

1 **The biogeography of relative abundance of soil fungi and bacteria in top surface soil**

2 Kailiang Yu<sup>1\*</sup>, Johan van den Hoogen<sup>1</sup>, Zhiqiang Wang<sup>2</sup>, Colin Averill<sup>1</sup>, Devin Routh<sup>1</sup>, Gabriel  
3 R. Smith<sup>3,1</sup>, Rebecca E. Drenovsky<sup>4</sup>, Kate M. Scow<sup>5</sup>, Fei Mo<sup>6</sup>, Mark P Waldrop<sup>7</sup>, Yuanhe  
4 Yang<sup>8,9</sup>, Weize Tang<sup>9,10</sup>, Franciska T. De Vries<sup>11</sup>, Richard D. Bardgett<sup>12</sup>, Peter Manning<sup>13</sup>, Felipe  
5 Bastida<sup>14</sup>, Sara G. Baer<sup>15</sup>, Elizabeth M. Bach<sup>16</sup>, Carlos García<sup>14</sup>, Qingkui Wang<sup>17</sup>, Linna Ma<sup>8</sup>,  
6 Baodong Chen<sup>18,9</sup>, Xianjing He<sup>19</sup>, Sven Teurlincx<sup>20</sup>, Amber Heijboer<sup>21,22</sup>, James A. Bradley<sup>23,24</sup>,  
7 Thomas W Crowther<sup>1\*</sup>

8 <sup>1</sup>Institute of Integrative Biology, ETH Zürich, Zürich, Switzerland

9 <sup>2</sup>Institute for Advanced Study, Chengdu University, Chengdu, China

10 <sup>3</sup>Department of Biology, Stanford University, California, USA

11 <sup>4</sup>Biology Department, John Carroll University, Ohio, USA

12 <sup>5</sup>Department of Land, Air and Water Resources, University of California, Davis, California,  
13 USA

14 <sup>6</sup>College of Agronomy, Northwest A&F University, Shaanxi, PR China

15 <sup>7</sup>U.S. Geological Survey. Geology, Minerals, Energy, and Geophysics Science Center. Menlo  
16 Park, California, USA

17 <sup>8</sup>State Key Laboratory of Vegetation and Environmental Change, Institute of Botany, Chinese  
18 Academy of Sciences, Beijing, China

19 <sup>9</sup>University of Chinese Academy of Sciences, Beijing, China

20 <sup>10</sup>Key Laboratory of Vegetation Restoration and Management of Degraded Ecosystems, South  
21 China Botanical Garden, Chinese Academy of Sciences, Guangzhou, China

22 <sup>11</sup>Institute of Biodiversity and Ecosystem Dynamics, University of Amsterdam, Amsterdam, the  
23 Netherlands

24 <sup>12</sup>Department of Earth and Environmental Sciences, University of Manchester, Oxford Road,  
25 Manchester, UK

26 <sup>13</sup>Senckenberg Biodiversity and Climate Research Centre, Frankfurt, Germany

27 <sup>14</sup>CEBAS-CSIC. Department of Soil and Water Conservation. Campus Universitario de  
28 Espinardo, Murcia, Spain.

29 <sup>15</sup>Kansas Biological Survey and Department of Ecology & Evolutionary Biology, University of  
30 Kansas, Lawrence, Kansas, USA

31 <sup>16</sup>The Nature Conservancy, Nachusa Grasslands, Franklin Grove, IL, USA

32 <sup>17</sup>Huitong Experimental Station of Forest Ecology, CAS Key Laboratory of Forest Ecology and  
33 Management, Institute of Applied Ecology, Shenyang, PR China

34 <sup>18</sup>State Key Laboratory of Urban and Regional Ecology, Research Center for Eco-Environmental  
35 Sciences, Chinese Academy of Sciences, Beijing, China

36 <sup>19</sup>Key Laboratory of the Three Gorges Reservoir Region's Eco-Environment, Ministry of  
37 Education, Chongqing University, Chongqing, China

38 <sup>20</sup>Department of Aquatic Ecology, Netherlands Institute of Ecology (NIOO-KNAW),  
39 Wageningen, Netherlands

40 <sup>21</sup>Biometris, Wageningen University & Research, Wageningen, Netherlands

41 <sup>22</sup>Ecology and Biodiversity Group, Department of Biology, Institute of Environmental Biology,  
42 Utrecht University, Padualaan, Netherlands

43 <sup>23</sup>School of Geography, Queen Mary University of London, London, E1 4NS, UK

44 <sup>24</sup>Interface Geochemistry, GFZ German Research Centre for Geosciences, Potsdam, Germany

45 Corresponding author: [kai86liang@gmail.com](mailto:kai86liang@gmail.com) or [tom.crowther@usys.ethz.ch](mailto:tom.crowther@usys.ethz.ch)

46

47 **Abstract.** Fungi and bacteria are the two dominant groups of soil microbial communities  
48 worldwide. By controlling the turnover of soil organic matter, these organisms directly regulate  
49 the ~~cycling~~exchange of carbon between the soil and the atmosphere. Fundamental differences in  
50 the physiology and life history of bacteria and fungi suggest that variation in the biogeography of  
51 soil fungal and bacterial relative abundance could drive striking differences in carbon  
52 decomposition and soil organic matter formation across different biomes. However, a lack of  
53 global and predictive information on the distribution of these organisms in terrestrial ecosystems  
54 has prevented the inclusion of soil fungal and bacterial relative abundance and the associated  
55 processes into global biogeochemical models. Here, we used a global scale dataset (~~>3000 distinct~~  
56 ~~observations of soil fungal and bacterial abundance~~) in the top soil surface (~~up to 15 cm~~)~~<3000~~  
57 ~~distinct observations of soil fungal and bacterial abundance~~) to generate the first quantitative and  
58 spatially high resolution ( $1\text{km}^2$ ) explicit map of soil fungal proportion, defined as fungi/fungi +  
59 bacteria, across terrestrial ecosystems. We reveal striking latitudinal trends where fungal  
60 dominance increases in cold and high latitude environments with large soil carbon stocks. There  
61 was strong non-linear response of fungal dominance to environmental gradient, i.e., mean annual  
62 temperature (MAT) and net primary productivity (NPP). Fungi and bacteria dominated in regions  
63 with low and high MAT and NPP, respectively, thus representing slow vs. fast soil energy channels,  
64 a concept with a long history in soil ecology. These high-resolution models provide the first steps  
65 towards representing the major soil microbial groups and their functional differences in global  
66 biogeochemical models to improve predictions of soil organic matter turnover under current and  
67 future climate scenarios.

68 **Keywords:**

设置了格式: 上标

69 **1. Introduction**

70 Fungi and bacteria are the dominant members of soil microbial communities worldwide in terms  
71 of diversity, abundance and biomass (Bahram et al., 2018). Representing distinct kingdoms of life,  
72 bacteria and fungi systematically differ in a multitude of physiological and life-history traits with  
73 direct implications for global soil biogeochemical cycles (Waring et al., 2013; Wieder et al., 2015)  
74 including the decomposition of organic matter, which contributes to one of the largest fluxes of  
75 CO<sub>2</sub> on Earth (Glassman et al., 2018). Compared to bacteria, fungi generally have slower growth  
76 and turnover rates (Rousk and Bååth, 2007), greater carbon (C) to nutrient stoichiometry (Waring  
77 et al., 2013), greater capacity to degrade a wider and more recalcitrant range of substrates  
78 (Strickland and Rousk, 2010) and potentially higher C use efficiency (Soares and Rousk, 2019).  
79 For these reasons, a new generation of soil and ecosystem models have begun to explicitly  
80 represent these fundamentally distinct fast and slow cycling microbial groups, suggesting that  
81 spatially-explicit information about the relative abundance of fungal and bacteria in a region can  
82 dramatically improve the accuracy of global carbon cycling model predictions (Shi et al., 2018;  
83 Sulman et al., 2014; Wieder et al., 2013, 2015). Generating an understanding of the factors  
84 affecting the biogeography of the relative abundance of fungal and bacteria in soil, and its  
85 connection to global carbon cycle, would represent a breakthrough step forward in our general  
86 understanding of the natural history of soil microbial life.

87       Temperature, precipitation, soil pH and soil C:N have all been linked to the balance of fungi  
88 vs. bacteria within soil communities across different spatial scales (Bahram et al., 2018;  
89 Strickland and Rousk, 2010; Tedersoo et al., 2014). Relative to fungi, bacteria tend to dominate  
90 in locations with high soil nutrient contents or in frequently disturbed soils that limit the growth  
91 of fungal hyphae or make N more available (Fierer et al., 2009; Van Der Heijden et al., 2008;

92 Strickland and Rousk, 2010). However, until now, the relative importance of these different  
93 environmental drivers remains relatively unclear at global scale, and the biogeography of these  
94 major functional groups (fungi vs. bacteria) has only been demonstrated at local and regional  
95 scales. A recent analysis suggested that the relative soil bacterial abundance is high in tropical  
96 latitudes and decreases in abundance towards the high-latitude boreal regions, where fungi tend  
97 to dominate (Bahram et al., 2018). Translating these broad-scale trends into quantitative,  
98 spatially explicit information will be necessary if we intend to represent regional variations in  
99 soil community functioning (Wieder et al., 2013, 2015), or predict future changes in terrestrial  
100 carbon and nutrient cycling.

101       Some progress was made in the quantitative and spatially explicit understanding of global  
102 biogeographic patterns of fungal and bacterial biomass and their biomass ratio. By synthesizing  
103 phospholipid-derived fatty acids data from 1323 locations across the globe, and extrapolating  
104 linear relationships with environmental factors, a recent study generate the global maps of fungal  
105 and bacterial biomass and their biomass ratio at the resolution of 0.5 degree for top soil (0-30  
106 cm) (He et al., 2020). This approach provided the support for the broad-scale latitudinal trends,  
107 with high fungal dominance in high-latitude regions. Yet to date, there are three crucial  
108 knowledge gaps to be addressed. First, we still lack a high resolution evaluation of the spatial  
109 patterns and regional contingencies in fungal:bacterial ratios, which would allow representation  
110 of microbial-mediated mechanisms that operate within and/or across ecosystems at fine scales  
111 (Frindte et al., 2019; Zhu et al., 2017). Second, the response of soil microbial community  
112 composition across environmental gradients are expected to be non-linear, with strong interactive  
113 effects of different environmental characteristics that give rise to thresholds that diverge from the  
114 global latitudinal trends (Sengupta et al., 2021; Wang et al., 2018; Waring et al., 2013). This

115 non-linear linkage of soil microbial communities with environmental resource gradient has not  
116 been assessed, while it has fundamental implications on ecosystem functions and management  
117 solutions (Sengupta et al., 2021; Wang et al., 2018). Third, there are distinct difference of soil  
118 nutrients, soil microbial community and the associated biogeochemical processes across soil  
119 depths, i.e., from top surface soil (i.e., 0-10 cm) to top subsurface soil (i.e., 0-30 cm) (Lavahun et  
120 al., 1996; Yue et al., 2015). A continental-scale empirical study further showed that strong  
121 positive associations among soil microbial community, fertility and plant productivity are limited  
122 to the top surface soil (Delgado-Baquerizo et al., 2017), thus highlighting its potential dominant  
123 role regulated by top surface soil microbial communities on ecosystem functions and the  
124 research needs of generating a global spatially explicit understanding of soil fungi and bacteria in  
125 top surface soil.

126 Here, we present a global analysis of total and relative abundance of soil fungi and bacteria  
127 in soil surfaces (defined as top 10-15 cm) informed from over 3000 spatially distinct surface soil  
128 observations from phospholipid-derived fatty acids (PLFA) (Fig. 1a). The use of PLFA data  
129 provides an opportunity to provide quantitative insights into the abundances of ~~the~~these major  
130 functional groups. We conducted the analysis on the abundances in view of the uncertainty in  
131 conversion factors used to convert abundance derived from PLFA to biomass (Frostegård et al.,  
132 2011; Klamer and Bååth, 2004). We used machine learning to link the variation in soil fungi and  
133 bacteria abundances to global variation in 95 climate, vegetation, and soil variables. This allowed  
134 us to 1) explore the environmental drivers of fungal and bacterial dominance, defined as fungal  
135 proportion - fungi/(fungi + bacteria), where values closer to 1 indicate a higher fungal dominance  
136 and values closer to zero indicate a greater bacterial dominance (see Methods); 2) examine the  
137 non-linear response or pattern of fungal proportion across environmental gradients, i.e., mean

138 annual temperature-MAT and net primary productivity-NPP. Based on the observed  
139 relationships (by accounting for the non-linearity), we generated a quantitative spatially-explicit  
140 global map (1 km<sup>2</sup>) of fungal proportion, and assessed how soil fungal and bacterial dominance  
141 varies with key climate, soil, vegetation and geographic drivers.

设置了格式: 上标

## 142 2. Material and methods

### 143 2.1. Data acquisition of soil microbe composition

144 We compiled data of abundance of soil fungi and bacteria and fungal proportion, defined as  
145 fungi/(fungi + bacteria). We focused on phospholipid-derived fatty acids (PLFA) and the data  
146 derived from PLFA reported the balance between fungal and bacterial PLFAs (Frostegård et al.,  
147 2011) can provide a valuable estimation of the comparative dominance of both functional groups.  
148 The data based on qPCR was not included because of difference in units with PLFA. With non-  
149 significant difference using data of fungal proportion and fungi : bacteria ratio, we focused on and  
150 reported the results on the fungal proportion rather than fungi : bacteria ratio because 1) The fungal  
151 proportion is insensitive to whether fungi or bacteria are the numerator (i.e. bacterial proportion =  
152 1 – fungal proportion), and 2) fungal proportion had more spread frequency distribution and thus  
153 led to better machine learning predictions (Fig. S1). The data was compiled by a primary literature  
154 review through Google Scholar, Web of Science (<http://apps.webofknowledge.com>) and China  
155 National Knowledge Infrastructure Database (<http://cnki.net>) till 30 June, 2020 using the keywords  
156 “fungi”, “bacteria”, “abundance”, “PLFA”. To be included in our data analysis, the study had to  
157 at least have the following metadata: longitude and latitude, sampling date, sampling depth,  
158 information on land use (agriculture, tree plantations, or natural sites), units and the methods used.  
159 In total, this led to 319 references. We further used the following criteria to select eligible  
160 references and datasets: (1) when the studies were manipulative experiments, we only included the

161 data from “control” plots (Chen et al., 2016). (2) we standardized our efforts by focusing on all  
162 samples that were collected from the top surface soils ( $\approx$  0-10/15 cm) because this layer contains  
163 greater biomass and has the majority of sample size. (3) we used the datasets based on reporting  
164 abundance with units of nmol,  $\mu$ mol, or mol% since the majority (>90%) of datasets reported  
165 abundance. Thus, we exclude all datasets reporting biomass instead of abundance. (4) we excluded  
166 observations located in sea since our study focuses on terrestrial ecosystems. (5) we only included  
167 the datasets on soil samples derived from field experiments and thus excluded the datasets from  
168 incubation experiments. (6) some datasets reported in original references as average across  
169 sampling sites or sampling dates were included.

170 The criteria were carefully scrutinized by three independent researchers and this ultimately  
171 led to 179 eligible references (see Supplementary references for PLFA) used for this study. In total,  
172 we compiled a dataset of fungal proportion ( $n = 3224$ ) at a global scale. The subset of data ( $n =$   
173 1795) with only natural ecosystems (Fig. S2a) were used to examine the potential role of land use  
174 change (see Supplementary Methods). The results showed minimal difference of the two scenarios  
175 of including all data and natural ecosystems. All data points falling within the same 30 arc-seconds  
176 ( $\sim 1\text{-km}^2$ ) pixel were aggregated via an average. The aggregated data of fungal proportion ( $n = 946$   
177 for all data;  $n = 716$  for natural ecosystems) were used to examine its environmental controls and  
178 geospatial modelling in making the global map (Fig. 1a; Fig. S2a).

179 The spatial variations of fungi and bacteria ratio or fungal proportion across latitude could be  
180 influenced by either changes (increases or decreases) in abundance of fungi or bacteria or both.  
181 Thus, to better understand the biogeographic pattern of fungal and bacterial composition, we also  
182 analysed the spatial patterns of abundance of fungi and bacteria by using the abundance data with  
183 the same unit ( $\text{nmol g}^{-1}$  PLFA). In total, our data compiling led to a final subset of 2753, and 2759



184 samples which were used for further analyses of abundance of fungi and bacteria, respectively (Fig.  
185 S3). As compared to the larger sample size of fungal proportion (n = 946 for all data), the data of  
186 abundance of fungi (n = 646 for all data) and bacteria (n = 647 for all data) aggregated within the  
187 30 arc-seconds (~1-km<sup>2</sup>) pixel via an average were used for the analysis of their spatial trends  
188 across vegetation biome, vegetation type and latitude (see Supplementary Methods).

## 189 **2.2. Geospatial modelling**

190 A stack (n = 95) of ecologically relevant, global map layers including soil physical, chemical and  
191 nutrient properties, climate conditions, vegetative indices, radiation and topographic variables and  
192 anthropogenic covariates (Supplementary Table 1) were used to derive the environmental factors  
193 which could affect fungal proportion. All of these covariate map layers were standardized at 30  
194 arc-seconds resolution ( $\approx 1\text{km}^2$  at the equator) (van den Hoogen et al., 2019). These covariates  
195 were then derived based on the georeferenced coordinates of the soil samples aggregated at 30 arc-  
196 seconds resolution.

197 We used the Random Forest machine learning algorithm (see Supplementary Methods) with  
198 the derived 95 covariates to extrapolate these relationships between fungal proportion and  
199 environmental conditions across the globe and generate the first spatially-explicit, quantitative  
200 map of fungal proportion at a global scale. The strength of prediction was evaluated using k-fold  
201 cross validation (with k = 10) and the best model having high coefficient of determination and low  
202 standard deviation in the mean cross-validation were used to generate the global map of fungal  
203 proportion. The standard error sharply decreased with increasing sample size across all vegetation  
204 biomes and the analysis showed that an efficient prediction required a large sample size (n > 500)  
205 (Fig. S4). To evaluate the sensitivity, we also generate the uncertainty (standard deviation as a  
206 fraction of the mean predicted value) map of fungal proportion by using a stratified bootstrapping

设置了格式: 上标

207 procedure (van den Hoogen et al., 2019). The stratification category was the sampled biomes of  
208 each point feature (fungal proportion) with the total number collection of fungal proportion points  
209 to avoid biases. In total, 100 bootstrap iterations were used, thus generating 100 global maps of  
210 fungal proportion used to quantify statistically robust 95% confidence intervals per pixel.

### 211 **2.3. Environmental drivers and statistic analysis**

212 To examine the environmental controls of soil microbial composition at a global scale, we chose  
213 the top drivers (Chen et al., 2016; Drenovsky et al., 2010a; de Vries et al., 2012) which include  
214 soil properties, climate conditions, vegetation index and human activities (see Supplementary  
215 Methods). These variables were examined to avoid multicollinearity using a matrix of pairwise  
216 correlations to remove any variable with high correlations ( $R > 0.7$ ) with other predictor variables  
217 (Anderegg et al., 2013). Random Forest machine learning algorithm was then used to determine  
218 variable importance for each variable (Breiman, 2001). Mean decrease in accuracy (%IncMSE)  
219 and mean decrease gini (IncNodePurity) were reported and the variables with greater values  
220 of %IncMSE and IncNodePurity are more important in influencing fungal proportion. Partial  
221 functions of most important variables (MAT and NPP) were plotted using forestFloor package to  
222 examine their influences on fungal proportion.

### 223 **3. Code and data availability of machine learning**

224 The code and data of machine learning is available at  
225 <https://github.com/KailiangYu/Biogeography-of-soil-microbes.git>.

## 226 **4. Results and discussion**

### 227 **4.1. Raw data patterns of fungal proportion**

228 Globally, we observed greater than 10-fold variation in soil fungal proportion across all sites,  
229 ranging from 0.01 to 0.6 (Fig. 1b). At a global scale, we found clear latitudinal trends, with the

230 abundance of both fungi and bacteria increasing in high-latitude regions. Yet, the abundance of  
231 fungi increased with latitude at a greater rate than the abundance of bacteria (Fig. S5), resulting in  
232 a higher proportion of fungi in the cold, high-latitude regions. These latitudinal trends lend support  
233 to the general global patterns detected in a previous broad-scale analysis (Bahram et al., 2018) and  
234 in a recent meta-data analysis (He et al., 2020). As such, the highest fungal dominance was  
235 observed in tundra and boreal forest ecosystems (mean  $\pm$  1SE:  $0.23 \pm 0.02$ ; Fig. 1b). In addition,  
236 high elevation and cold grasslands (i.e., Montane grasslands) with large soil organic C (SOC)  
237 content generally harbor higher proportion of fungi, relative to bacteria (Fig. 1b).

238         Within similar climates, soil fungal and bacterial abundance as well as fungal proportion  
239 was greatest in ecosystems harboring woody vegetation compared to grasslands and managed  
240 (agricultural) ecosystems (Fig. S6). This finding is consistent with the idea that ecosystems  
241 dominated by woody plants generate lignified, more recalcitrant and nutrient poor soil C inputs  
242 that characteristically favor fungal dominance (Fierer et al., 2009; Strickland and Rousk, 2010),  
243 and have a biomass stoichiometry better suited to low nutrient environments (Waring et al., 2013).  
244 But we stress that this link of belowground soil microbial composition (fungi vs bacteria) with  
245 aboveground plant community composition (woody plants vs grasses) can be complex, non-linear  
246 and even divergent, as demonstrated by the non-existence of woody plants in grasslands and  
247 scarcity of grasses in forests but with well mixed fungi vs bacteria abundances. This raises the  
248 curiosity whether the interactions, associations or couplings of belowground soil microbial  
249 composition vs aboveground plant community composition are stronger in ecosystems where  
250 woody plants and grasses interact or coexist (i.e., savannas) (Yu and D’Odorico, 2015). It also  
251 remains unclear how this coupling could improve our understanding of ecosystem carbon cycling  
252 and other services.

253 Management of agricultural ecosystems often disrupts soil fungal networks (i.e. tillage,  
254 frequent dry/wet cycles due to irrigation, machine operations, etc.), which decreases the abundance  
255 of fungi relative to bacteria in agricultural soils (Fig. S6) (Drenovsky et al., 2010b; Jangid et al.,  
256 2011; Waldrop et al., 2017). A central concern in agricultural ecosystems is the tradeoff of  
257 increased food production to feed the increasing population vs the decreased soil carbon storage  
258 to accelerate the global climate change (Sanderman et al., 2017). This study showed the higher  
259 bacterial abundance relative to fungal abundance in soils of agricultural lands where soil carbon  
260 storage is low; this corresponds with the global trends of bacterial dominance in low latitude where  
261 soil carbon storage is low. These results suggest the potential strong but complex interactions and  
262 feedbacks of soil microbial composition and soil functions (i.e., soil carbon storage) (Bardgett et  
263 al., 2008), while the mechanistic links need further studies.

#### 264 **4.2. Drivers of fungal proportion**

265 Globally, the fungal proportion in soil can be predicted by few primary environmental drivers (Fig.  
266 2; Fig. S7). Specifically, mean annual temperature (MAT) and primary productivity (NPP) were  
267 strong determinants of fungal dominance. The responses of fungal proportion to both MAT and  
268 NPP were strongly non-linear, with warmer, more productive regions of the world (i.e. tropical  
269 forest biomes) showing lower dominance of fungi as compared to colder, less productive  
270 ecosystems (i.e. boreal forest and tundra biomes, Fig. 3; Fig. S8). This pattern is consistent with  
271 the idea that fungi and bacteria represent slow vs. fast soil energy channels, respectively (Crowther  
272 et al., 2019; Malik et al., 2016), a concept with a long history in soil ecology (Moore et al., 2003;  
273 Moore and William Hunt, 1988). This finding is important because it could potentially link the  
274 belowground slow – fungi vs fast – bacteria energy channels with aboveground plant slow growth  
275 rates – woody plants vs fast growth rates – grasses, while the linkage could be complex, non-linear

276 or even divergent. The fast vs slow concept or spectrum have fundamentally improved the  
277 understandings and predictions of land carbon storage across resource gradient or under global  
278 change. The faster growth could be typically trade off with higher mortality or heterotrophic  
279 respiration with resource (i.e., CO<sub>2</sub>) enriched conditions (Jiang et al., 2020; Terrer et al., 2021; Yu  
280 et al., 2019), thus constraining land carbon storage. This raises the question of how the  
281 belowground fast vs slow energy channels and the aboveground fast vs slow growth spectrum  
282 could be potentially linked or integrated to assess land carbon storage.

283         Temperature can affect soil microbial composition in complex ways, via directly  
284 physiology or via indirectly soil substrate (Romero-Olivares et al., 2017). Previous studies have  
285 shown the non-linear response of soil fungal and bacterial ratio to soil substrate (Waring et al.,  
286 2013). The non-linear trends of the temperature sensitivity (Q<sub>10</sub>) of soil organic C decomposition,  
287 as regulated by soil fungal and bacterial ratio, were also found along latitude (Wang et al., 2018).  
288 Other environmental variables such as soil C to nitrogen ratio (C:N) have previously been found  
289 to be important drivers in influencing fungal proportion within local and regional scale analyses  
290 (Fierer et al., 2009; Waring et al., 2013). Our results suggest a more complicated relationship  
291 between fungal proportion and the soil C:N. In the low range of soil C:N values, fungal proportion  
292 decreased with soil C:N (Fig. S9a), suggesting the likely role of site-specific differences (i.e.,  
293 climate or plant community) in out-weighting the influence of N availability (Soares and Rousk,  
294 2019). Aside from these ecosystems, we observed a positive relationship between fungal  
295 proportion and soil C:N at a global scale, consistent with previous work at local and regional scales  
296 (Strickland and Rousk, 2010; Waring et al., 2013). Additionally, pH has been thought as a critical  
297 driver of microbial diversity and biomass in soils. At local scales, previous studies reported either  
298 no relationship, a negative correlation or convex curve between fungal and bacterial ratio and soil

设置了格式: 下标

299 pH (Rousk et al., 2009, 2010; de Vries et al., 2012). Our global scale analysis suggests a convex  
300 relationship between fungal proportion and soil pH, with fungi dominating only within a narrow  
301 pH range (<5-6) (Fig. S9b).

### 302 **4.3. Biogeographic pattern from the machine learning model**

303 Across all samples, the machine learning model was able to predict the variation in fungal and  
304 bacterial dominance with high predictive accuracy ( $R^2 = 0.43/0.35$  in 10-fold cross validation;  $R^2$   
305  $= 0.92/0.91$  in final model; Fig. S10a-b). By extrapolating these relationships across terrestrial  
306 ecosystems, we could identify clear global trends in fungal dominance. Despite these general  
307 global scale patterns of increase in fungi dominance with latitude, our models also revealed  
308 regional contingencies that diverge from the global trends (Fig. 4a; Fig. S11a). For instance,  
309 Northeastern Europe is dominated by woody vegetation and exhibits high fungal proportion, while  
310 the United Kingdom and northern Kazakhstan have much lower fungal proportion despite being  
311 at comparable latitude, likely because these areas are dominated by herbaceous vegetation with  
312 lower lignin content than in woody tissues. Tibetan alpine grasslands are at comparatively much  
313 lower latitude but have high values of fungal proportion in part due to very high SOC stocks and  
314 cold temperatures. Model predictions of fungal proportion had high uncertainty in dry regions  
315 (i.e., Northern and Southern Africa, Australia, Western USA, eastern Mongolia) (Fig. 4b; Fig.  
316 S11b), presumably because of the low sample size in drylands and/or complex response of fungi  
317 and bacteria to water availability (Fierer et al., 2009; Strickland and Rousk, 2010). Indeed,  
318 our datasets are mostly concentrated to US, Europe and East Asia, thus highlighting the data gaps  
319 at tropical and boreal biomes. Even for the temperate biome, there were data gaps in west Australia  
320 and central Asia. Because of the unbalanced sample distribution, we also used a bootstrapping  
321 strategy (100 iterations) by randomly sampling 90% data with replacement. The results showed

322 the similar spatial patterns of fungal proportion (Fig. S12a) and uncertainty (Fig. S12b) as the  
323 scenario of using full dataset without bootstrapping.

324 Our study differs from a previous study (He et al., 2020) in several aspects including  
325 sample size ( $n > 3000$ ), spatial resolution ( $1\text{km}^2$ ), consideration of non-linearity (through random  
326 forest analysis), soil depth (soil surface 0-10/15 cm). We also note that our analysis sticks to the  
327 original data of abundance derived from PLFA instead of converting abundance to biomass.  
328 Conversion of abundance to biomass needs the conversion factor, which has large uncertainty  
329 (Frostegeård et al., 2011; Klamer and Bååth, 2004). Our high resolution map would allow  
330 representation of microbial-mediated mechanisms at fine scales to link with ecosystem functions.  
331 For instance, the significant functional differences between fungi and bacterial mean that the  
332 relative dominance of fungi vs. bacteria is likely to influence a wide range of ecosystem  
333 functions such as C use efficiency (CUE) of the decomposer community (Six et al., 2006; Soares  
334 and Rousk, 2019) and enzymatic activity in soil N vs P acquisition (Caldwell, 2005; Crowther et  
335 al., 2019). At fine, local or even regional scales, these relationships between soil microbial  
336 composition and ecosystem functions could only be well identified using fine scale maps of soil  
337 microbial composition.

#### 338 **4.4. Implications and limitations of this study**

339 It is generally accepted that the soil microbiome exerts major control over soil processes, and in  
340 turn ecosystem functioning, and by extension the global biogeochemical cycles (Bahram et al.,  
341 2018; Crowther et al., 2019; Van Der Heijden et al., 2008; Jenny, 1941). Fungi and bacteria  
342 represent most of the diversity of life on Earth (Bardgett and van der Putten, 2014; Locey and  
343 Lennon, 2016). Yet, inclusion of fungal and bacterial abundance into quantitative ecosystem and  
344 Earth system models has been hindered by the paucity of information about organisms at

设置了格式: 上标

345 appropriate spatial scales. Here, we impose a global top-down constraint on the broad composition  
346 of soil microbial life. By doing so, we hope to empower microbial, ecosystem and Earth-system  
347 scientists to consider how this broad constraint on the soil biodiversity may inform and transform  
348 how we understand terrestrial ecosystem functioning. As we develop a spatially-explicit  
349 understanding of the global soil community, we will be able to better parameterize and benchmark  
350 our predictions about the rate and efficiency of carbon turnover in soil and the feedbacks to  
351 ongoing climate change.

352         Despite of the progress made in this study, here we clarify two limitations on this study.  
353 First, our study highlights the data gaps in fungal proportion prediction in low latitude – tropical  
354 biome and high latitude – boreal biome (i.e., boreal forests and tundra). Tropical vs boreal  
355 biomes are hotspots or debated regions with their relative capacity and capability to sequester  
356 atmospheric CO<sub>2</sub> and mitigate climate change in an increasingly changing climate (Schimel et  
357 al., 2015; Tagesson et al., 2020). They are also regions with striking differences of soil  
358 microbial composition (fungal proportion), plant communities and soil carbon storage, thus  
359 suggesting their potentially strong interactions and feedbacks in these regions (Bardgett et al.,  
360 2008). Boreal biome contains large amount of soil organic carbon which could be sensitive to  
361 global change (i.e., warming), whereby soil microbial community (i.e., total biomass or the  
362 relative abundance of of soil fungi and bacteria) could play an essential role. Second, microbial  
363 biomass (C) is more relevant to be linked with soil carbon cycling and carbon stock in term of  
364 their own contribution by living carbon pools and the impacts of its microbial necromass (Liang  
365 et al., 2019), while the conversion factor of converting abundance into biomass across space is  
366 currently not available. To mechanistically and explicitly incorporate soil microbial composition  
367 into biogeochemical models, the biogeographic patterns of abundance or biomass of each major



368 group (fungi vs bacteria), the relative ratio within fungi (i.e., saprotrophic fungi, arbuscular  
369 mycorrhiza fungi vs ectomycorrhizal fungi) and/or bacteria (i.e., gram positive bacteria vs gram  
370 negative bacteria) would also be critical in view of their striking functional difference (Averill et  
371 al., 2014; Crowther et al., 2019). These knowledge gaps highlight the urgent research needs in  
372 these new research endeavors with the increasing availability of datasets.

### 373 5. Conclusions

374 This study used a global scale dataset (>3000 distinct observations of soil fungal and bacterial  
375 abundance) in the top soil surface (up to 15 cm) (~~>3000 distinct observations of soil fungal and~~  
376 ~~bacterial abundance~~) to generate the first quantitative and spatially high resolution (1 km<sup>2</sup>) explicit  
377 maps of soil fungal and bacterial relative abundance across global terrestrial ecosystems. Our  
378 machine learning approach (random forest) enabled us to link the variation in fungal proportion to  
379 global variation in climate, soil, vegetation and other environmental drivers, whilst accounting for  
380 the interactions and non-linearities among them. We found the striking latitudinal trends where  
381 fungal dominance increases in cold and high latitude environments with large soil carbon stocks.  
382 The fungal proportion in soil can be predicted by few primary environmental drivers – temperature  
383 and NPP with strong non-linear effects of temperature and NPP. We demonstrated that fungi and  
384 bacteria represent slow vs fast energy channels, whereby they dominate in regions of low MAT  
385 and NPP vs high MAT and NPP, respectively. Overall, our spatially-explicit model would enable  
386 us to explicitly represent the different contributions of fast - bacterial vs. slow – fungal energy  
387 channels in spatially-explicit biogeochemical models, with the potential to enhance the accuracy  
388 of soil carbon turnover and carbon storage predictions. We further highlight the data gaps in tropical  
389 and boreal regions and needs of future research endeavors in generating high resolution

设置了格式: 上标

390 biogeographic patterns of biomass of each major microbial group, the relative biomass ratios  
391 across and within major microbial groups.

392

393

394

395

396

397 **References**

- 398 Anderegg, L. D. L., Anderegg, W. R. L., Abatzoglou, J., Hausladen, A. M. and Berry, J. A.:  
 399 Drought characteristics' role in widespread aspen forest mortality across Colorado, USA,  
 400 *Glob. Chang. Biol.*, 19(5), 1526–1537, doi:10.1111/gcb.12146, 2013.
- 401 Averill, C., Turner, B. L. and Finzi, A. C.: Mycorrhiza-mediated competition between plants and  
 402 decomposers drives soil carbon storage, *Nature*, 505(7484), doi:10.1038/nature12901, 2014.
- 403 Bahram, M., Hildebrand, F., Forslund, S. K., Anderson, J. L., Soudzilovskaia, N. A., Bodegom,  
 404 P. M., Bengtsson-Palme, J., Anslan, S., Coelho, L. P., Harend, H., Huerta-Cepas, J.,  
 405 Medema, M. H., Maltz, M. R., Mundra, S., Olsson, P. A., Pent, M., Pölme, S., Sunagawa, S.,  
 406 Ryberg, M., Tedersoo, L. and Bork, P.: Structure and function of the global topsoil  
 407 microbiome, *Nature*, 560(7717), 233–237, doi:10.1038/s41586-018-0386-6, 2018.
- 408 Bardgett, R. D. and van der Putten, W. H.: Belowground biodiversity and ecosystem  
 409 functioning., *Nature*, 515(7528), 505–11, doi:10.1038/nature13855, 2014.
- 410 Bardgett, R. D., Freeman, C. and Ostle, N. J.: Microbial contributions to climate change through  
 411 carbon cycle feedbacks, *ISME J.*, 2(8), doi:10.1038/ismej.2008.58, 2008.
- 412 Breiman, L.: Random forests, *Mach. Learn.*, 45(1), 5–32, doi:10.1023/A:1010933404324, 2001.
- 413 Caldwell, B. A.: Enzyme activities as a component of soil biodiversity: A review, in  
 414 *Pedobiologia*, vol. 49., 2005.
- 415 Chen, Y. L., Ding, J. Z., Peng, Y. F., Li, F., Yang, G. B., Liu, L., Qin, S. Q., Fang, K. and Yang,  
 416 Y. H.: Patterns and drivers of soil microbial communities in Tibetan alpine and global  
 417 terrestrial ecosystems, *J. Biogeogr.*, 43(10), 2027–2039, doi:10.1111/jbi.12806, 2016.
- 418 Crowther, T. W., van den Hoogen, J., Wan, J., Mayes, M. A., Keiser, A. D., Mo, L., Averill, C.  
 419 and Maynard, D. S.: The global soil community and its influence on biogeochemistry,  
 420 *Science* (80-. ), doi:10.1126/science.aav0550, 2019.
- 421 Delgado-Baquerizo, M., Powell, J. R., Hamonts, K., Reith, F., Mele, P., Brown, M. V., Dennis,  
 422 P. G., Ferrari, B. C., Fitzgerald, A., Young, A., Singh, B. K. and Bissett, A.: Circular  
 423 linkages between soil biodiversity, fertility and plant productivity are limited to topsoil at the  
 424 continental scale, *New Phytol.*, 215(3), doi:10.1111/nph.14634, 2017.
- 425 Drenovsky, R. E., Steenwerth, K. L., Jackson, L. E. and Scow, K. M.: Land use and climatic  
 426 factors structure regional patterns in soil microbial communities, *Glob. Ecol. Biogeogr.*,  
 427 19(1), 27–39, doi:10.1111/j.1466-8238.2009.00486.x, 2010a.
- 428 Drenovsky, R. E., Steenwerth, K. L., Jackson, L. E. and Scow, K. M.: Land use and climatic  
 429 factors structure regional patterns in soil microbial communities, *Glob. Ecol. Biogeogr.*,  
 430 doi:10.1111/j.1466-8238.2009.00486.x, 2010b.
- 431 Fierer, N., Strickland, M. S., Liptzin, D., Bradford, M. A. and Cleveland, C. C.: Global patterns  
 432 in belowground communities, *Ecol. Lett.*, 12(11), 1238–1249, doi:10.1111/j.1461-  
 433 0248.2009.01360.x, 2009.
- 434 Frindte, K., Pape, R., Werner, K., Löffler, J. and Knief, C.: Temperature and soil moisture  
 435 control microbial community composition in an arctic–alpine ecosystem along elevational  
 436 and micro-topographic gradients, *ISME J.*, 13(8), doi:10.1038/s41396-019-0409-9, 2019.
- 437 Frostegård, Å., Tunlid, A. and Bååth, E.: Use and misuse of PLFA measurements in soils, *Soil*  
 438 *Biol. Biochem.*, doi:10.1016/j.soilbio.2010.11.021, 2011.
- 439 Glassman, S. I., Weihe, C., Li, J., Albright, M. B. N., Looby, C. I., Martiny, A. C., Treseder, K.  
 440 K., Allison, S. D. and Martiny, J. B. H.: Decomposition responses to climate depend on  
 441 microbial community composition, *Proc. Natl. Acad. Sci. U. S. A.*,

442 doi:10.1073/pnas.1811269115, 2018.

443 He, L., Mazza Rodrigues, J. L., Soudzilovskaia, N. A., Barceló, M., Olsson, P. A., Song, C.,  
444 Tedersoo, L., Yuan, F., Yuan, F., Lipson, D. A. and Xu, X.: Global biogeography of fungal  
445 and bacterial biomass carbon in topsoil, *Soil Biol. Biochem.*, 151,  
446 doi:10.1016/j.soilbio.2020.108024, 2020.

447 Van Der Heijden, M. G. A., Bardgett, R. D. and Van Straalen, N. M.: The unseen majority: Soil  
448 microbes as drivers of plant diversity and productivity in terrestrial ecosystems, *Ecol. Lett.*,  
449 doi:10.1111/j.1461-0248.2007.01139.x, 2008.

450 van den Hoogen, J., Geisen, S., Routh, D., Ferris, H., Traunspurger, W., Wardle, D. A., de  
451 Goede, R. G. M., Adams, B. J., Ahmad, W., Andriuzzi, W. S., Bardgett, R. D., Bonkowski,  
452 M., Campos-Herrera, R., Cares, J. E., Caruso, T., de Brito Caixeta, L., Chen, X., Costa, S.  
453 R., Creamer, R., Mauro da Cunha Castro, J., Dam, M., Djigal, D., Escuer, M., Griffiths, B.  
454 S., Gutiérrez, C., Hohberg, K., Kalinkina, D., Kardol, P., Kergunteuil, A., Korthals, G.,  
455 Krashevskaya, V., Kudrín, A. A., Li, Q., Liang, W., Magilton, M., Marais, M., Martín, J. A.  
456 R., Matveeva, E., Mayad, E. H., Mulder, C., Mullin, P., Neilson, R., Nguyen, T. A. D.,  
457 Nielsen, U. N., Okada, H., Rius, J. E. P., Pan, K., Peneva, V., Pellissier, L., Carlos Pereira da  
458 Silva, J., Pitteloud, C., Powers, T. O., Powers, K., Quist, C. W., Rasmann, S., Moreno, S. S.,  
459 Scheu, S., Setälä, H., Sushchuk, A., Tiunov, A. V., Trap, J., van der Putten, W., Vestergård,  
460 M., Villenave, C., Waeyenberge, L., Wall, D. H., Wilschut, R., Wright, D. G., Yang, J. and  
461 Crowther, T. W.: Soil nematode abundance and functional group composition at a global  
462 scale. *Nature*, doi:10.1038/s41586-019-1418-6, 2019.

463 Jangid, K., Williams, M. A., Franzluebbers, A. J., Schmidt, T. M., Coleman, D. C. and Whitman,  
464 W. B.: Land-use history has a stronger impact on soil microbial community composition  
465 than aboveground vegetation and soil properties, *Soil Biol. Biochem.*, 43(10), 2184–2193,  
466 doi:10.1016/j.soilbio.2011.06.022, 2011.

467 Jenny, H.: Factors of Soil Formation, *Soil Sci.*, doi:10.1097/00010694-194111000-00009, 1941.

468 Jiang, M., Medlyn, B. E., Drake, J. E., Duursma, R. A., Anderson, I. C., Barton, C. V. M., Boer,  
469 M. M., Carrillo, Y., Castañeda-Gómez, L., Collins, L., Crous, K. Y., De Kauwe, M. G., dos  
470 Santos, B. M., Emmerson, K. M., Facey, S. L., Gherlenda, A. N., Gimeno, T. E., Hasegawa,  
471 S., Johnson, S. N., Kännaste, A., Macdonald, C. A., Mahmud, K., Moore, B. D., Nazaries,  
472 L., Neilson, E. H. J., Nielsen, U. N., Niinemets, Ü., Noh, N. J., Ochoa-Hueso, R., Pathare,  
473 V. S., Pendall, E., Pihlblad, J., Piñeiro, J., Powell, J. R., Power, S. A., Reich, P. B., Renchon,  
474 A. A., Riegler, M., Rinnan, R., Rymer, P. D., Salomón, R. L., Singh, B. K., Smith, B.,  
475 Tjoelker, M. G., Walker, J. K. M., Wujeska-Klaue, A., Yang, J., Zaehle, S. and Ellsworth,  
476 D. S.: The fate of carbon in a mature forest under carbon dioxide enrichment, *Nature*,  
477 580(7802), doi:10.1038/s41586-020-2128-9, 2020.

478 Klamer, M. and Bååth, E.: Estimation of conversion factors for fungal biomass determination in  
479 compost using ergosterol and PLFA 18:2 $\omega$ 6,9, *Soil Biol. Biochem.*, 36(1),  
480 doi:10.1016/j.soilbio.2003.08.019, 2004.

481 Lavahun, M. F. E., Joergensen, R. G. and Meyer, B.: Activity and biomass of soil  
482 microorganisms at different depths, *Biol. Fertil. Soils*, 23(1), doi:10.1007/BF00335816,  
483 1996.

484 Liang, C., Amelung, W., Lehmann, J. and Kästner, M.: Quantitative assessment of microbial  
485 necromass contribution to soil organic matter, *Glob. Chang. Biol.*, 25(11),  
486 doi:10.1111/gcb.14781, 2019.

487 Locey, K. J. and Lennon, J. T.: Scaling laws predict global microbial diversity, *Proc. Natl. Acad.*

488 Sci. U. S. A., doi:10.1073/pnas.1521291113, 2016.

489 Malik, A. A., Chowdhury, S., Schlager, V., Oliver, A., Puissant, J., Vazquez, P. G. M., Jehmlich,  
490 N., von Bergen, M., Griffiths, R. I. and Gleixner, G.: Soil fungal: Bacterial ratios are linked  
491 to altered carbon cycling, *Front. Microbiol.*, 7(AUG), doi:10.3389/fmicb.2016.01247, 2016.

492 Moore, J. C. and William Hunt, H.: Resource compartmentation and the stability of real  
493 ecosystems, *Nature*, doi:10.1038/333261a0, 1988.

494 Moore, J. C., McCann, K., Setälä, H. and De Ruiter, P. C.: Top-down is bottom-up: Does  
495 predation in the rhizosphere regulate aboveground dynamics?, *Ecology*, doi:10.1890/0012-  
496 9658(2003)084[0846:TIBDPI]2.0.CO;2, 2003.

497 Romero-Olivares, A. L., Allison, S. D. and Treseder, K. K.: Soil microbes and their response to  
498 experimental warming over time: A meta-analysis of field studies, *Soil Biol. Biochem.*,  
499 doi:10.1016/j.soilbio.2016.12.026, 2017.

500 Rousk, J. and Bååth, E.: Fungal biomass production and turnover in soil estimated using the  
501 acetate-in-ergosterol technique, *Soil Biol. Biochem.*, 39(8), 2173–2177,  
502 doi:10.1016/j.soilbio.2007.03.023, 2007.

503 Rousk, J., Brookes, P. C. and Bååth, E.: Contrasting soil pH effects on fungal and bacterial  
504 growth suggest functional redundancy in carbon mineralization, *Appl. Environ. Microbiol.*,  
505 75(6), 1589–1596, doi:10.1128/AEM.02775-08, 2009.

506 Rousk, J., Bååth, E., Brookes, P. C., Lauber, C. L., Lozupone, C., Caporaso, J. G., Knight, R. and  
507 Fierer, N.: Soil bacterial and fungal communities across a pH gradient in an arable soil,  
508 *ISME J.*, 4(10), 1340–1351, doi:10.1038/ismej.2010.58, 2010.

509 Sanderman, J., Hengl, T. and Fiske, G. J.: Soil carbon debt of 12,000 years of human land use,  
510 *Proc. Natl. Acad. Sci. U. S. A.*, 114(36), doi:10.1073/pnas.1706103114, 2017.

511 Schimel, D., Stephens, B. B. and Fisher, J. B.: Effect of increasing CO<sub>2</sub> on the terrestrial carbon  
512 cycle, *Proc. Natl. Acad. Sci. U. S. A.*, doi:10.1073/pnas.1407302112, 2015.

513 Sengupta, A., Fansler, S. J., Chu, R. K., Danczak, R. E., Garayburu-Caruso, V. A., Renteria, L.,  
514 Song, H. S., Toyoda, J., Wells, J. and Stegen, J. C.: Disturbance triggers non-linear microbe-  
515 environment feedbacks, *Biogeosciences*, 18(16), doi:10.5194/bg-18-4773-2021, 2021.

516 Shi, Z., Crowell, S., Luo, Y. and Moore, B.: Model structures amplify uncertainty in predicted  
517 soil carbon responses to climate change, *Nat. Commun.*, doi:10.1038/s41467-018-04526-9,  
518 2018.

519 Six, J., Frey, S. D., Thiet, R. K. and Batten, K. M.: Bacterial and Fungal Contributions to Carbon  
520 Sequestration in Agroecosystems, *Soil Sci. Soc. Am. J.*, 70(2), 555,  
521 doi:10.2136/sssaj2004.0347, 2006.

522 Soares, M. and Rousk, J.: Microbial growth and carbon use efficiency in soil: Links to fungal-  
523 bacterial dominance, SOC-quality and stoichiometry, *Soil Biol. Biochem.*,  
524 doi:10.1016/j.soilbio.2019.01.010, 2019.

525 Strickland, M. S. and Rousk, J.: Considering fungal: Bacterial dominance in soils - Methods,  
526 controls, and ecosystem implications, *Soil Biol. Biochem.*,  
527 doi:10.1016/j.soilbio.2010.05.007, 2010.

528 Sulman, B. N., Phillips, R. P., Oishi, A. C., Shevliakova, E. and Pacala, S. W.: Microbe-driven  
529 turnover offsets mineral-mediated storage of soil carbon under elevated CO<sub>2</sub>, *Nat. Clim.*  
530 *Chang.*, doi:10.1038/nclimate2436, 2014.

531 Tagesson, T., Schurgers, G., Horion, S., Ciais, P., Tian, F., Brandt, M., Ahlström, A., Wigneron,  
532 J. P., Ardö, J., Olin, S., Fan, L., Wu, Z. and Fensholt, R.: Recent divergence in the  
533 contributions of tropical and boreal forests to the terrestrial carbon sink, *Nat. Ecol. Evol.*,

534 doi:10.1038/s41559-019-1090-0, 2020.

535 Tedersoo, L., Bahram, M., Pöhlme, S., Kõljalg, U., Yorou, N. S., Wijesundera, R., Ruiz, L. V.,  
536 Vasco-Palacios, A. M., Thu, P. Q., Suija, A., Smith, M. E., Sharp, C., Saluveer, E., Saitta,  
537 A., Rosas, M., Riit, T., Ratkowsky, D., Pritsch, K., Põldmaa, K., Piepenbring, M., Phosri,  
538 C., Peterson, M., Parts, K., Pärtel, K., Otsing, E., Nouhra, E., Njouonkou, A. L., Nilsson, R.  
539 H., Morgado, L. N., Mayor, J., May, T. W., Majuakim, L., Lodge, D. J., Lee, S., Larsson, K.  
540 H., Kohout, P., Hosaka, K., Hiiesalu, I., Henkel, T. W., Harend, H., Guo, L. D., Greslebin,  
541 A., Grelet, G., Geml, J., Gates, G., Dunstan, W., Dunk, C., Drenkhan, R., Dearnaley, J., De  
542 Kesel, A., Dang, T., Chen, X., Buegger, F., Brearley, F. Q., Bonito, G., Anslan, S., Abell, S.  
543 and Abarenkov, K.: Global diversity and geography of soil fungi, *Science* (80-. ),  
544 346(6213), doi:10.1126/science.1256688, 2014.

545 Terrer, C., Phillips, R. P., Hungate, B. A., Rosende, J., Pett-Ridge, J., Craig, M. E., van  
546 Groenigen, K. J., Keenan, T. F., Sulman, B. N., Stocker, B. D., Reich, P. B., Pellegrini, A. F.  
547 A., Pendall, E., Zhang, H., Evans, R. D., Carrillo, Y., Fisher, J. B., Van Sundert, K., Vicca,  
548 S. and Jackson, R. B.: A trade-off between plant and soil carbon storage under elevated  
549 CO<sub>2</sub>, *Nature*, 591(7851), doi:10.1038/s41586-021-03306-8, 2021.

550 de Vries, F. T., Manning, P., Tallowin, J. R. B., Mortimer, S. R., Pilgrim, E. S., Harrison, K. A.,  
551 Hobbs, P. J., Quirk, H., Shipley, B., Cornelissen, J. H. C., Kattge, J. and Bardgett, R. D.:  
552 Abiotic drivers and plant traits explain landscape-scale patterns in soil microbial  
553 communities, *Ecol. Lett.*, 15(11), 1230–1239, doi:10.1111/j.1461-0248.2012.01844.x, 2012.

554 Waldrop, M. P., Holloway, J. M., Smith, D. B., Goldhaber, M. B., Drenovsky, R. E., Scow, K.  
555 M., Dick, R., Howard, D., Wylie, B. and Grace, J. B.: The interacting roles of climate, soils,  
556 and plant production on soil microbial communities at a continental scale, *Ecology*,  
557 doi:10.1002/ecy.1883, 2017.

558 Wang, Q., Liu, S. and Tian, P.: Carbon quality and soil microbial property control the latitudinal  
559 pattern in temperature sensitivity of soil microbial respiration across Chinese forest  
560 ecosystems, *Glob. Chang. Biol.*, doi:10.1111/gcb.14105, 2018.

561 Waring, B. G., Averill, C. and Hawkes, C. V.: Differences in fungal and bacterial physiology  
562 alter soil carbon and nitrogen cycling: Insights from meta-analysis and theoretical models,  
563 *Ecol. Lett.*, 16(7), 887–894, doi:10.1111/ele.12125, 2013.

564 Wieder, W. R., Bonan, G. B. and Allison, S. D.: Global soil carbon projections are improved by  
565 modelling microbial processes, *Nat. Clim. Chang.*, 3(10), 909–912,  
566 doi:10.1038/nclimate1951, 2013.

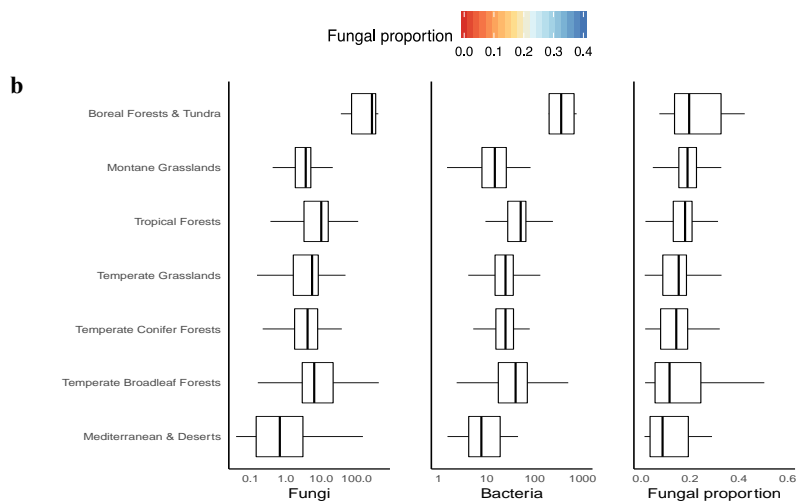
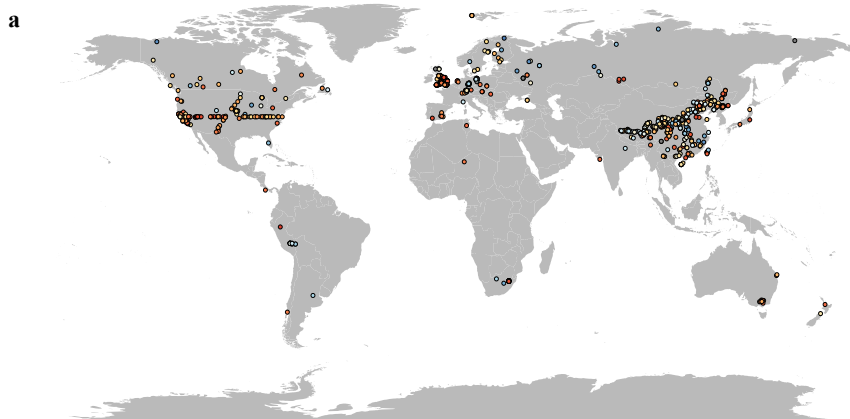
567 Wieder, W. R., Allison, S. D., Davidson, E. A., Georgiou, K., Hararuk, O., He, Y., Hopkins, F.,  
568 Luo, Y., Smith, M. J., Sulman, B., Todd-Brown, K., Wang, Y. P., Xia, J. and Xu, X.:  
569 Explicitly representing soil microbial processes in Earth system models, *Global*  
570 *Biogeochem. Cycles*, 29(10), 1782–1800, doi:10.1002/2015GB005188, 2015.

571 Yu, K. and D’Odorico, P.: Hydraulic lift as a determinant of tree-grass coexistence on savannas,  
572 *New Phytol.*, 207(4), 1038–1051, doi:10.1111/nph.13431, 2015.

573 Yu, K., Smith, W. K., Trugman, A. T., Condit, R., Hubbell, S. P., Sardans, J., Peng, C., Zhu, K.,  
574 Peñuelas, J., Cailleret, M., Levanic, T., Gessler, A., Schaub, M., Ferretti, M. and Anderegg,  
575 W. R. L.: Pervasive decreases in living vegetation carbon turnover time across forest climate  
576 zones, *Proc. Natl. Acad. Sci. U. S. A.*, doi:10.1073/pnas.1821387116, 2019.

577 Yue, H., Wang, M., Wang, S., Gilbert, J. A., Sun, X., Wu, L., Lin, Q., Hu, Y., Li, X., He, Z.,  
578 Zhou, J. and Yang, Y.: The microbe-mediated mechanisms affecting topsoil carbon stock in  
579 Tibetan grasslands, *ISME J.*, 9(9), doi:10.1038/ismej.2015.19, 2015.

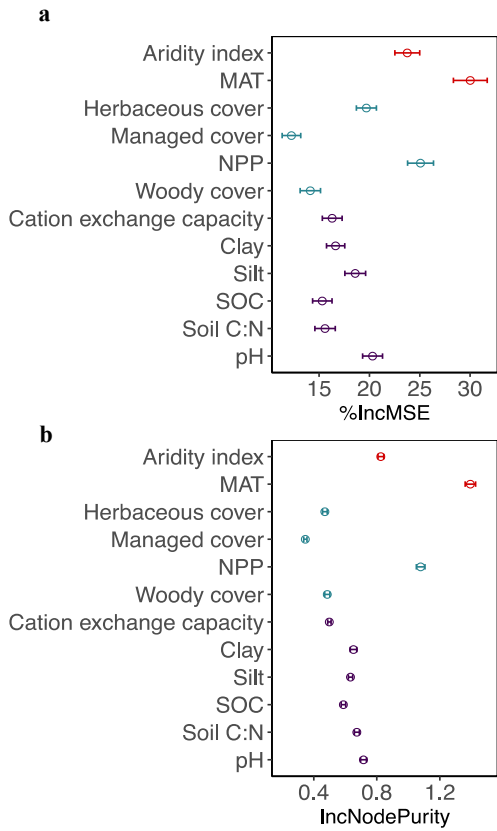
580 Zhu, Q., Riley, W. J. and Tang, J.: A new theory of plant-microbe nutrient competition resolves  
581 inconsistencies between observations and model predictions, *Ecol. Appl.*, 27(3),  
582 doi:10.1002/eap.1490, 2017.  
583



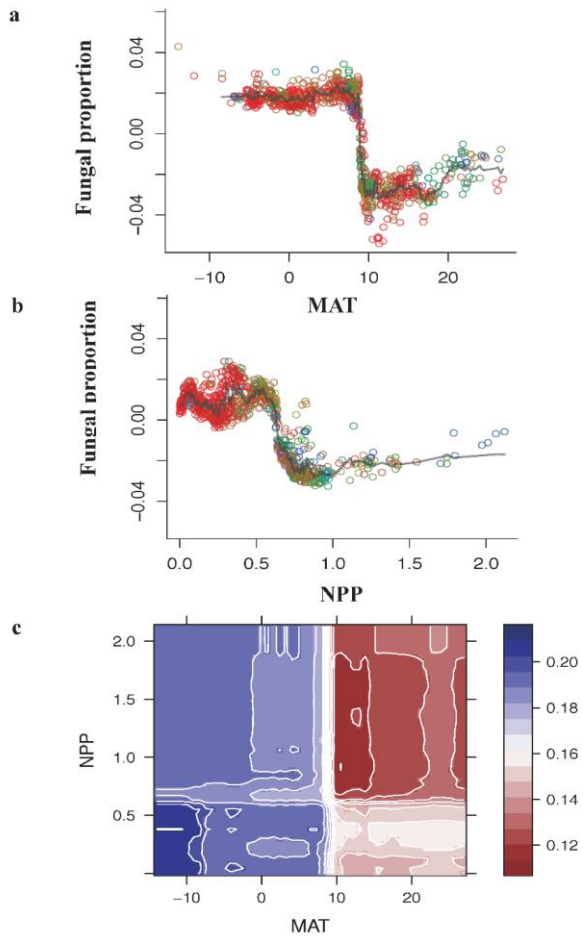
584 **Figure 1. Map of sample locations and fungal and bacterial abundance and fungal**  
 585 **proportion data. a**, Sampling sites. A total of 3224 samples were collected and aggregated into  
 586 943 1-km<sup>2</sup> pixels that were used for geospatial modelling. **b**, The median and interquartile range  
 587 of abundance of fungi and bacteria and fungal proportion across vegetation biomes. Tundra and  
 588 boreal forest, Mediterranean and desert have low sample sizes (<25) and thus were combined.  
 589

设置了格式: 上标



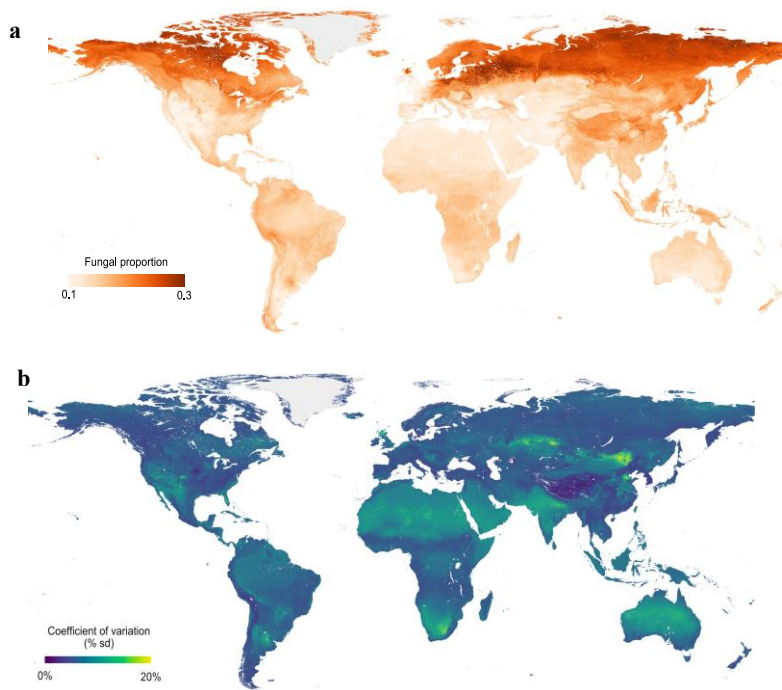


590  
 591 **Figure 2. Mean decrease in accuracy (%IncMSE, mean and SD, a) and mean decrease gini**  
 592 **(IncNodePurity, mean and SD, b) estimated from 1000 simulations of random forests. This**  
 593 **is used to evaluate the importance of top environmental drivers on proportion of fungi derived**  
 594 **from ‘all’ dataset.**



595  
 596 **Figure 3. Fungal proportion is primarily associated with climate- mean annual**  
 597 **temperature (MAT) and net primary productivity (NPP). a–b, Partial feature contributions**  
 598 **of primary environmental variables (a, MAT; b, NPP) to fungal proportion. c, Partial feature**  
 599 **contributions of primary environmental variable interactions (MAT vs NPP) to fungal**  
 600 **proportion.**

601



602  
 603  
 604 **Figure 4.** Global map of fungal proportion (a) and bootstrapped (100 iterations) coefficient  
 605 of variation (b) at the 30 arcsec (approximately 1 km<sup>2</sup>) pixel scale. Bootstrapped coefficient  
 606 of variation is standard deviation divided by the mean predicted value as a measure of prediction  
 607 accuracy. Sampling was stratified by biome.

设置了格式: 上标

608  
 609  
 610

611 Author contributions

612 KLY and TWC designed the project. KLY built the PLFA datasets with help from JVDH and

613 ZQW. KLY performed the analysis with inputs from DR and CA. KLY, CA, and TWC wrote the

614 paper with revisions from all other coauthors. GRS, RED, KMS, FM, MPW, YHY, FTDV,

615 RDB, PM, FB, SGB, EMB, CG, QKW, LM, BD C, XJH, WZT, ST, AH, JAB contributed to

616 PLFA datasets.

617

设置了格式: 英语(美国)

设置了格式: 英语(美国)

设置了格式: 英语(美国)