

# Hyperspectral reflectance dataset of pristine, weathered and biofouled plastics

Giulia Leone<sup>1,2,4,5, \*</sup>, Ana I. Catarino<sup>1</sup>, Liesbeth De Keukelaere<sup>3</sup>, Mattias Bossaer<sup>1</sup>, Els Knaeps<sup>3§</sup> Gert Everaert<sup>1§</sup>

<sup>1</sup>Flanders Marine Institute (VLIZ), Oostende, Belgium

<sup>2</sup>Ghent University, Research Group Aquatic Ecology, Ghent, Belgium

<sup>3</sup>Flemish Institute for Technological Research (VITO), Belgium

<sup>4</sup> Research Institute for Nature and Forest, Aquatic Management (INBO), Brussels, Belgium

<sup>5</sup> Research Foundation – Flanders (FWO), Brussels, Belgium

§ Shared senior co-authorship

\*Correspondence to: Giulia Leone (giulia.leone@ugent.be)

## Abstract

This work presents a hyperspectral reflectance dataset of macroplastic samples acquired using Analytical Spectral Devices (ASD) FieldSpec 4. Samples analyzed consisted of pristine, artificially weathered, and biofouled plastic items and plastic debris samples collected in the docks of the Port of Antwerp and in the river Scheldt near Temse Bridge (Belgium). The hyperspectral signal of each sample was measured in controlled dry conditions in an optical calibration facility at the Flemish Institute for Technological Research, and, for a subset of plastics, under wet and submerged conditions in a silo tank at Flanders Hydraulics. The wet and submerged hyperspectral signals were measured in a mesocosm setting that mimicked environmentally relevant concentrations of freshwater microalgae and suspended sediment. The ASD was equipped with an 8° field of view at the calibration facility, and a 1° field of view was used in the mesocosm setting. The dataset obtained complies with the Findability, Accessibility, Interoperability, and Reuse (FAIR) principles and is available in the open-access repository Marine Data Archive (<https://doi.org/10.14284/530>, Leone et al., 2021).

## 27 **1 Introduction**

28 The spectral reflectance measurements, collected in the framework of the Plastic Flux for Innovation and  
29 Business Opportunities in Flanders (PLUXIN) project, contribute to the current knowledge on the  
30 detection of plastics using remote sensing techniques.

31 In recent years, focus has been placed on the detection of plastic litter using remote sensing techniques  
32 such as optical sensors on satellites, aircraft, and drones (Garaba and Dierssen, 2018; Martínez-  
33 Vicente et al., 2019). With the increasing demand for these technologies, it is crucial to generate  
34 knowledge on the diagnostic spectral properties of not only pristine but also weathered and biofouled  
35 plastics (Moshtaghi et al., 2021) that are representative of the variety of environmental plastics.

36 Currently, the spectral reflectance of dry plastics is known and is already applied in the field of material  
37 recycling (Moroni et al., 2015), but it is restricted to items assessed for dry measurements. To be able to  
38 identify plastic litter in aquatic environments such as rivers, harbors, and oceans, we require the  
39 acquisition of the spectral features when plastics are either wet or submerged (Moshtaghi et al., 2021),  
40 as water absorbs the light in both the near (NIR) and shortwave infrared (SWIR). In addition, other water  
41 constituents such as sediment or algae could further impact the reflected signal of plastic items  
42 (Moshtaghi et al., 2021).

43 To date, only a limited number of high-quality datasets consisting of hyperspectral measurements of wet  
44 and submerged plastic litter, have been published in open-access repositories (e.g., Garaba and  
45 Dierssen, 2018, Garaba and Dierssen, 2020; Knaeps et al., 2021). As plastics in our environment are so  
46 diverse in polymer type, color, transparency, thickness, state (pristine, biofouled, weathered, wrinkled)  
47 and wetness (dry, wet, submerged), it is critical to generate, within the scientific community,  
48 substantiated data sets which represent plastics in many different facets. The collected spectra can  
49 serve as references or endmember spectra in future Remote Sensing plastic detection techniques and  
50 help to understand the complexity of plastic detection through spectral analysis. It is recognized that not  
51 all possible scenarios can be measured in an experimental way, therefore the dataset can further be  
52 used to compare with and complement numerical simulations. The dataset described in the current paper  
53 aims at complementing the existing datasets by adding new information about the hyperspectral  
54 reflectance of pristine plastic items, harvested plastic litter, and artificially weathered and biofouled plastic

55 samples. In addition, the dataset reports the optical features of plastics acquired in various water turbidity  
56 conditions obtained by adding sediment or algae to the water, at selected concentrations.

57 The dry spectral reflectance of plastic specimens, consisting of different polymers, was measured using  
58 an Analytical Spectral Device (ASD) FieldSpec 4. For a selection of these samples, the spectral  
59 reflectance was also collected in wet and submerged settings.

60 The presented data can be used to generate insights into the spectral properties of plastic litter and how  
61 these features change when plastics are exposed to natural agents or different depths. The dataset was  
62 compiled following the FAIR (Findable, Accessible, Interoperable, Reusable; go-fair.org) principles and  
63 it is publicly available at <https://doi.org/10.14284/530> (Leone et al., 2021).

64

## 65 **2. Data collection**

66 Data collection consisted of measuring the spectral reflectance of different plastic specimens using the  
67 ASD. The measurements were conducted in two different settings: at an optical calibration facility located  
68 at the Flemish Institute for Technological Research (Mol, Belgium), and at a silo tank at the Flanders  
69 Hydraulics Research facility (Antwerp, Belgium). The spectral reflectance of each plastic specimen was  
70 collected at least 5 times (“pseudo-replicates”) within 1 minute. These pseudo-replicates are  
71 measurements taken by slightly changing the position of the plastic sample. Therefore, very  
72 homogeneous plastics will have less variation in the pseudo-replicates’ reflectance.

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### 74 **2.1 Plastic specimens**

75 We aimed to collect the spectral reflectance of plastic items as representative as possible of commonly  
76 found plastics in the environment (LI et al., 2016), and to provide additional information on their  
77 reflectance when exposed to natural agents such as sunlight (UV radiation) or biofilm growth. To do so,  
78 we have selected and analyzed a total of 10 polymer types. The selected polymers were Polyethylene  
79 (PE), Polypropylene (PP), Polystyrene (PS), and two types of Polyethylene terephthalate (PET),  
80 crystalline and amorphous, Fluorocarbon, Thermoplastic elastomer, Polyvinyl chloride (PVC), Nylon 6  
81 (PA6) and Paraffin (Table 1; Table S1). In addition, we have further artificially weathered and biofouled  
82 a selection of 6 plastic polymer types. The polymer discrimination was based on the available information  
83 from the supplier or marked on the plastic itself (e.g., plastic bottles or bags with an identifiable polymer  
84 tag). In addition, for a set of samples, we confirmed the polymer type using a micro-Fourier Transform

85 Infrared Spectroscopy ( $\mu$ FTIR, PerkinElmer, FTIR spectrometer Frontier). The percentage of matching  
 86 scores with a library was recorded and it is reported in the dataset. Table 1 provides an overview of  
 87 measured plastic types within this study and compared them to existing datasets. This dataset adds  
 88 additional information on plastics spectral reflectance of similar polymers to what is already available in the  
 89 literature, allowing comparison, but also novel conditions and treatments (Table 1).

90

91 **Table 1.** Comparison of polymers and conditions analyzed in this dataset and in the literature. (D = Dry; W = Wet;  
 92 S = Submerged; PE = polyethylene, PP = polypropylene, PS = polystyrene, PET = Polyethylene terephthalate,  
 93 PVC = Polyvinyl chloride, PA6, PA 6.6. = Polyamide 6 and 66, LD-PE = low density polyethylene, FEB = fluorinated  
 94 ethylene propylene Teflon, ABS = terpolymer lustran 752, PMMA = polymethyl methacrylate)

Polymer type	This dataset			Knaeps et al., 2022			Garaba and Dierssen, 2018			Garaba and Dierssen, 2020		
	D	W	S	D	W	S	D	W	S	D	W	S
<b><i>Pristine Sample</i></b>												
PE	X	X	X									
PP	X	X	X	X	X	X	X				X	
PS	X	X	X				X				X	
PET amorphous	X	X	X									
PET crystalline	X	X	X									
PET unspecified	X			X			X				X	
PVC	X						X				X	
Thermoplastic elastomer	X											
PA6 and PA 6.6	X						X				X	
Paraffin	X											
LD-PE	X			X			X				X	
Polyester				X								
FEB							X				X	
ABS							X				X	
Merlon							X				X	
PMMA							X				X	
Non specified polymers				X	X	X						
<b><i>Environmentally weathered and biofouled samples</i></b>												
PE	X	X	X									
PP	X	X	X									
PS	X											
PET unspecified	X	X	X									
Fluorocarbon	X											
Paraffin	X											
Non specified polymers				X			X	X			X	X

<b>Artificially biofouled samples</b>												
<b>PE</b>	X	X	X									
<b>PP</b>	X	X	X									
<b>PS</b>	X	X	X									
<b>PET amorphous</b>	X	X	X									
<b>PET crystalline</b>	X	X	X									
<b>PVC</b>	X											
<b>Artificially weathered samples</b>												
<b>PE</b>	X	X	X									
<b>PP</b>	X	X	X									
<b>PS</b>	X	X	X									
<b>PET amorphous</b>	X	X	X									
<b>PET crystalline</b>	X	X	X									
<b>PVC</b>	X											

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## 2.2.1 Pristine plastic specimens

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To enhance the physio-chemical properties of an end plastic product, plastic manufacturers often incorporate additive substances into the pristine polymer (Garaba et al., 2021). Additives included in plastics, such as colored plastics, may be influencing the spectra (Garaba et al., 2021). Thus, to avoid confounding measurements, sheets of 6 x 6 cm were obtained from a commercial supplier (Carat, Germany, <https://www.carat-lab.com>) as pure pristine plastic polymers without any additional additive for the following polymers: PP, PE, PS, PET amorphous, PET crystalline. In addition, no color or stains were added to the commercial plastics, and as such, these selected samples were transparent or white. The other pristine plastic samples were either purchased from a local shop (Oostende, Belgium) or obtained from other institutes (Marine Remote Sensing Group, University of the Aegean in Plastic Litter Projects (2021 and 2022), The Ocean Cleanup).

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## 2.1.2 Weathered plastics

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To mimic the effect of solar radiation on plastic items in the environment, pristine plastics without additives were exposed to UV radiation in an Atlas SunTest CPS+ weathering chamber, simulating one solar year in central Europe (Gewert et al., 2018) (Table 2). Prior to the treatment, plastic specimens were cut, using a hot knife, into 2 x 10 cm sheets in order to fit them into closed Quartz cuvettes. Although UV exposure in a laboratory cannot perfectly mimic environmental conditions, to reproduce them as accurately as possible we have tested different treatments for weathering conditions: i.e., dry UV

115 exposure, seawater UV exposure (35 PSU), and dark controls (dry and wet), and each treatment consisted  
116 of a set of three independent replicates for each polymer type (Table 4).  
117

118 **Table 2.** Parameters of the UV chamber, used in artificial weathering experiments

UV irradiation W / m <sup>2</sup>	Black Stand Temperature (BST) °C	Chamber Temperature (CHT) °C	Test durations h
60	50	30 - 35	917

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### 120 **2.1.3 Biofouled plastics**

121 To test the effects of biofilm attached to the surface of plastic items on their spectral reflectance, we  
122 induced biofilm growth on the two surfaces of pristine samples. Since surface roughness can affect  
123 biofilm growth and its survival (Rodriguez, D., B. Einarsson, 2012), one of the two surfaces of the  
124 commercial plastic specimens, was manually treated for approximately 10 seconds with sandpaper  
125 (grain size 80) to create a rougher surface compared to the smooth and untouched one. An aquarium  
126 was filled with unfiltered seawater collected from the port of Ostend (Belgium), which was renewed every  
127 two weeks, kept at 20 °C, and aerated with an air pump. Plastic specimens were suspended in the  
128 aquarium with paper clippers and rope (Fig. 1) to allow the biofilm to form on all of the surfaces.



129

130 **Figure 1.** Aquarium set-up for biofilm growth on pristine plastic specimens. The picture was taken by Giulia Leone

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### 132 **2.1.4 Field plastic items**

133 Plastic specimens were collected from the Port of Antwerp and in the Scheldt River, near Temse Bridge  
134 (Belgium) and are naturally exposed to weathering and biofouling and are, compared to pristine polymers,

135 non-homogeneous. The plastic coming from the river Scheldt was collected by a plastic collector  
136 installed by the Belgian company Dredging, Environmental and Marine Engineering NV (DEME)  
137 Environmental Contractors (DEC) on behalf of Vlaamse Waterweg (the Flemish authority responsible for  
138 waterways in Flanders, Belgium).

139

## 140 **2.2 Spectral measurements of plastic items**

141 The spectral reflectance measurements of plastic specimens were performed using an Analytical  
142 Spectral Device (ASD) Field-Spec4 (Table 3). The reflectance was automatically derived and normalized  
143 to a 99% Labsphere Spectralon® Lambertian panel. In the silo tank, the Spectralon panel was positioned  
144 on the holder of the adjustable arm at the same distance from the ASD field of view as the dry plastic  
145 specimen.

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147 **Table 3.** Hyperspectral-radiometer specifications used in the two locations of the study, calibration facility and water silo tank  
148 ASD: Analytical Spectral Device; VNIR: visible-near infrared; SWIR: shortwave infrared

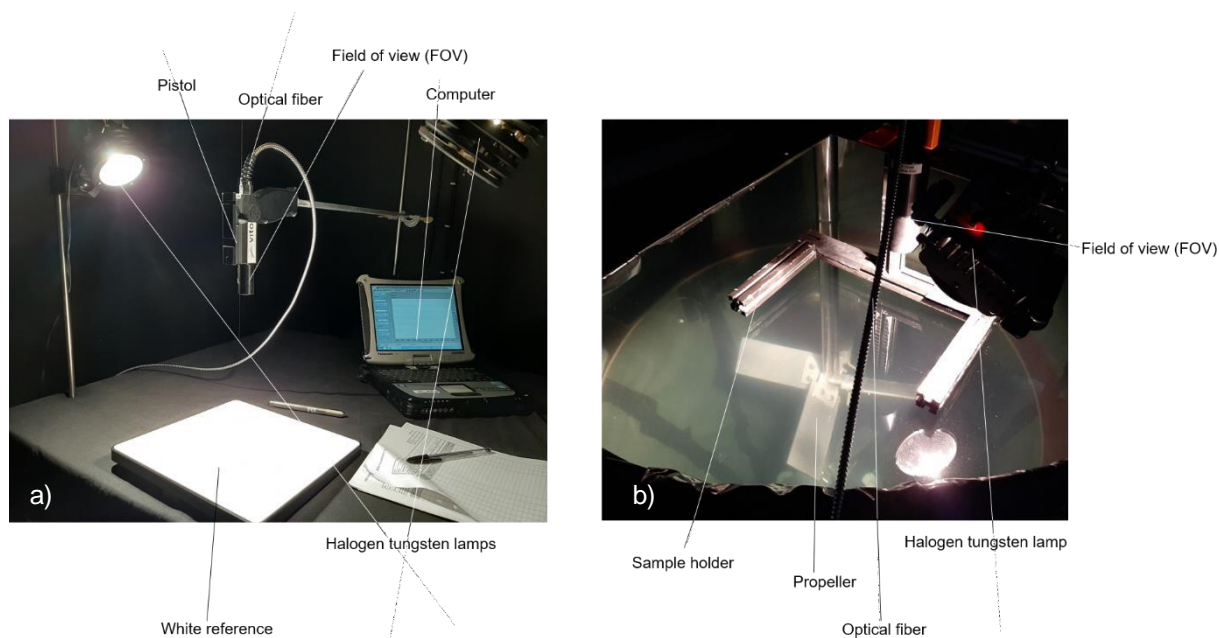
149

	ASD specifications in the calibration facility	ASD specifications in the tank
Spectral range (nm)	350 - 2500	350 - 2500
Spectral resolution (nm)	VNIR: ca. 3 nm SWIR: 10 - 12 nm	VNIR: ca. 3 nm SWIR: 10 - 12 nm
Scans per measurements	30	10
Replicate measurements	5	5
Foreoptic field of view	8°	1°

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151 **3 Experimental set-ups**

152 Following the procedure described by Knaeps et al., 2021, we measured the spectral reflectance of the  
153 different plastic samples at two different facilities: 1) Measurements done in dry conditions were  
154 performed at the optical calibration laboratory of the Flemish Institute for Technological Research (VITO),  
155 Belgium; 2) the series of wet and submerged measurements were acquired in a mesocosm setting, with  
156 experiments being performed in a conical shaped silo tank at the Flanders Hydraulics Research facility  
157 in Antwerp, Belgium. In both laboratory and silo tank set-ups (Fig. 2), we have attached a laser pen to  
158 the ASD pistol grip to ensure that, at all times, the fiber optics of the ASD were pointing at the plastic  
159 samples.



160  
161 **Figure 2.** Experimental set-up. a) laboratory set-up; b) silo tank set-up. Pictures were taken by Giulia Leone

162 **3.1 Laboratory set-up**

163 All the plastic specimens were measured in the optical calibration facility (VITO, Belgium). This laboratory  
164 consisted of a dark room where a desk equipped with two halogen tungsten lamps and a holder for the  
165 ASD field of view, allowed us to collect the measurements.



## 166 **3.1 Silo tank set-up**

167 The water silo tank (Diameter 2m, Depth 3m), at the Flanders Hydraulics Research (Belgium), was  
168 equipped with a controlled mixer with a double pitch blade impeller that permitted the mixing of the water  
169 to obtain suspensions of sediment or algae. No information about the rpm was available on the metal  
170 impeller at the water tank. We attached a tailor-made aluminum frame to the water tank for the mounting  
171 of the spectroradiometer detector and light, together with a plexiglass sample holder to lower the  
172 plastic specimens in the tank and measure their spectral reflectance at different water depths. The  
173 plexiglass was cut to fit the plastic samples which were held by black plastic paper clips. In the current  
174 study, we acquired data using a single lamp attached to the frame, with an angle of 40 degrees. To  
175 reduce undesired stray light, a dark environment was created with a black plastic cover around the  
176 experimental set-up and in addition, black plastic bags were held against the tank by means of  
177 wooden rings with weights and waterproof tape. In the tank, samples attached to the plexiglass sample  
178 holder were first measured at dry conditions, just above the water level and then carefully lowered in  
179 the water at fixed depths. The water level above the sample was measured with a ruler and the chosen  
180 depths were: 1 cm, 2 cm, 4 cm, and 8 cm. After the submersion, the plastic was measured again above  
181 the water level as a wetted sample. The same steps were performed in clear and turbid water.

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### 183 **3.1.1 Water with added sediment**

184 Natural sediment was collected using a manual Van Veen grab by the Flanders Marine Institute (VLIZ)  
185 during a sampling campaign as part of the PLUXIN project, in Nieuwpoort (Belgium). The sediment was  
186 transported to the laboratory (VLIZ), placed into a metal container, and dried in the oven for 4 days at 60  
187 °C, and was afterward manually crushed using a mortar, and stored in the dark until further use. To add  
188 the crushed sediment into the tank, before any measurements, the tank metal impeller was activated for  
189 one and a half minutes to allow the sediment to stay in suspension in the water. This action was repeated  
190 approximately every 30 minutes to maintain the sediment in suspension. The actual concentration of  
191 sediment added to the tank was measured by Flanders Hydraulics. The suspended sediment  
192 concentration (SSC) was determined gravimetrically, after filtration of the sample through a filter with a  
193 pore size of 0.45 µm and drying at a temperature of 105 °C. The results showed a low concentration of  
194 sediment of 4 mg / L and a higher concentration of 16 mg / L.

195

### 196 **3.1.2 Water with added microalgae**

197 To evaluate how the readings of spectral reflectance of plastic specimens can be affected by turbidity  
198 in the water due to microalgae suspension, measurements were performed using two concentrations of  
199 freshwater *Pseudokirchneriella subcapitata*. The concentrations were selected to mimic the natural  
200 conditions of spring and late summer seasonal concentrations of green algae for a Western European  
201 River (Ibelings et al., 1998). The nominal concentrations of microalgae were 1500 cells / mL and 3000  
202 cell / mL. The microalgae *P. subcapitata* was cultured at the Research unit “Health” at VITO, and the  
203 stock solution of  $8,18 \times 10^6$  algae / mL was kept for 11 days in a cold room and dark before being used  
204 in the experimental set-up. After pouring 450 mL of stock solution, before any measurements, the metal  
205 impeller was activated for one and a half minutes to allow the algae to suspend in the water. This  
206 action was repeated approximately every 30 minutes to maintain the algae in suspension. To obtain a  
207 higher concentration of algae that would mimic a nominal concentration of 3000 cells / mL, other 450 mL  
208 of stock solution were then added into the tank.

209

## 210 **4. Data description**

211

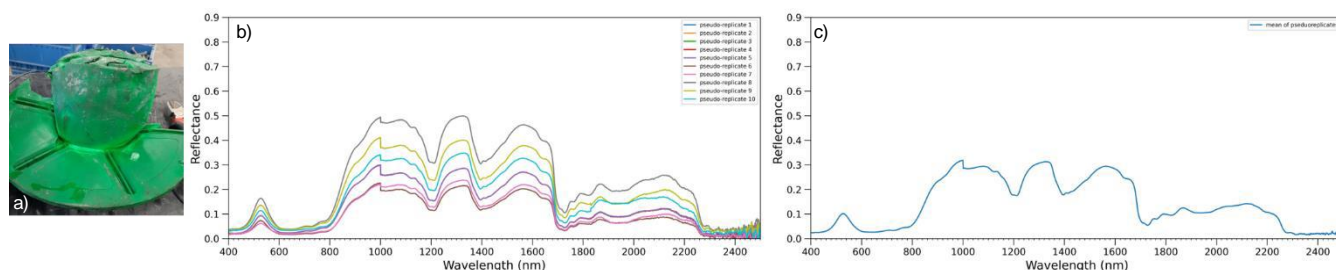
212 In this study we created a dataset with the spectral reflectance of ten plastic polymers undergone two  
213 treatments (i.e., artificial weathering and artificial biofouling) in addition to field collected samples and  
214 pristine plastics (Table 4). All data included in the presented dataset were curated before submission  
215 and consisted of the raw reflectance data of the samples. A metadata section, included in the dataset,  
216 explains the data, adding additional, and essential information. For instance, information on the polymer  
217 type,  $\mu$ FTIR results with the corresponding library matching score, the origin of the sample, date and time  
218 of collection. The sample code was created following the naming convention of Knaeps et al., 2021. To  
219 ensure that end users can correctly interpret each section of the metadata, a README file is available  
220 together with the dataset. From the presented dataset it is possible to visually show the spectral  
221 reflectance of different plastic polymers and compare the different conditions experimentally tested (Fig.  
222 1S; Fig. 2S). For instance, it is possible to derive the effect of weathering and biofouling on the spectral  
223 reflectance of a polymer when compared with the same pristine one.

224 We suggest that users of this dataset perform splice correction on the data using Python or any suitable  
 225 software. This processing step will lead to the obtention of spectra that do not present radiometric steps  
 226 at the joints of the detectors. In addition, pseudo-replicates of each plastic sample were taken, and  
 227 therefore, we advise that users calculate the mean of these measurements to obtain a single spectrum  
 228 for each observed item (Fig 3).

229 **Table 4.** Overview of the polymer and treatment performed during the study (SSC: suspended sediment concentration)

Treatment	Polymer tested	Origin/Supplier	State of samples	Condition of the water in the tank
<ul style="list-style-type: none"> <li>• Pristine</li> <li>• Artificially dry weathered</li> <li>• Artificially seawater weathered</li> <li>• Artificially biofouled on pristine surface</li> <li>• Artificially biofouled on rough surface</li> <li>• Field</li> </ul>	<ul style="list-style-type: none"> <li>• PS</li> <li>• PE</li> <li>• PP</li> <li>• PET amorphous</li> <li>• PET crystalline</li> <li>• PVC</li> <li>• Extruded polystyrene</li> <li>• Thermoplastic elastomer</li> <li>• Fluorocarbon</li> <li>• Paraffin</li> </ul>	<ul style="list-style-type: none"> <li>• Carat</li> <li>• The Ocean Cleanup</li> <li>• Shop</li> <li>• Port of Antwerp</li> <li>• Scheldt</li> <li>• VITO</li> <li>• Marine Remote Sensing Group</li> </ul>	<ul style="list-style-type: none"> <li>• Dry</li> <li>• Wet</li> <li>• Submerged</li> </ul>	<ul style="list-style-type: none"> <li>• Clear water, no turbidity</li> <li>• Algae:               <ol style="list-style-type: none"> <li>1. Nominal conc. 3000 cells/ml</li> <li>2. Nominal conc. 1500 cells/ml</li> </ol> </li> <li>• Sediment:               <ol style="list-style-type: none"> <li>1. SSC 4mg/L</li> <li>2. SSC 16mg/L</li> </ol> </li> </ul>

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**Figure 3.** Spectra of field sample and figure of the object from which the spectra were obtained (a). Spectrum (b) shows the pseudo-replicates of the low homogeneous field samples. Spectrum (c) is the mean of all the pseudo-replicates. No splice correction was applied. The picture was taken by Giulia Leone.

235

## 236 5. Data availability

237 The data are available in the open access repository Marine Data Archive at <https://doi.org/10.14284/530>  
 238 (Leone et al., 2021)

239

## 240 6. Conclusions

241 The use of remote sensing technologies can be used in the detection, observation, and monitoring of  
242 marine plastic pollution. However, due to a lack of knowledge of the optical features of environmental  
243 plastics, small steps can be made in designing algorithms to appropriately detect plastic pollution.  
244 The presented hyperspectral dataset is a step forward in the knowledge of the optical features of plastic  
245 litter when exposed to natural agents such as UV radiations or the growth of biofilm. In addition, from the  
246 data presented it is possible to investigate the effects that biofouling and weathering have on different  
247 polymers. Lastly, the conditions in which a plastic polymer is (i.e., dry, wet, or submerged with different  
248 turbidity) are also described and assessed in the presented dataset. Therefore, we anticipate that this  
249 dataset will contribute to the definition of optical spectral bands and assist in the development of  
250 algorithms for the observation, monitoring, and discrimination of plastics in a (semi-) operational  
251 environment.

252

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259

## 260 **Credit author statement**

261 **Giulia Leone:** Conceptualization, Methodology, Writing – original draft, Writing – review & editing,  
262 Visualization, Funding acquisition, Data curation, Investigation. **Ana I. Catarino:** Conceptualization,  
263 Methodology, Writing – review & editing, Supervision. **Liesbeth De Keukelaere:** Conceptualization,  
264 Writing – review & editing. **Mattias Bossaer:** Investigation, Methodology, Writing – review & editing. **Els**  
265 **Knaeps:** Conceptualization, Methodology, Writing – review & editing, Supervision, Funding acquisition.  
266 **Gert Everaert:** Conceptualization, Methodology, Funding acquisition, Supervision, Writing – review &  
267 editing

268

## 269 **Competing interests**

270 The authors declare that they have no conflict of interest.

271

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277

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