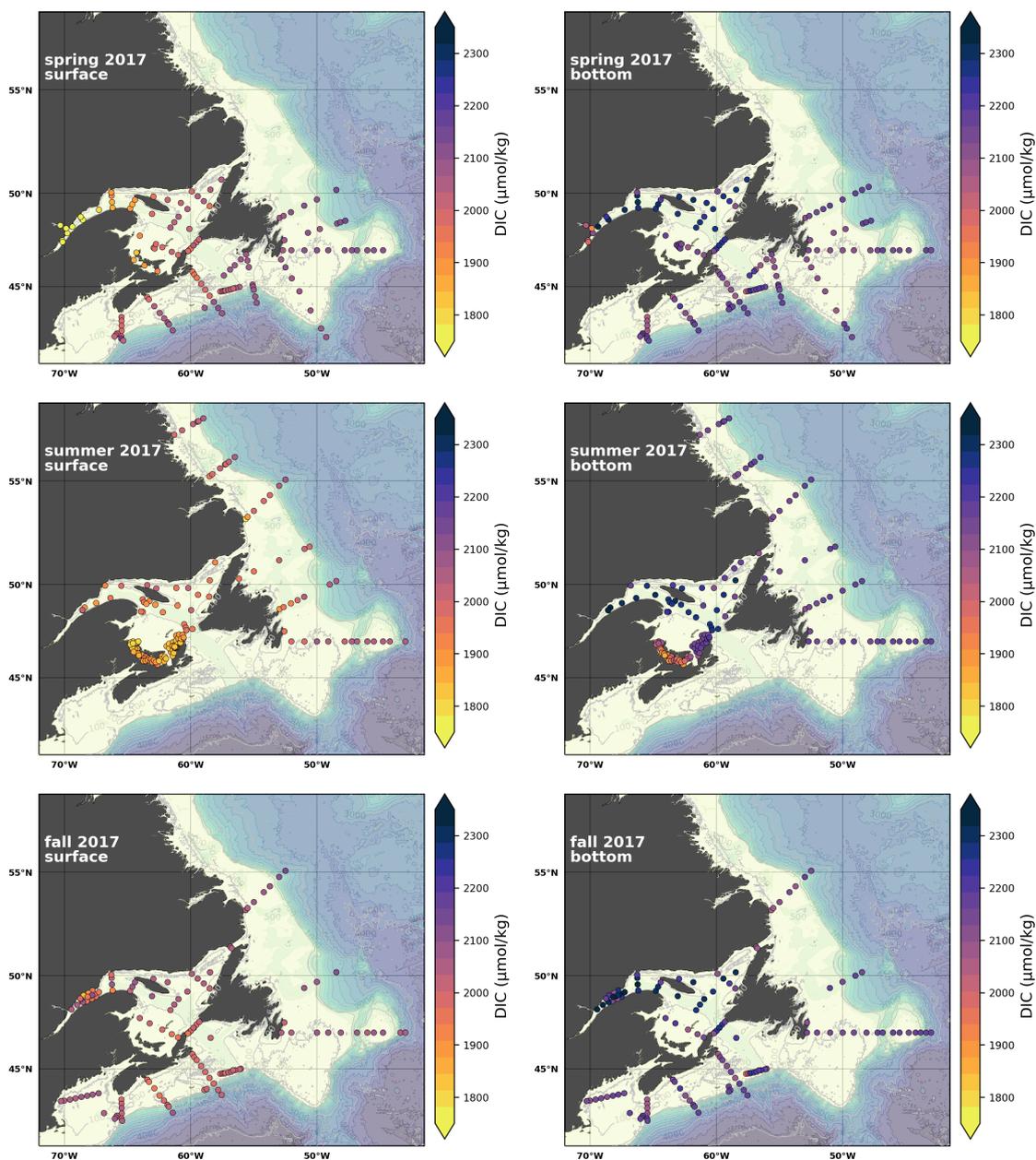


Figure A5. Same as in Figure A1, but for TIC.

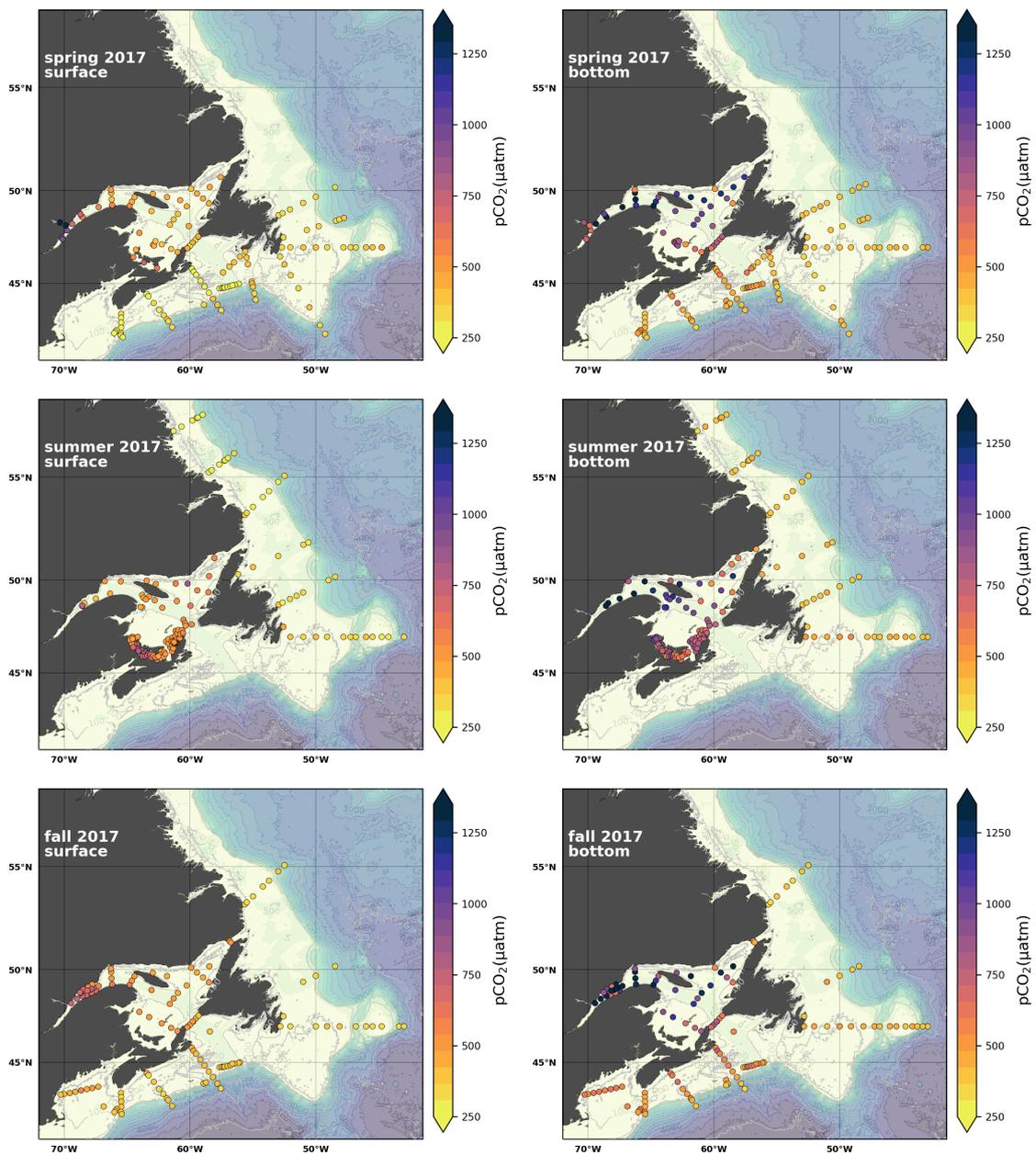


Figure A6. Same as in Figure A1, but for pCO<sub>2</sub>

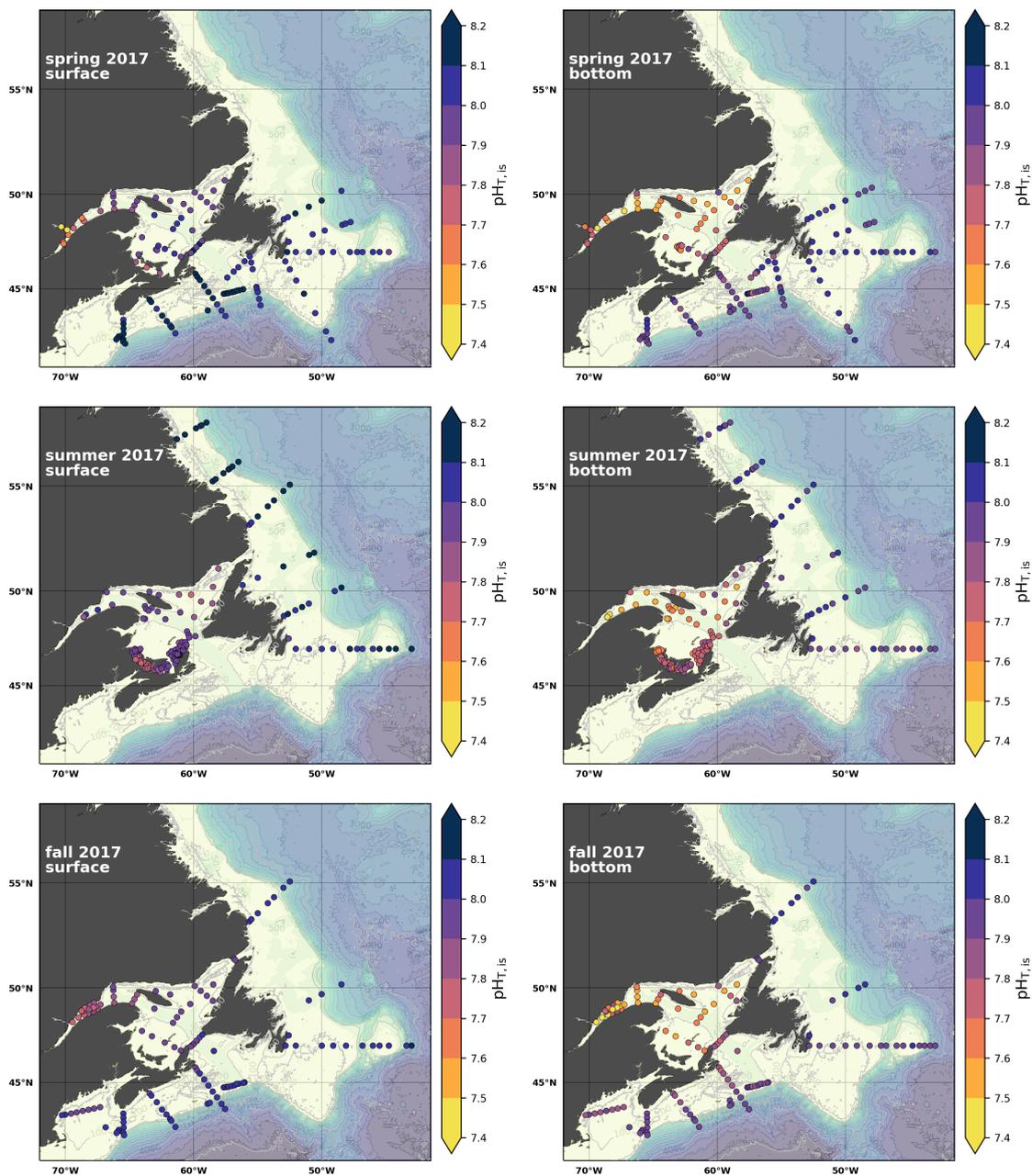


Figure A7. Same as in Figure A1, but for pH.

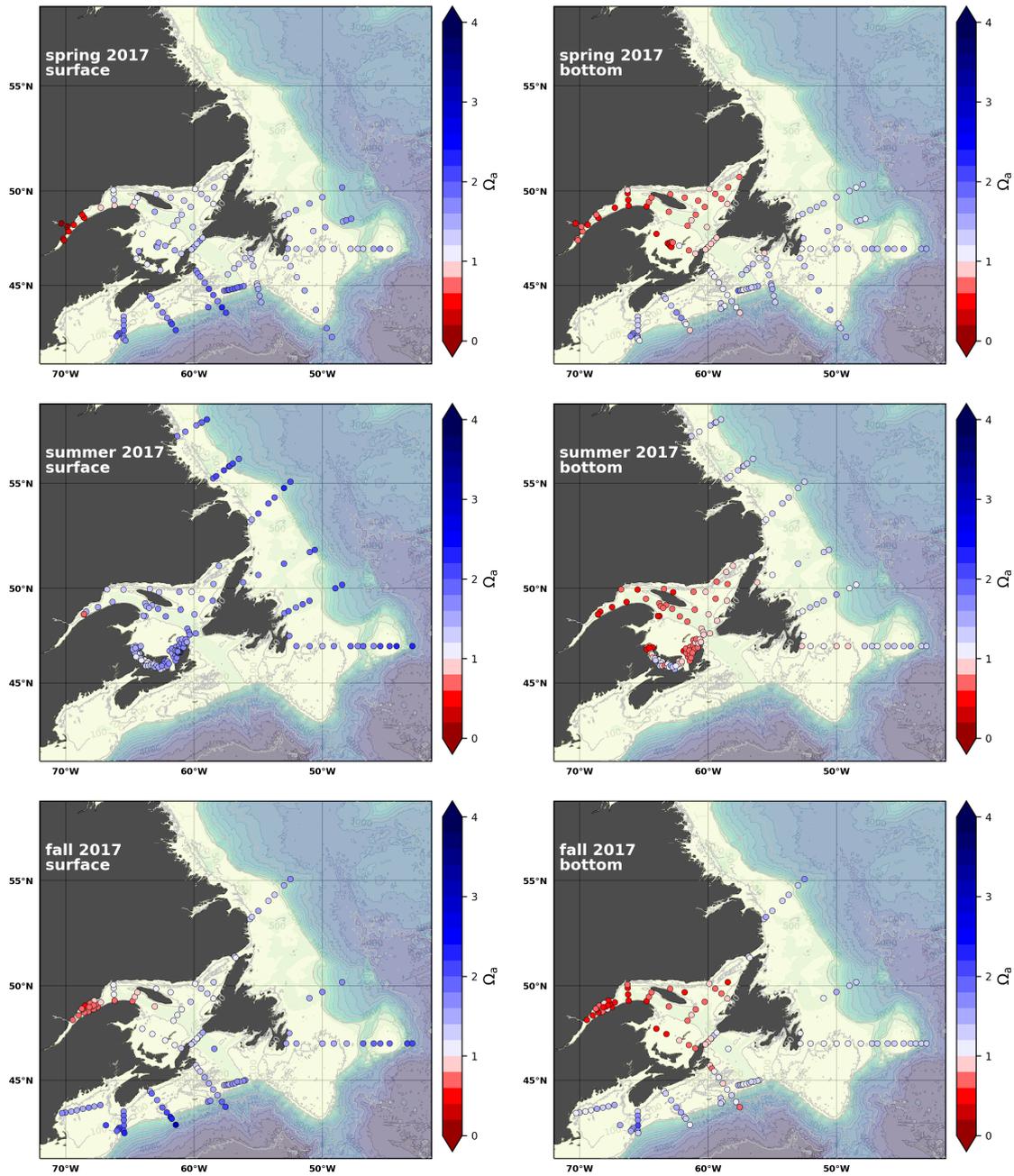


Figure A8. Same as in Figure A1, but for  $\Omega_{arg}$ .

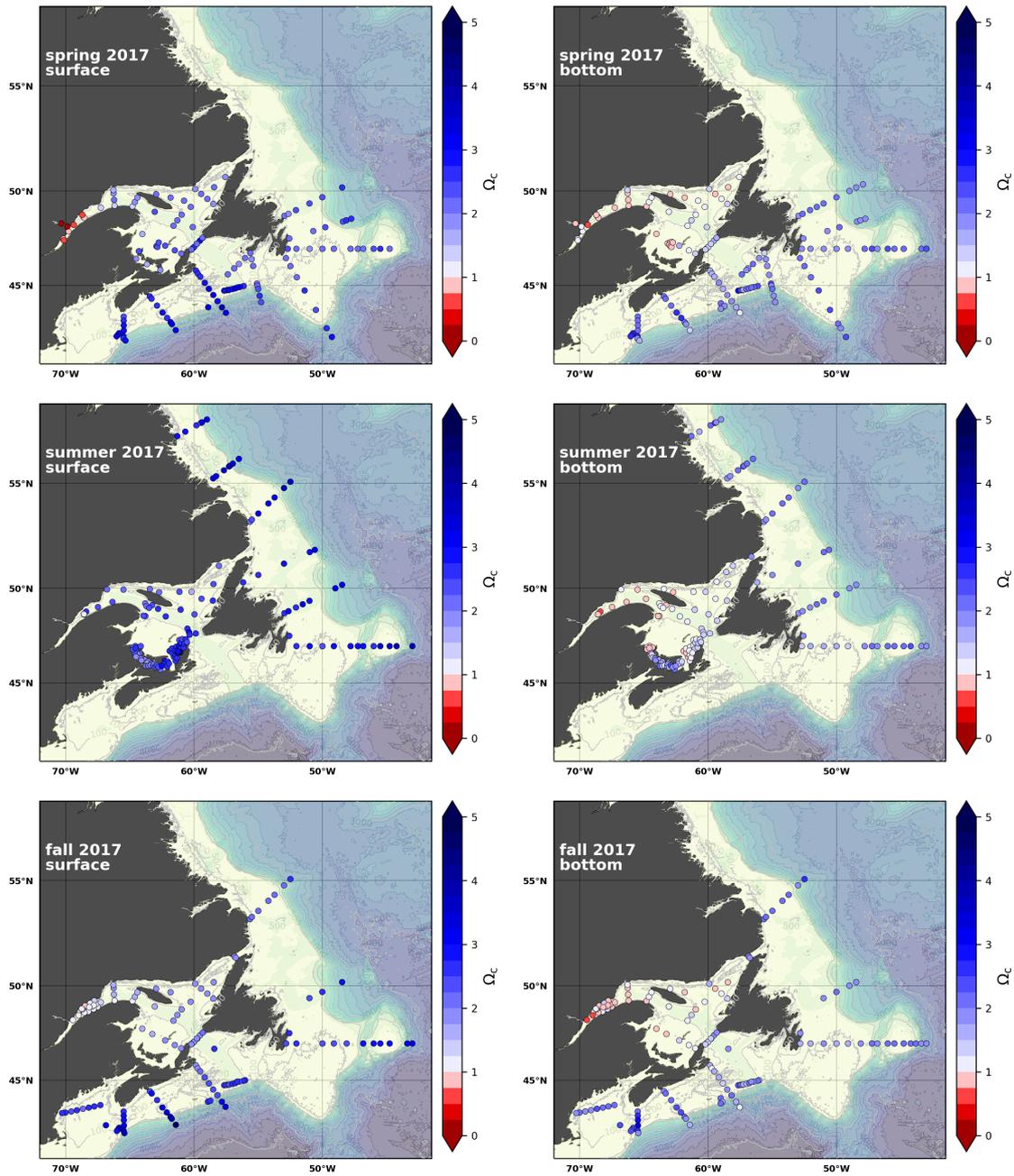
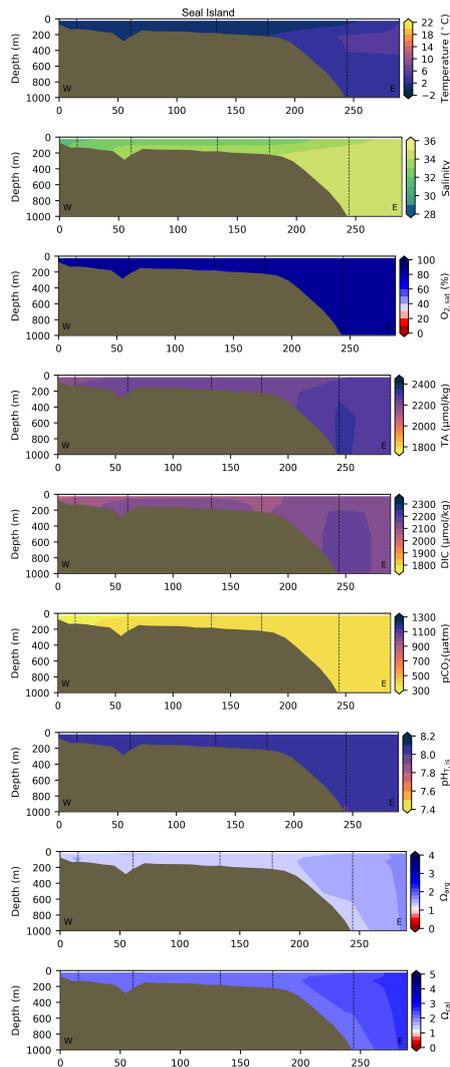
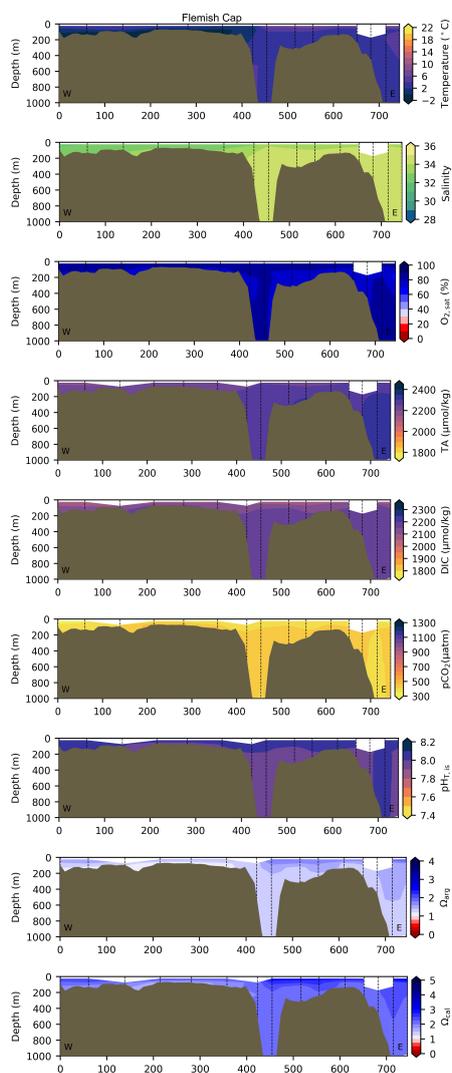


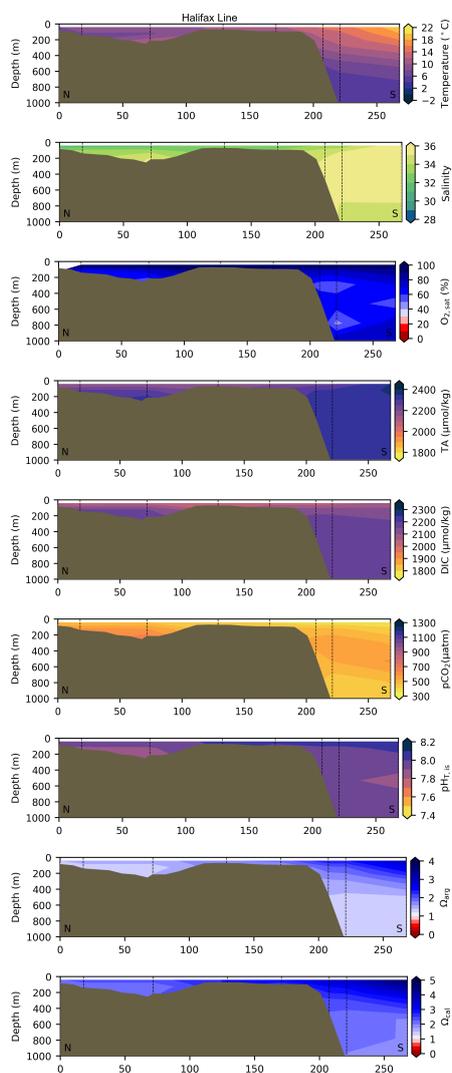
Figure A9. Same as in Figure A1, but for  $\Omega_{cal}$ .



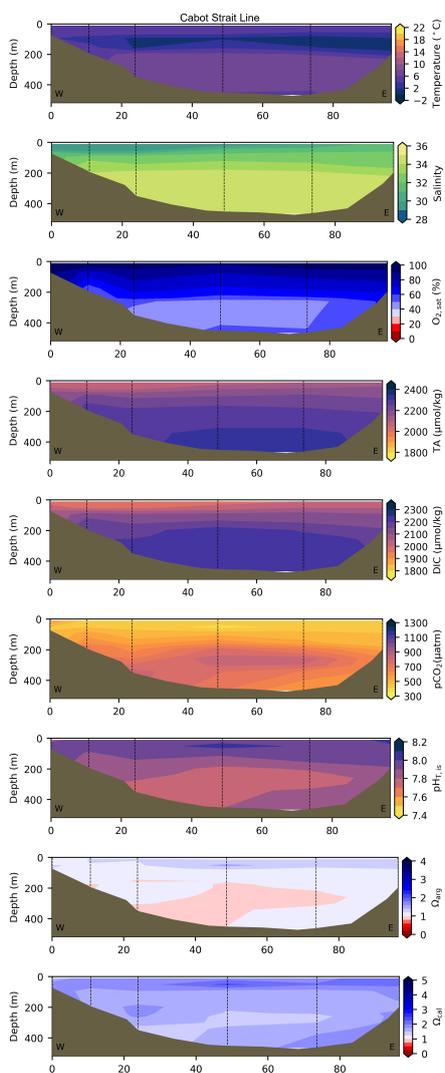
**Figure A10.** Contour plots of various physico-chemical parameters collected during the fall 2017 AZMP survey along hydrographic section Seal Island (NL region, see Figure 3 for location). From top to bottom, these parameters are respectively: Temperature, Salinity,  $O_{2,sat}$ , TA, TIC,  $pCO_2$ , pH,  $\Omega_{arg}$  and  $\Omega_{cal}$ .



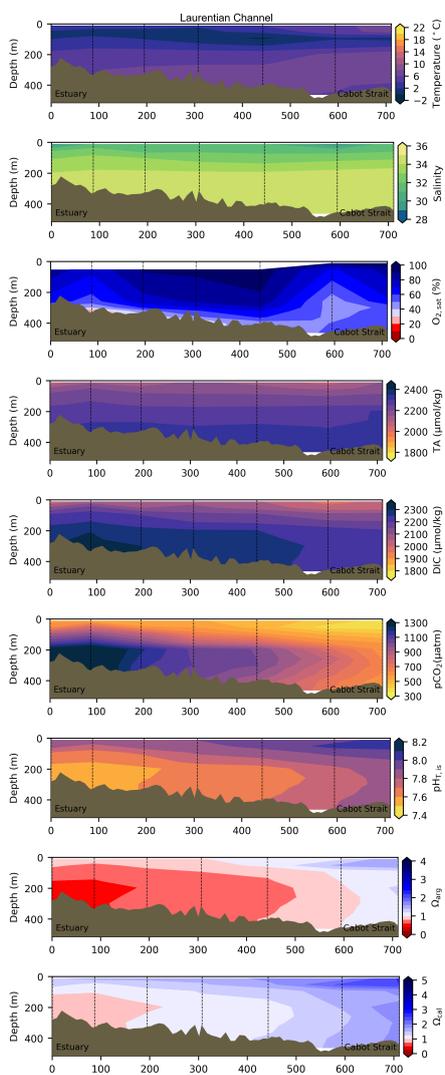
**Figure A11.** Same as in Figure A10, but for hydrographic section Flemish Cap (NL region, see Figure 3 for location).



**Figure A12.** Same as in Figure A10, but for hydrographic section Halifax (MAR region, see Figure 3 for location).



**Figure A13.** Same as in Figure A10, but for hydrographic section Cabot Strait (GSL region, see Figure 3 for location).



**Figure A14.** Same as in Figure A10, but for hydrographic section Laurentian Channel (GSL region, see Figure 3 for location).