



1 **Hyperspectral reflectance dataset of pristine, weathered and biofouled** 2 **plastics**

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14 **Abstract**

15 This work presents a hyperspectral reflectance dataset of macroplastic samples acquired using
16 Analytical Spectral Devices (ASD) FieldSpec 4. Samples analysed consisted of pristine, artificially
17 weathered and biofouled plastic items and plastic debris samples collected in the docks of the Port of
18 Antwerp and in the river Scheldt near Temse Bridge (Belgium). The hyperspectral signal of each sample
19 was measured in controlled dry conditions in an optical calibration facility at the Flemish Institute for
20 Technological Research, and, for a subset of plastics, under wet and submerged conditions in a silo tank
21 at Flanders Hydraulics. The wet and submerged hyperspectral signals were measured in a mesocosm
22 setting that mimicked environmentally relevant concentrations of freshwater microalgae and of
23 suspended sediment. The ASD was equipped with an 8° field of view at the calibration facility, and a 1°
24 field of view was used in the mesocosm setting. The dataset obtained complies with the Findability,
25 Accessibility, Interoperability, and Reuse (FAIR) principles and is available in the open-access repository
26 Marine Data Archive (<https://doi.org/10.14284/530>, Leone et al., 2021).



27 **1 Introduction**

28 The spectral reflectance measurements, collected in the framework of the Plastic Flux for Innovation and
29 Business Opportunities in Flanders (PLUXIN) project, contribute to the current knowledge on the
30 detection of plastics using remote sensing techniques.

31 In recent years, focus has been placed on the detection of plastic litter using remote sensing techniques
32 such as optical sensors on satellites, aircrafts, and drones (Garaba and Dierssen, 2018; Martínez-
33 Vicente et al., 2019). With the increasing demand of these technologies, it is crucial to generate
34 knowledge on the diagnostic spectral properties of not only pristine, but also weathered and biofouled
35 plastics (Moshtaghi et al., 2021) that are representative of the variety of environmental plastics.

36 Currently the spectral reflectance of dry plastics is known, and is already applied in the field of material
37 recycling (Moroni et al., 2015), but it is restricted to items assessed for dry measurements. To be able to
38 identify plastic litter in aquatic environments such as rivers, harbours, and oceans, we require the
39 acquisition of the spectral features when plastics are either wet or submerged (Moshtaghi et al., 2021),
40 as water absorbs the light in both the near (NIR) and shortwave infrared (SWIR). In addition, other water
41 constituents such as sediment or algae, could further impact the reflected signal of plastic items
42 (Moshtaghi et al., 2021).

43 To date, only a limited number of high-quality datasets consisting of hyperspectral measurements of wet
44 and submerged plastic litter, have been published in open-access repositories (e.g., Garaba and
45 Dierssen, 2019, Garaba and Dierssen, 2020; Knaeps et al., 2021). The dataset described in the current
46 paper aims at complementing the existing datasets by adding new information about the hyperspectral
47 reflectance of pristine plastic items, harvested plastic litter, and artificially weathered and biofouled plastic
48 samples. In addition, the dataset reports the optical features of plastics acquired in various water turbidity
49 conditions obtained by adding sediment or algae to the water, at selected concentrations.

50 The dry spectral reflectance of plastic specimens, consisting of different polymers, was measured using
51 an Analytical Spectral Device (ASD) FieldSpec 4. For a selection of these samples, the spectral
52 reflectance was also collected in wet and submerged settings.

53 The presented data can be used to generate insights on the spectral properties of plastic litter and how
54 these features change when plastics are exposed to natural agents or different depths. The dataset was



55 compiled following the FAIR (Findable, Accessible, Interoperable, Reusable; go-fair.org) principles and
56 it is publicly available at <https://doi.org/10.14284/530>.

57

58 **2. Data collection**

59 Data collection consisted in measuring the spectral reflectance of different plastic specimens using the
60 ASD. The measurements were conducted in two different settings: at an optical calibration facility located
61 at the Flemish Institute for Technological Research (Mol, Belgium), and at a silo tank at the Flanders
62 Hydraulics Research facility (Antwerp, Belgium). The spectral reflectance of each plastic specimen was
63 collected at least 5 times (“pseudo-replicates”) within 1 minute. These pseudo-replicates are
64 measurements taken by slightly changing the position of the plastic sample. Therefore, very
65 homogeneous plastics will have less variation in the pseudo-replicates’ reflectance.

66



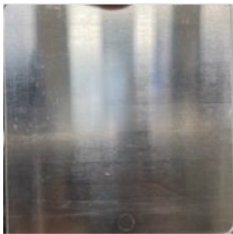
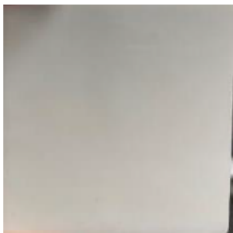

67 **2.1 Plastic specimens**

68 We aimed to collect the spectral reflectance of plastic items as representative as possible of commonly
69 found plastics in the environment (LI et al., 2016), and to provide additional information on their
70 reflectance when exposed to natural agents such as sunlight (UV radiation) or biofilm growth. To do so,
71 we have selected and analysed a total of 10 polymer types (Table 1). The selected polymers were
72 Polyethylene (PE), Polypropylene (PP), Polystyrene (PS), and two types of Polyethylene terephthalate
73 (PET), crystalline and amorphous, Fluorocarbon, Thermoplastic elastomer, Polyvinyl chloride (PVC),
74 Nylon 6 (PA6) and Paraffin. In addition, we have further artificially weathered and biofouled a selection
75 of 6 plastic polymer types. The polymer discrimination was based on the available information from the
76 supplier or marked on the plastic itself (e.g., plastic bottle or bags with an identifiable polymer tag). In
77 addition, for a set of samples, we confirmed the polymer type using a micro-Fourier Transform Infrared
78 Spectroscopy (μ FTIR, PerkinElmer, FTIR spectrometer Frontier). The percentage of matching score with
79 a library was recorded and it is reported in the dataset.

80

81 **Table 1.** Overview of the plastic specimens analysed in the current work. Polymer type refers to the available information from
82 the supplier or product, unless stated otherwise. Selected polymer types were confirmed by μ FTIR. Pictures in this table are
83 taken by Giulia Leone and Mattias Bossaer.





Type	Plastic specimen	Condition of the assessment	Picture	Polymer type	Available measurements (d: dry, w: wet, s: submerged)	Dimensions (cm)
Commercial plastics	Sheet	Dry		PE (confirmed by μ FTIR)	d, w, s	6 x 6
	Sheet	Dry		PP (confirmed by μ FTIR)	d, w, s	6 x 6
	Sheet	Dry		PET amorphous (confirmed by μ FTIR)	d, w, s	6 x 6
	Sheet	Dry		PET crystalline (confirmed by μ FTIR)	d, w, s	6 x 6
	Sheet	Dry		PS (confirmed by μ FTIR)	d, w, s	6 x 6









Weathered plastics from shops	Sheet	Dry		PVC (Thermoplastic elastomer under μ FTIR)	d	2 x 4
	Sheet	Dry		PP (confirmed by μ FTIR)	d	2 x 4
	Sheet	Dry		PS (confirmed by μ FTIR)	d	2 x 4
Biofilm plastics from shops	Sheet	Dry		PVC (Thermoplastic elastomer under μ FTIR)	d	2 x 10
	Sheet	Dry		PP (confirmed by μ FTIR)	d	2 x 10
	Sheet	Dry		PS (confirmed by μ FTIR)	d	2 x 10


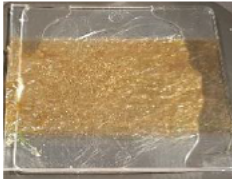







Plastic from shops	Sheet	Dry	ND	PVC (Thermoplastic elastomer under μ FTIR)	d	2 x 10
	Sheet	Dry	ND	PP (confirmed by μ FTIR)	d	2 x 10
	Sheet	Dry	ND	PS (confirmed by μ FTIR)	d	2 x 10
	Spar bag	Dry		LDPE (paraffin under μ FTIR)	d	ND
	Jumbo bag	Dry		ND	d	ND
	Bottle	Dry		ND	d	ND










Biofilm commercial plastics	Sheet	Wet rough		PP (confirmed by μ FTIR)	d, w, s	6 x 6
	Sheet	Wet smooth		PP (confirmed by μ FTIR)	d, w, s	6 x 6
	Sheet	Wet rough		PE (confirmed by μ FTIR)	d, w, s	6 x 6
	Sheet	Wet smooth		PE (confirmed by μ FTIR)	d, w, s	6 x 6
	Sheet	Wet rough		PS (confirmed by μ FTIR)	d, w, s	6 x 6
	Sheet	Wet smooth		PS (confirmed by μ FTIR)	d, w, s	6 x 6



Sheet	Wet rough		PET Amorphous (confirmed by μ FTIR)	d, w, s	6 x 6
Sheet	Wet smooth		PET amorphous (confirmed by μ FTIR)	d, w, s	6 x 6
Sheet	Wet rough		PET Crystalline (confirmed by μ FTIR)	d, w, s	6 x 6
Sheet	Wet smooth		PET crystalline (confirmed by μ FTIR)	d, w, s	6 x 6
Sheet	Dry rough		PS (confirmed by μ FTIR)	d	6 x 6
Sheet	Dry smooth		PS (confirmed by μ FTIR)	d	6 x 6
Sheet	Dry rough		PE (confirmed by μ FTIR)	d	6 x 6



Sheet	Dry smooth		PE (confirmed by μ FTIR)	d	6 x 6
Sheet	Dry rough		PET amorphous (confirmed by μ FTIR)	d	6 x 6
Sheet	Dry smooth		PET amorphous (confirmed by μ FTIR)	d	6 x 6
Sheet	Dry rough		PP (confirmed by μ FTIR)	d	6 x 6
Sheet	Dry smooth		PP (confirmed by μ FTIR)	d	6 x 6
Sheet	Dry rough		PET crystalline (confirmed by μ FTIR)	d	6x6
Sheet	Dry smooth		PET crystalline (confirmed by μ FTIR)	d	6x6

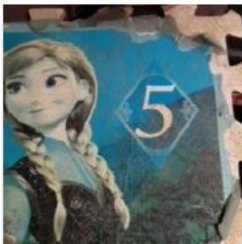

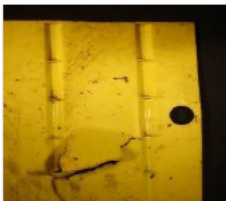





Artificially weathered	Sheet	Dry		PS (confirmed by μ FTIR)	d, w, s	2 x 4
	Sheet	Seawater		PS (confirmed by μ FTIR)	d, w, s	2 x 4
	Sheet	Dry		PE (confirmed by μ FTIR)	d, w, s	2 x 4
	Sheet	Seawater		PE (confirmed by μ FTIR)	d, w, s	2 x 4
	Sheet	Dry		PP (confirmed by μ FTIR)	d, w, s	2 x 4
	Sheet	Seawater		PP (confirmed by μ FTIR)	d, w, s	2 x 4
	Sheet	Dry		PET amorphous (confirmed by μ FTIR)	d, w, s	2 x 4






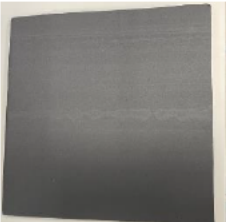


	Sheet	Seawater		PET amorphous (confirmed by μ FTIR)	d, w, s	2 x 4
	Sheet	Dry		PET crystalline (confirmed by μ FTIR)	d, w, s	2 x 4
	Sheet	Seawater		PET crystalline (confirmed by μ FTIR)	d, w, s	2 x 4
Real samples	Green Buoy	Real		PP (confirmed by μ FTIR)	d, w, s	ND
	Milk bottle	Real		HDPE (confirmed by μ FTIR)	d, w, s	ND
	Foam	Real		PS	d	ND








Toy placemat	Real		PP – PE (confirmed by μ FTIR)	d, w, s	ND
Green Bottle	Real		PET (confirmed by μ FTIR)	d, w, s	ND
Yellow sheet	Real		HDPE (confirmed by μ FTIR)	d	ND
Bag dog food	Real		PE (confirmed by μ FTIR)	d	ND
Rope	Real		PP (confirmed by μ FTIR)	d	ND
Shoe	Real		Fluorocarbon (confirmed by μ FTIR)	d	ND



	Bubble wrap	Real		PE (confirmed by μ FTIR)	d	ND
	Foam	Real		PE (confirmed by μ FTIR)	d	ND
	Red sheet	Real		PP (confirmed by μ FTIR)	d	ND
	Cup	Real		PP (confirmed by μ FTIR)	d	ND
The Ocean Cleanup samples	HDPE	Pristine		PE	d	30x30
	Extruded PS	Pristine		Extruded PS	d	30x30



PP	Pristine		PP	d	30x30
PET	Pristine		PET	d	30x30
PVC	Pristine		PVC	d	30x30
PA6	Pristine		PA6	d	30x30
Wood	Real		NA	d	NA

84

85

86 2.2.1 Pristine plastic specimens

87 To enhance the physio-chemical properties of an end plastic product, plastic manufacturers often
88 incorporate additive substances into the pristine polymer (Garaba et al., 2021). Additives included in
89 plastics, such as coloured plastics, may be influencing the spectra (Garaba et al., 2021). Thus, to avoid
90 confounding measurements, sheets of 6 x 6 cm were obtained from a commercial supplier (Carat,



91 Germany, <https://www.carat-lab.com>) as pure pristine plastic polymers without any additional additive
92 for the following polymers: PP, PE, PS, PET amorphous, PET crystalline. In addition, no colour or stains
93 were added to the commercial plastics, and as such, these selected samples were transparent or white.
94 The other pristine plastic samples were either purchased from a local shop (Oostende, Belgium), or
95 obtained from other institutes (Marine Remote Sensing Group, University of the Aegean in Plastic Litter
96 Projects (2021 and 2022), The Ocean Cleanup).

97

98 **2.1.2 Weathered plastics**

99 To mimic the effect of solar radiation on plastic items in the environment, pristine plastics without
100 additives were exposed to UV radiation in an Atlas SunTest CPS+ weathering chamber, simulating one
101 solar year in central Europe (Gewert et al., 2018) (Table 2). Prior to the treatment, plastic specimens
102 were cut, using a hot knife, into 2 x 10 cm sheets in order to fit them into closed Quartz cuvettes. Different
103 treatments for weathering conditions were tested: i.e., dry UV exposure, seawater UV exposure (35 psu)
104 and dark controls (dry and wet), and each treatment consisted of a set of three independent replicates
105 for each polymer type (Table 4).

106

107 **Table 2.** Parameters of the UV chamber, used in artificial weathering experiments

UV irradiation W / m ²	Black Stand Temperature (BST) °C	Chamber Temperature (CHT) °C	Test durations h
60	50	30 - 35	917

108

109 **2.1.3 Biofouled plastics**

110 To test the effects of biofilm attached to the surface of plastic items on their spectral reflectance, we
111 induced biofilm growth on the two surfaces of pristine samples. Since surface roughness can affect
112 biofilm growth and its survival (Rodriguez, D., B. Einarsson, 2012), one of the two surfaces of the
113 commercial plastic specimens, was manually treated for approximately 10 seconds with sandpaper
114 (grain size 80) to create a rougher surface compared to the smooth and untouched one. An aquarium
115 was filled with unfiltered seawater collected from the port of Ostend (Belgium), which was renewed every
116 two weeks, kept at 20 °C, and aerated with an air pump. Plastic specimens were suspended in the
117 aquarium with paper clippers and rope (Fig. 1) to allow the biofilm to form on all of the surfaces.



118
119 **Figure 1.** Aquarium set-up for biofilm growth on pristine plastic specimens. Picture taken by Giulia Leone
120

121 **2.1.4 Field plastic items**

122 Plastic specimens were collected from the Port of Antwerp and in the Scheldt River, near Temse Bridge
123 (Belgium) (Fig.2). The plastic coming from the river Scheldt were collected by a plastic collector installed
124 by the Belgian company Dredging, Environmental and Marine Engineering NV (DEME) Environmental
125 Contractors (DEC) on behalf of Vlaamse Waterweg (the Flemish authority responsible for waterways in
126 Flanders, Belgium).



127

128 **Figure 2.** Example of field litter items collected in 2020 from the river Scheldt, near Temse Bridge (Belgium) by the plastic
129 collector installed by the Belgian company DEME Environmental Contractors (DEC) on behalf of Vlaamse Waterweg (the
130 Flemish authority responsible for waterways in Flanders, Belgium). Picture taken by Giulia Leone

131

132 2.2 Spectral measurements of plastic items

133 The spectral reflectance measurements of plastic specimens were performed using an Analytical
134 Spectral Device (ASD) Field-Spec4 (Table 3). The reflectance was automatically derived and normalized
135 to a 99% Labsphere Spectralon® Lambertian panel. In the silo tank, the Spectralon panel was positioned
136 on the holder of the adjustable arm, at the same distance from the ASD field of view as the dry plastic
137 specimen.

138

139 **Table 3.** Hyperspectral-radiometer specifications used in the two locations of the study, calibration facility and water silo tank
140 ASD: Analytical Spectral Device; VNIR: visible-near infrared; SWIR: shortwave infrared

141



	ASD specifications in the calibration facility	ASD specifications in the tank
Spectral range (nm)	350 - 2500	350 - 2500
Spectral resolution (nm)	VNIR: ca. 3 nm SWIR: 10 - 12 nm	VNIR: ca. 3 nm SWIR: 10 - 12 nm
Scans per measurements	30	10
Replicate measurements	5	5
Foreoptic field of view	8°	1°

142

143 3 Experimental set-ups

144 Following the procedure described by Knaeps et al., 2021, we measured the spectral reflectance of the
145 different plastic samples at two different facilities: 1) Measurements done in dry conditions were
146 performed at the optical calibration laboratory of the Flemish Institute for Technological Research (VITO),
147 Belgium; 2) the series of wet and submerged measurements were acquired in a mesocosm setting, with
148 experiments being performed in a conical shaped silo tank at the Flanders Hydraulics Research facility
149 in Antwerp, Belgium.

150

151 3.1 Laboratory set-up

152 All the plastic specimens were measured in the optical calibration facility (VITO, Belgium). This laboratory
153 consisted of a dark room where a desk equipped with two halogen tungsten lamps and a holder for the
154 ASD field of view, allowed us to collect the measurements.

155



156 **3.1 Silo tank set-up**

157 The water silo tank (2 top diameter x 3 depth m), at the Flanders Hydraulics Research (Belgium), was
158 equipped with a controlled mixer with a double pitch blade impeller that permitted the mixing of the water
159 to obtain suspensions of sediment or algae. No information about the rpm was available on the metal
160 impeller at the water tank. We attached a tailor-made aluminium frame to the water tank for the mounting
161 of spectroradiometer detector and light, together with a plexiglass sample holder to lower the plastic
162 specimens in the tank and measure their spectral reflectance at different water depths. The plexiglass
163 was cut to fit the plastic samples which were held by black plastic paper clips. In the current study, we
164 acquired data using a single lamp attached to the frame, with an angle of 40 degrees. To reduce
165 undesired stray light, a dark environment was created with black plastic cover around the experimental
166 set-up and in addition black plastic bags were held against the tank by means of wooden rings with
167 weights and waterproof tape. In the tank, samples attached to the plexiglass sample holder were first
168 measured at dry conditions, just above the water level and then carefully lowered in the water at fixed
169 depths. The water level above the sample was measured with a ruler and the chosen depths were: 1
170 cm, 2 cm, 4 cm and 8 cm. After the submersion, the plastic was measured again above the water level
171 as wetted sample. The same steps were performed in clear and turbid water.

172

173 **3.1.1 Water with added sediment**

174 Natural sediment was collected using a manual Van Veen grab by the Flanders Marine Institute (VLIZ)
175 during a sampling campaign as part of the PLUXIN project, in Nieuwpoort (Belgium). The sediment was
176 transported to the laboratory (VLIZ), placed into a metal container, and dried in the oven for 4 days at 60
177 °C, and was afterwards manually crushed using a mortar, and stored in the dark until further use. To add
178 the crushed sediment into the tank, before any measurements, the tank metal impeller was activated for
179 a one and a half minute to allow the sediment to stay in suspension in the water. This action was repeated
180 approximately every 30 minutes to maintain the sediment in suspension. The actual concentration of
181 sediment added in the tank was measured by Flanders Hydraulics. The suspended sediment
182 concentration (SSC) was determined gravimetrically, after filtration of the sample through a filter with a
183 pore size of 0.45 µm and drying at a temperature of 105 °C. The results showed a low concentration of
184 sediment of 4 mg / L and a higher concentration of 16 mg / L.



185

186 **3.1.2 Water with added microalgae**

187 To evaluate how the readings of spectral reflectance of plastic specimens can be affected by turbidity
188 in the water due to microalgae suspension, measurements were performed using two concentrations of
189 freshwater *Pseudokirchneriella subcapitata*. The concentrations were selected to mimic the natural
190 conditions of spring and late summer seasonal concentration of green algae for a Western European
191 River (Ibelings et al., 1998). The nominal concentrations of microalgae were 1500 cells / mL and 3000
192 cell / mL. The microalgae *P. subcapitata* was cultured at the Research unit “Health” at VITO, and the
193 stock solution of $8,18 \times 10^6$ algae / mL was kept for 11 days in a cold room and in dark before being used
194 in the experimental set-up. After pouring 450 mL of stock solution, before any measurements, the metal
195 impeller was activated for a one and a half minute to allow the algae to suspend into the water. This
196 action was repeated approximately every 30 minutes to maintain the algae in suspension. To obtain a
197 higher concentration of algae that would mimic a nominal concentration of 3000 cells / mL, other 450 mL
198 of stock solution were then added into the tank.

199

200 **4. Data description**

201

202 In this study we created a dataset with the spectral reflectance of ten plastic polymers undergone to two
203 treatments (i.e., artificial weathering and artificial biofouling) in addition to field collected samples and
204 pristine plastics (Table 4). All data included in the presented dataset were curated before submission
205 and consist of the raw reflectance data of the samples. A metadata section, included in the dataset,
206 explains the data, adding additional and essential information. For instance, information on the polymer
207 type, μ FTIR results with the corresponding library matching score, origin of the sample, date and time of
208 collection. The sample code was created following the naming convention of Knaeps et al., 2021. To
209 ensure that end users can correctly interpret each section of the metadata, a README file is available
210 together with the dataset.

211 We suggest that users of this dataset perform splice correction on the data using Python (Fig. 3) or any
212 suitable software. This processing step will lead to the obtention of spectra that do not present
213 radiometric steps at the joints of the detectors. In addition, pseudo-replicates of each plastic sample were



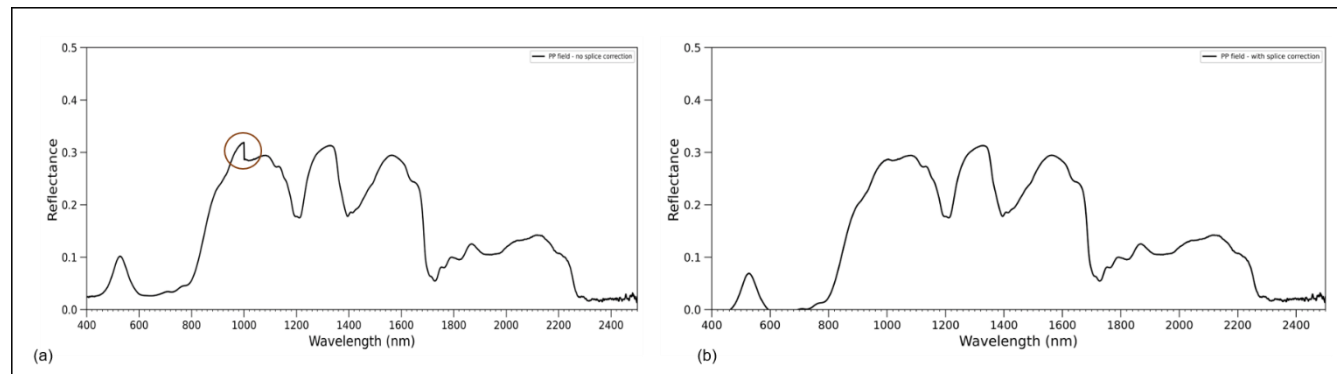
214 taken, and therefore we advise that users calculate the mean of these measurements to obtain a single
 215 spectrum for each observed item (Fig. 4).

216

217 **Table 4.** Overview of the polymer and treatment performed during the study (SSC: suspended sediment concentration)

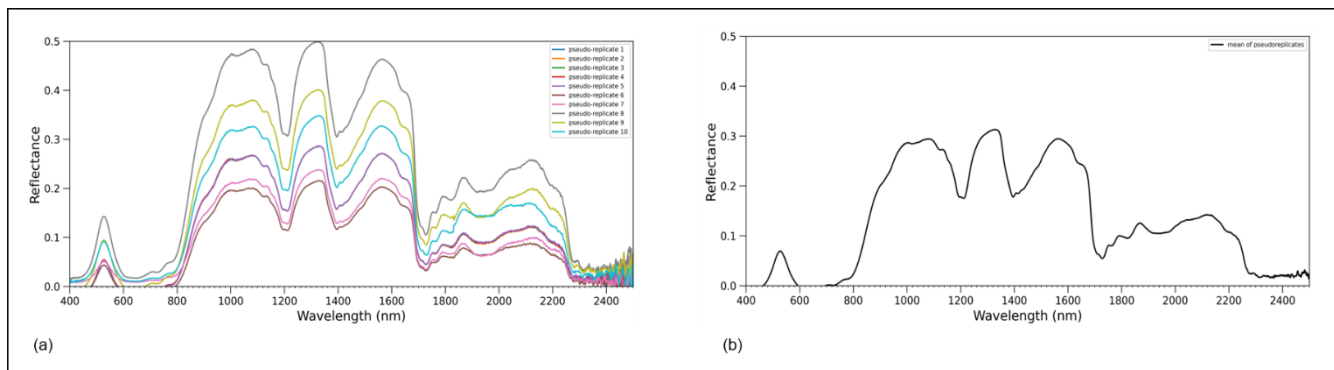
Treatment	Polymer tested	Origin/Supplier	State of samples	Condition of the water in the tank
<ul style="list-style-type: none"> • Pristine • Artificially dry weathered • Artificially seawater weathered • Artificially biofouled on pristine surface • Artificially biofouled on rough surface • Field 	<ul style="list-style-type: none"> • PS • PE • PP • PET amorphous • PET crystalline • PVC • Extruded polystyrene • Thermoplastic elastomer • Fluorocarbon • Paraffin 	<ul style="list-style-type: none"> • Carat • The Ocean Cleanup • Shop • Port of Antwerp • Scheldt • VITO • Marine Remote Sensing Group 	<ul style="list-style-type: none"> • Dry • Wet • Submerged 	<ul style="list-style-type: none"> • Clear water, no turbidity • Algae: <ol style="list-style-type: none"> 1. Nominal conc. 3000 cells/ml 2. Nominal conc. 1500 cells/ml • Sediment: <ol style="list-style-type: none"> 1. SSC 4mg/L 2. SSC 16mg/L

218



219

220 **Figure 3.** Spectrum of polypropylene (PP) collected in the field without (a) and with splice correction (b). The circle indicates
 221 the jump around 1000nm due to the connection between the different ASD cameras.
 222
 223



224
225 **Figure 4.** Spectra of field sample. Spectrum (a) shows the pseudo-replicates of the low homogeneous field samples. Spectrum
226 (b) is the mean of all the pseudo-replicates.

227 228 **5. Data availability**

229 The data are available in the open access repository Marine Data Archive at <https://doi.org/10.14284/530>
230 (Leone et al., 2021)

231 232 **6. Conclusions**

233 The use of remote sensing technologies can be used in the detection, observation and monitoring of
234 marine plastic pollution. However, due to a lack of knowledge of the optical features of environmental
235 plastics, small steps can be made in designing algorithms to appropriately detecting plastic pollution.
236 The presented hyperspectral dataset is a step forward in the knowledge of the optical features of plastic
237 litter when exposed to natural agents such as UV radiations or the growth of biofilm. We anticipate that
238 this dataset will contribute to the definition of optical spectral bands and assist in the development of
239 algorithms for the observation, monitoring and discrimination of plastics in a (semi-) operational
240 environment.

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249

250 Credit author statement

251 **Giulia Leone:** Conceptualization, Methodology, Writing – original draft, Writing – review & editing,
252 Visualization, Funding acquisition, Data curation, Investigation. **Ana I. Catarino:** Conceptualization,
253 Methodology, Writing – review & editing, Supervision. **Liesbeth De Keukelaere:** Conceptualization,
254 Writing – review & editing. **Mattias Bossaer:** Investigation, Methodology, Writing – review & editing. **Els**
255 **Knaeps:** Conceptualization, Methodology, Writing – review & editing, Supervision, Funding acquisition.
256 **Gert Everaert:** Conceptualization, Methodology, Funding acquisition, Supervision, Writing – review &
257 editing

258

259 Competing interests

260 The authors declare that they have no conflict of interest.

261

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266 Aegean (Greece) and The Ocean Cleanup (The Netherlands) for sharing plastic specimens with us.

267

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