A comprehensive dataset of microbial abundance, dissolved organic carbon, and nitrogen in Tibetan Plateau glaciers

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Abstract. Glaciers are recognized as a biome dominated by microorganisms and a reservoir of organic carbon and nutrients. Global warming remarkably increases glacier melting and runoff, which has a significant impact on the carbon and nitrogen

15   cycle of downstream ecosystems. The Tibetan Plateau (TP), dubbed "the water tower of Asia", owns the largest mountain glacial area at mid- and low-latitudes. However, limited data of microbial abundance, organic carbon, and nitrogen in TP glaciers have been reported, which severely hinders our understanding of the regional carbon and nitrogen cycle. This work constructed a new dataset on microbial abundance, dissolved organic carbon (DOC), and total nitrogen (TN) for TP glaciers. In this dataset, there are 5,409 records from 12 glaciers for microbial abundance of ice cores and snow pits, and there are

20   2,556 records from 40 glaciers for DOC and TN of five habitats, including ice core, snow pit, surface ice, surface snow, and proglacial runoff. These glaciers cover diverse geographic and climatic regions, where the multiyear average temperature range from -13.4 °C to 2.9 °C and the multiyear average precipitation range from 76.9 mm to 927.8 mm. This makes the constructed dataset qualified for large-scale researches across the TP. To the best of our knowledge, this is the first dataset of microbial abundance and TN in TP glaciers and also the first dataset of DOC in ice cores on the TP. This new dataset

25   provides important information for the studies on carbon and nitrogen cycle in glacial ecosystems, especially for the assessment of potential impacts of glacier retreat on downstream ecosystems under global warming. The datasets are available   from   the   National   Tibetan   Plateau/Third   Pole   Environment   Data   Center (https://doi.org/10.11888/Cryos.tpdc.271841, Liu, 2021).

1 Introduction

30   Glaciers are not only an important component of the global climate system, but also recognized as a biome dominated by microorganism and a reservoir of organic material and nutrients (Anesio and Laybourn-Parry, 2012; Hood et al., 2015).

There exist abundant and active microorganisms in multiple types of habitats such as surface snow and ice, cryoconite holes, proglacial runoff, the englacial ice and subglacial sediments despite the harsh environmental conditions (e.g. sustained low temperature, lack of nutrition and strong radiation) (Anesio et al., 2017; Hodson et al., 2008). These glacial microorganisms play an important role in the carbon cycle of glacial ecosystem and draw increasing attention in recent years (Liu et al., 2016a; Egge et al., 2021).

Due to the carbon fixation of glacial microorganisms and atmospheric deposition, glaciers are large pools of biologically available carbon and nitrogen, two indispensable elements to life (Hodson et al., 2008; Dubnick et al., 2017), and serve as important material and energy sources for downstream aquatic ecosystems, which is especially important for the nutrient-poor ecosystems like the sparsely vegetated high-alpine valleys and oligotrophic alpine lakes (Hood et al., 2015). Glaciers are also key species pools for downstream aquatic ecosystems and it has been found that the downstream bacterial and fungal communities were significantly affected by glacier-originating taxa (Fell et al., 2021; Liu et al., 2021). Under global climate warming, global glaciers are melting at an accelerating rate. Therefore, the carbon, nitrogen and microorganisms in glaciers are becoming more and more important for downstream ecosystems, and need more investigations and studies (Hood et al., 2009; Singer et al., 2012; Fellman et al., 2015; Wadham et al., 2016).

The Tibetan Plateau (TP), dubbed "the water tower of Asia", owns the largest mountain glacial area at mid- and low-latitudes (Yao et al., 2012). Glaciers on the TP are the sources of large rivers in Asia and thus of great importance for regional environments (Yao et al., 2019). However, due to harsh climate conditions and poor traffic accessibility, there have been limited observations of organic carbon, nitrogen and microorganisms in TP glaciers. In particular, very few such data in ice cores has been published, and even for surface snow and ice data, the spatial coverage is poor (Hood et al., 2015; Liu et al., 2016b). Data scarcity significantly hindered our understanding on the biogeochemical cycle in TP glaciers and the surrounding downstream regions.

In this study, we aim to construct a comprehensive dataset of microorganisms, dissolved organic carbon (DOC), and total nitrogen (TN) in TP glaciers by compiling deep ice-core samples and extensive surface snow and ice samples, which can provide valuable data for the researches on glacier biogeochemical cycle and the impact of glacier retreat on downstream ecosystems.

2 Study area

The TP covers an area of about 2.5 million km^2, with an average altitude of more than 4000 m above sea level. Climate over the TP is primarily influenced by the interaction between the Asian monsoon and the westerly wind (Tian et al., 2001). The TP and its surrounding regions contain the largest number of glaciers outside the poles with an area of about 47 thousand km^2 (Yao et al., 2012). These glaciers are important water resources for downstream areas and play a crucial role in regional water supply (Immerzeel et al. 2010). The glaciers on the TP are mainly distributed in the Kunlun, Nyainqntanglha, Himalayas and Karakoram mountains, and most glaciers are located between 4500-6500 m above sea level (Liu et al., 2015).

The glaciers can be divided into three types (Shi et al., 2000): marine glaciers (mainly distributed in the southeast of TP),
65 subcontinental glaciers (mainly distributed in the northeast and southern margin of TP), and the continental glaciers (mainly
distributed in the west of TP). Most of the TP glaciers except those in the Karakoram region are experiencing strong retreat
under climate warming (Yao et al., 2012; Wang et al., 2021).

3 Sample distribution and laboratory measurements

3.1 Sample distribution

70 In this study, 5,409 microbial abundance samples from 12 glaciers across the TP were collected as shown in Fig. 1(a),
including 5,210 ice core samples from 7 glaciers and 199 snow-pit samples from 7 glaciers. For DOC and TN, 2,556 samples
from 40 glaciers were collected as shown in Fig. 1(b), including 1,625 ice core samples from 7 glaciers, 180 surface ice
samples from 17 glaciers, 100 snow pit samples from 4 glaciers, 254 surface snow samples from 28 glaciers, and 397 runoff
samples from 16 glaciers.
75 The sampled glaciers covered diverse climate conditions. The multiyear average temperature ranged from -13.4 ℃ (the
Guliya glacier) to 2.9 ℃ (the Zhuxigou glacier), and the multiyear average precipitation ranged from 76.9 mm (the No.15
glacier) to 927.8 mm (the 24K glacier). These conditions cover the main types of environments for glaciers on the TP, which
made the dataset comprehensive and representative.

3.2 Sample distribution

80 Ice cores were drilled from the accumulation zones of nine glaciers to depths from 11 to 173 m (Fig. 1). Both microbial
abundance and DOC/TN samples were collected in five glaciers (i.e. Muztagh Ata (MSTG), Cuopugou (CPG), Zuoqiupu
(ZQP), Noijin Kangsang (NJKS), and East Rongbuk (ERB)), there were only microbial abundance samples in the Laohugou
(LHG) and Geladandong (GLDD) glacier, and there were only DOC and TN samples in the Muji (MJ) and Dunde (DD)
glacier. The MJ, MSTG, LHG, DD, and GLDD glacier are mainly influenced by the westerly, while the CPG, ZQP, NJKS,
85 and ERB glacier are strongly influenced by monsoon. The samples were transported frozen to the laboratory.
Snow pits were digged at the accumulation zones of seven glaciers (Fig 1). Both microbial abundance and DOC/TN samples
were collected in the GLDD, Zhadang (ZD), Palon No.4 (PL4), and ERB glacier, and there were only microbial abundance
samples in the DD, Mengdagangri (MDGR), and Yala glacier. Fourteen snow pits in total were sampled for microbial
abundance measurement, including six at the ZD glacier during April, May, June, August, September and October in 2006,
90 two at the MDGR glacier during 2006 and 2007, two at different altitudes at the PL4 glacier, and one for each at the DD,
GLDD, ERB, and Yala glacier, respectively. Four snow pits were sampled for DOC and TN concentration measurement
with one snow pit in each glacier among GLDD, ZD, PL4 and ERB. At each pit, snow was sampled from top to bottom at 5
or 10 cm interval using steel scoop.

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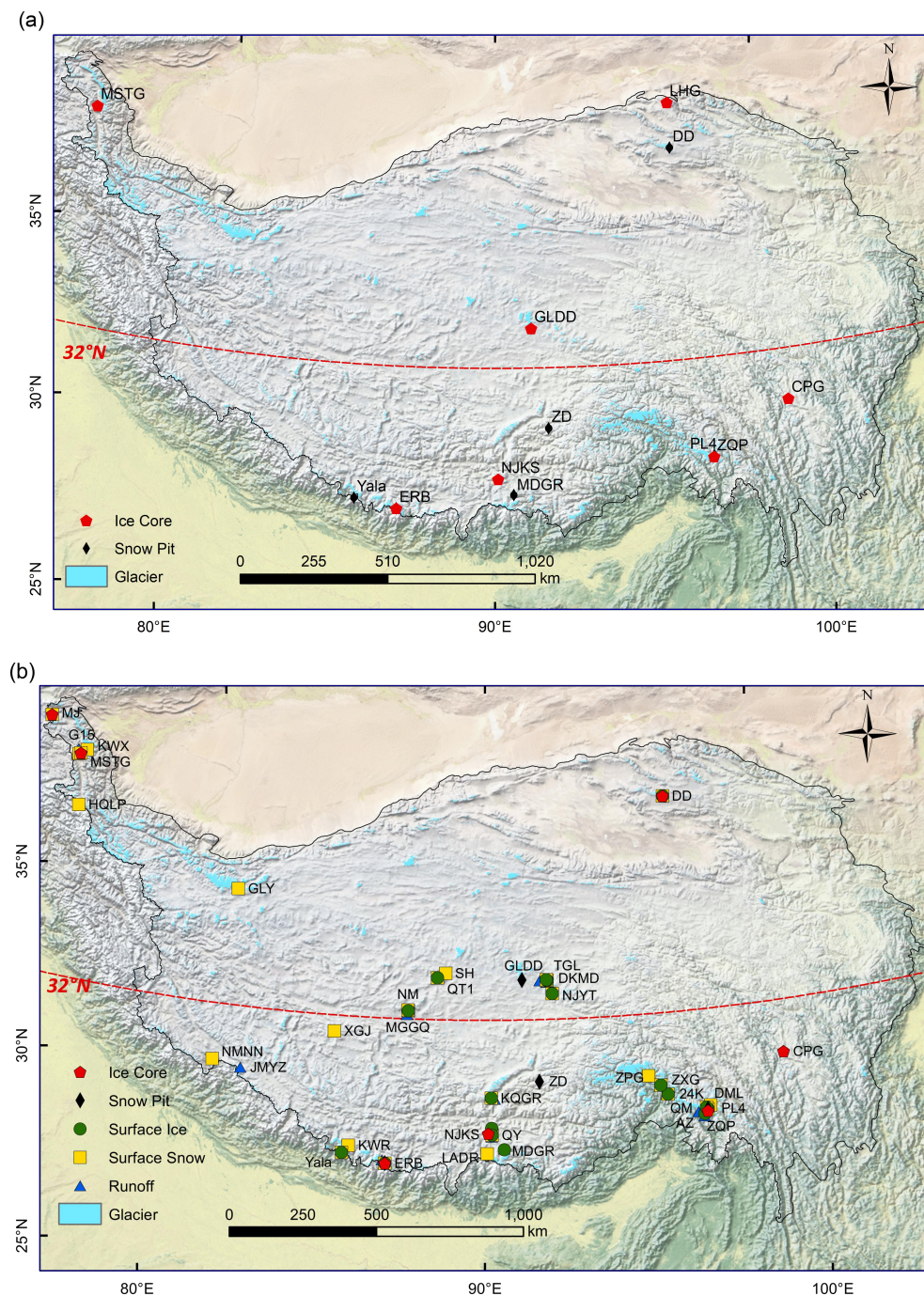
Figure 1: Location of sampled glaciers for microbial abundance (a), DOC and TN (b) on the Tibetan Plateau. The abbreviations of glacier names were labelled in the map and the full names were available in the supplement.
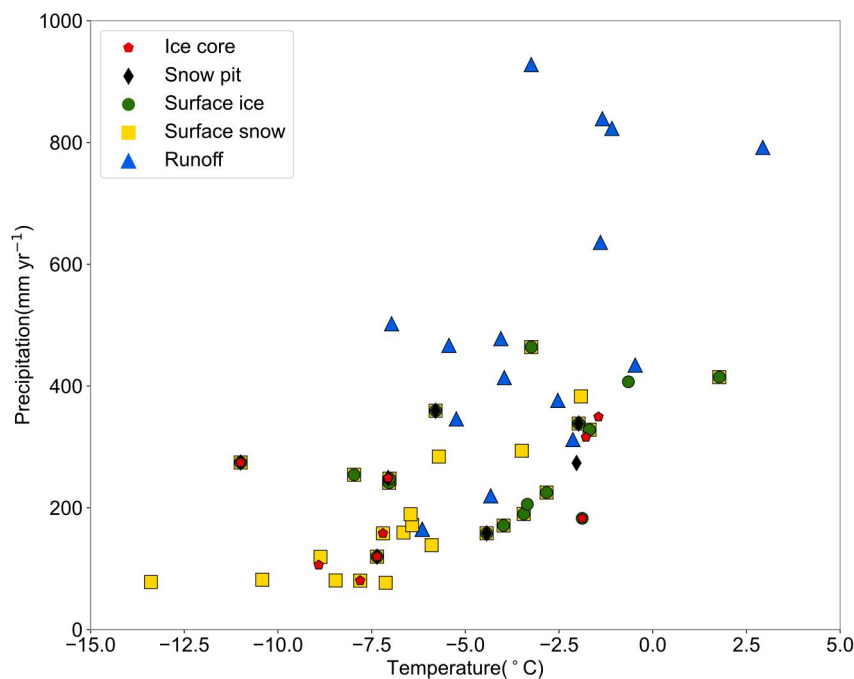
Figure 2: Muti-year average temperature and precipitation of sampled glaciers.

Surface snow (within 10 cm deep) were sampled using steel scoop at different altitudes from ablation to accumulation zone in 28 glaciers for DOC and TN concentration measurement. Surface ice samples were collected with a precleaned ice axe at the ablation zones in 17 glaciers. Ice and snow samples were stored in well-sealed WhirlPak bags (WhirlPak®, Nasco, USA) and transported to laboratory under frozen conditions.

Water samples were collected from proglacial runoff of 16 glaciers using 100 mL polycarbonate bottles pre-cleaned using 1% HCl (Nalgene, USA). Totally eight sets of 24-h water samples were collected at six glaciers (i.e. MSTG, Tanggula Longxiazailongba (TGL), Qiangtang No.1 (QT1), Qiangyong (QY), Tonkmadi (DKMD), and PL4) to investigate the diurnal variation of conductivity, pH, DOC, NO3- and TN of water. Water samples were transported in a dark container with ice keeping cold. In the laboratory, samples were kept in dark and frozen at -20°C until analysis.

3.3 Laboratory measurements

Each ice core was cut into 5-10 cm long sections in a -20 °C clean room. After cutting away the outer 1-2 cm annulus with knife, the inner ice was used. Knife and containers were pre-cleaned using 1% HCl and filter water. Snow and ice were melted overnight and the meltwater were aliquoted into 20 mL glass bottles. The glass bottles were leached using 1% HCl, rinsed by deionized water for 3 times and combusted (450 °C for > 3 h) before use.

For the enumeration of bacteria, 1.98 mL of meltwater was fixed with glutaraldehyde (final concentration: 1%), stored at 4 °C, and analysed within 8 hours after staining with SYBR Green I (Company, Country) (Marie et al., 1997). Samples were

processed on an EPICS ALTRA II flow cytometer (Beckman Coulter, USA) (Liu et al., 2016a). Duplicate samples were
measured with a relative standard deviation lower than 10%. Flow cytometry data were collected and analysed with
120   CytoWin 4.31 software.

The DOC and TN concentrations were measured with a TOC-Lcph (Shimadzu Corp., Japan) following standard methods
(Greenberg et al., 1992). Concentrations of ions were measured using a Dionex ion Chromatograph System 2000 (Dionex
Corp, USA) as previously described (Liu et al., 2016a).

4 Data description of microbial abundance

125   ## 4.1 Snow pit

The microbial abundance in snow pits of the seven sampled glaciers (i.e. DD, GLDD, ZD, PL4, MDGR, Yala, and ERB)
ranged from 212 to 721,305 cells mL^{-1}, with a mean value of 80,191±160,812 cells mL^{-1}. The range of these measurements
was consistent with the results of existing researches using the flow cytometer method (e.g. 3.7-25.0×10^4 cells mL^{-1} in the
Kuytun 51 Glacier, Tianshan Mountains) (Xiang et al., 2009). Fig. 3(a) shows the spatial distribution of log(microbial
130   abundance) for sampled snow pits averaged in each glacier. Generally, there were lower microbial abundance in the westerly
region than in the monsoon region. The DD glacier, located in the northeast of the TP, had the lowest microbial abundance
(i.e. 2,177 cells mL^{-1}), while the ZD glacier, located in the south of the TP, had the highest abundance (i.e. 218,305 cells mL^{-1}), which was 100 times higher as that in DD. Figure 4 shows the variation of log(microbial abundance) with depth in each
snow pit, but no consistent patterns were found.
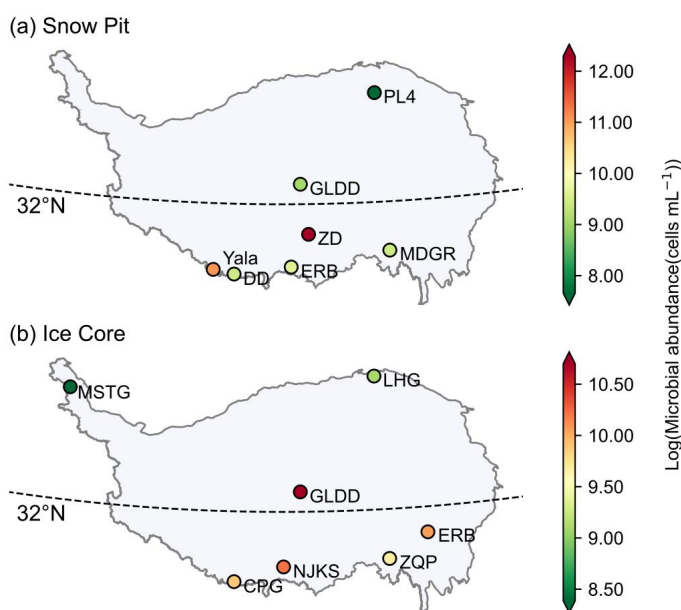
Figure 3: The spatial distribution of log(microbial abundance) for sampled snow pits (a) and ice cores (b) averaged in each glacier.

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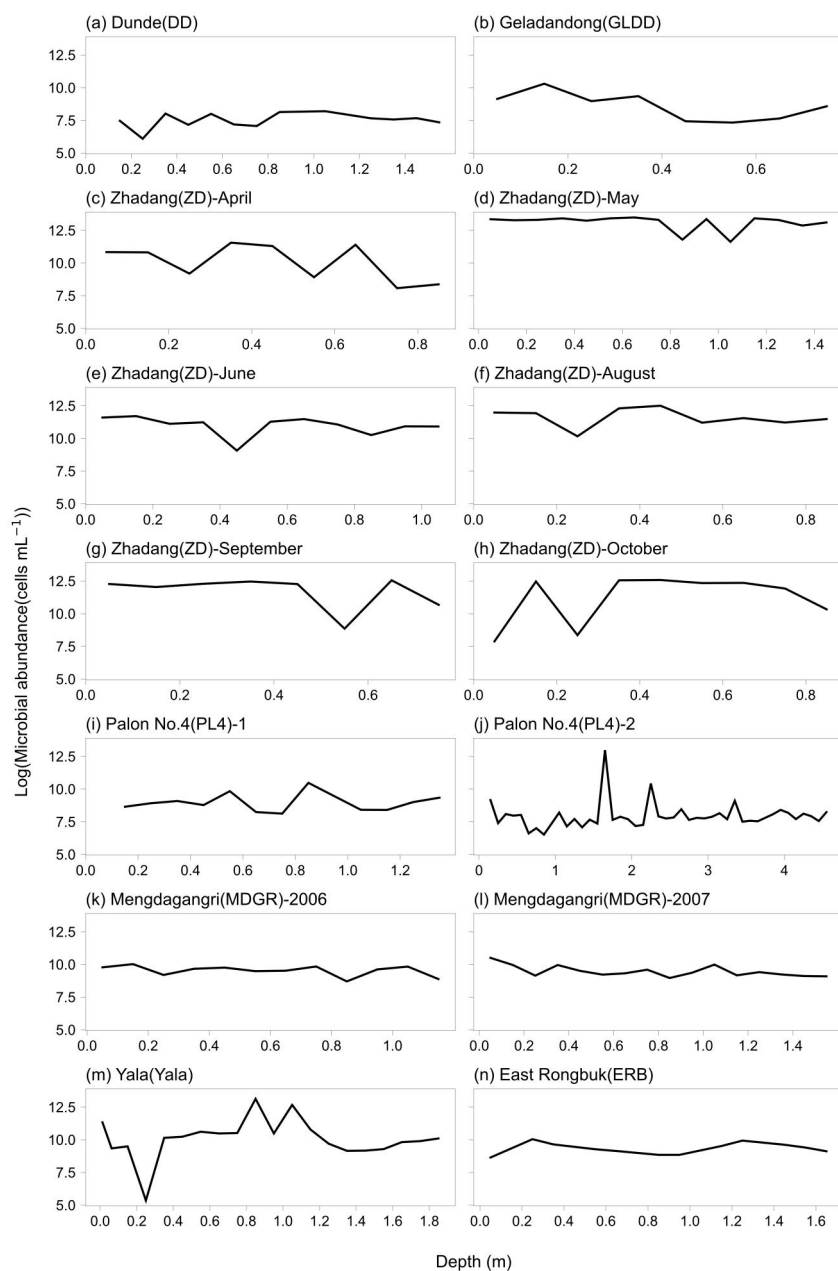
Figure 4: The variation of log(microbial abundance) with depth in each sampled snow pit.

4.2 Ice core

140   The microbial abundance in ice cores of the seven sampled glaciers (i.e. MSTG, LHG, GLDD, CPG, ZQP, NJKS, and ERB) had a wide range from 63 to 1,130,080 cells mL^{-1} with a mean value of 12,675±39,343 cells mL^{-1}. These values are on the same order of magnitude as existing researches using the flow cytometer method (e.g. 6.53×10^3 - 2.89×10^5 cells mL^{-1} in

the West Antarctic Ice Sheet Divide ice core) (Santibanez et al., 2016). The spatial distribution of average log(microbial abundance) in each ice core was shown in Fig. 3(b). Two ice cores in the north (i.e. MSTG and LHG) had obvious lower

145   microbial abundance (i.e. 4,389 and 8,958 cells mL^{-1}, respectively) than those in the south where the microbial abundance was at least 15,688 cells mL^{-1}. Fig. 4 showed the variation of log(microbial abundance) along depth in each ice core. The microbial abundance in ice cores in the CPG and NJKS glacier had decreasing trends with depth, while that in the MSTG glacier had an increasing trend. There existed low-frequency fluctuations for the microbial abundance in ice cores in the ERB, GLDD, and LHG glacier, and there were mainly high-frequency fluctuations in the ZQP glacier.
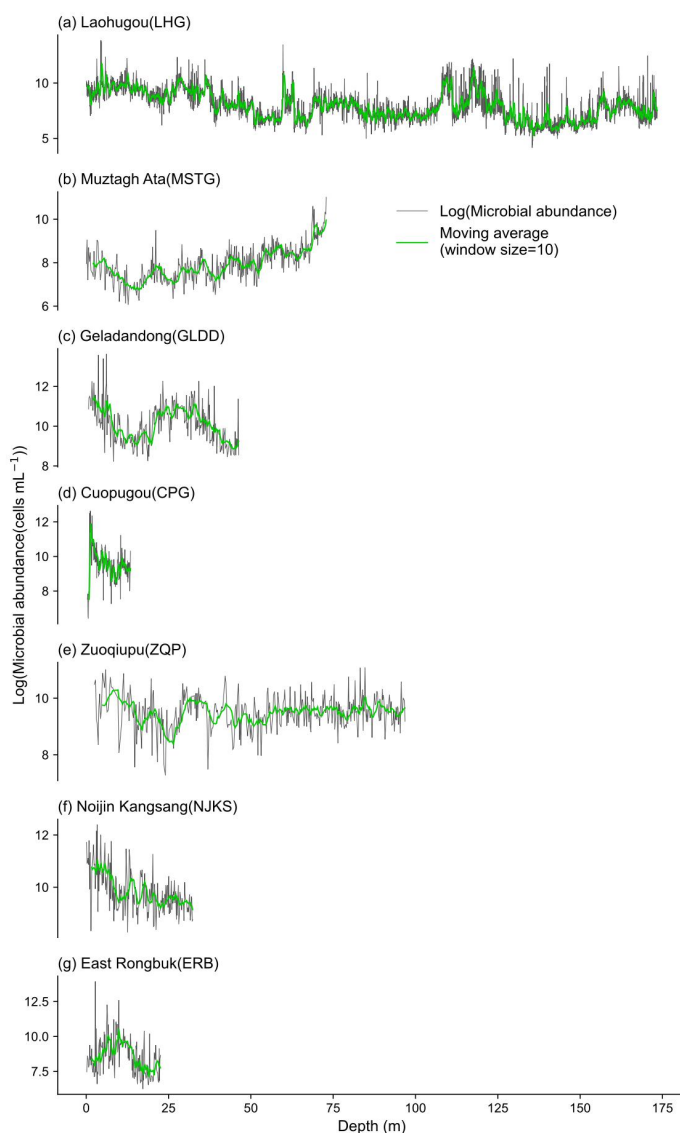

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Figure 5: The variation of log(microbial abundance) along depth in seven ice cores. The window size used in the moving average method was 10.

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5 Data description of DOC and TN

5.1 Ice core

155 The DOC concentrations in ice core samples ranged from 0.005 to 5.05 mg L^{-1} with an average value of 0.54±0.38 mg L^{-1}. These values are larger than the englacial DOC concentrations in global mountain glaciers reported by Hood et al. (2015) (0.01 to 1.20 mg L^{-1}, 0.29±0.03 mg L^{-1} on average), which may be related to the higher aerosol concentration in the area around the TP (Spracklen et al., 2011). The TN concentrations ranged from 0.001 to 1.15 mg L^{-1} with an average value of 0.24±0.16 mg L^{-1}. Fig. 6 shows the variation of DOC and TN concentrations along vertical profiles in ice cores of the MSTG

160 and ERB glacier. Generally, there are decreasing trends of DOC and TN with depth, suggesting that atmospheric deposition has been increasing in recent years. There exist large inter-annual variations in the serial data with some occasional large values, which may be related to historical large sand storms.
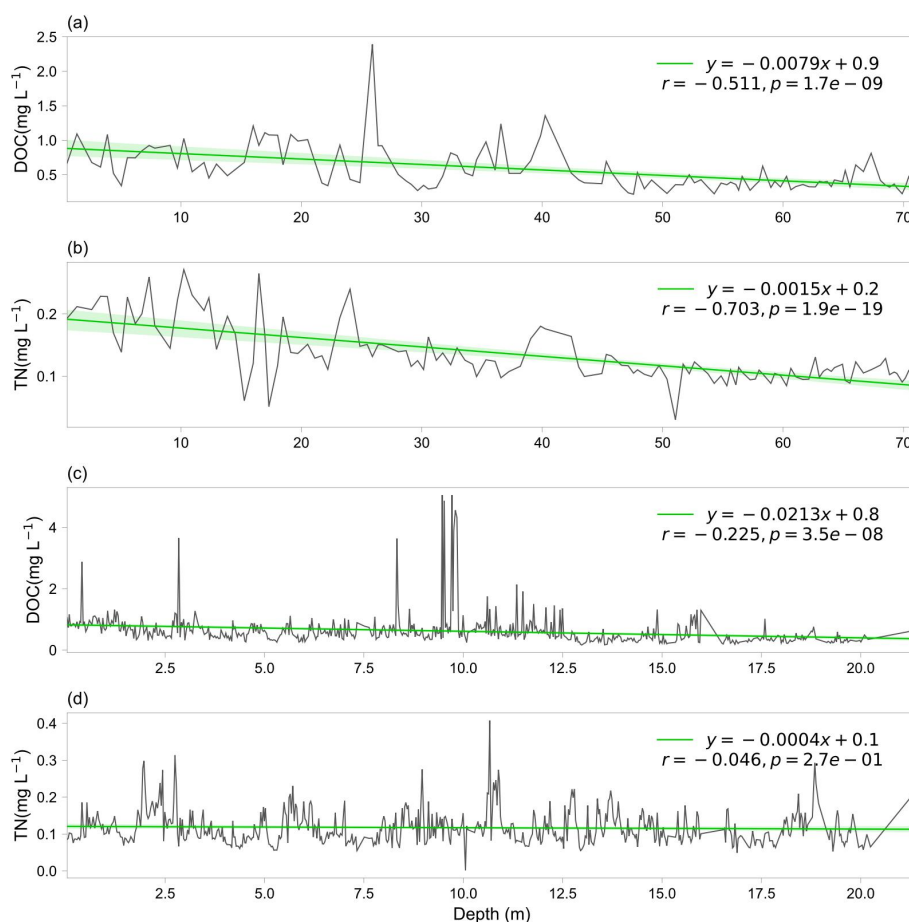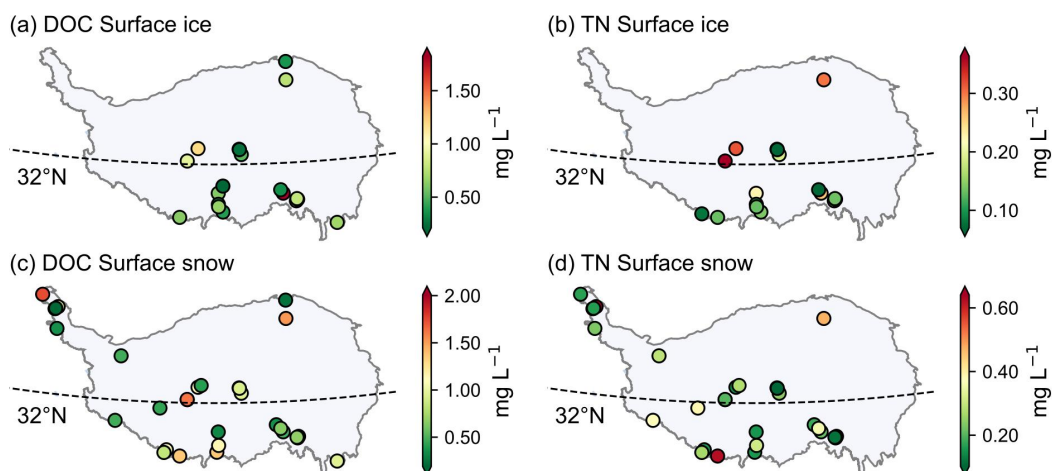

Figure 6: Variation of DOC (a, c) and TN (b, d) concentrations along vertical profiles in ice cores of the MSTG (Muztagh Ata)
165 **glacier (a, b) and the ERB (East Rongbuk) glacier (c,d).**

5.2 Surface ice, surface snow and snow pit

The DOC concentrations ranged from 0.08 to 9.0 mg L^{-1} (0.89±1.05 mg L^{-1}), from 0.12 to 11.65 mg L^{-1} (1.19±1.78 mg L^{-1}), and from 0.05 to 16.15 mg L^{-1} (0.72±1.71 mg L^{-1}) in surface ice, surface snow, and snow pits, respectively. These values are comparable to the DOC concentrations of surface ice in four TP glaciers reported by Liu et al. (2016) (i.e. 1.01±0.22 mg L^{-1}),

170   and those of surface snow (mean values ranging from 0.16 to 1.17 mg L^{-1}) and snow pits (mean values ranging from 0.21 to 0.81 mg L^{-1}) in TP glaciers summarized by Gao et al. (2020). The TN concentrations ranged from 0.01 to 1.88 mg L^{-1} (0.19±0.22 mg L^{-1}), from 0.07 to 3.06 mg L^{-1} (0.34±0.35 mg L^{-1}), and from 0.02 to 0.84 mg L^{-1} (0.15±0.15 mg L^{-1}) in surface ice, surface snow, and snow pits, respectively. Fig. 7 shows the spatial distribution of DOC and TN concentrations for surface snow and surface ice. For the DOC concentrations of surface snow, there is a decreasing pattern from south to north

175   in the monsoon dominant region (i.e. located on the south the 32°N). The DOC and TN concentrations in the westerly and monsoon regions were not significantly different according to the Mann Whitney U test. The differences of TN concentrations between the two regions were evident with higher values in the westerly region although the Mann Whitney U test was not significant with a p value of 0.056.

180   **Figure 7: The spatial distribution of DOC (a, c) and TN (b, d) concentrations for surface ice (a, b) and surface snow (c, d).**
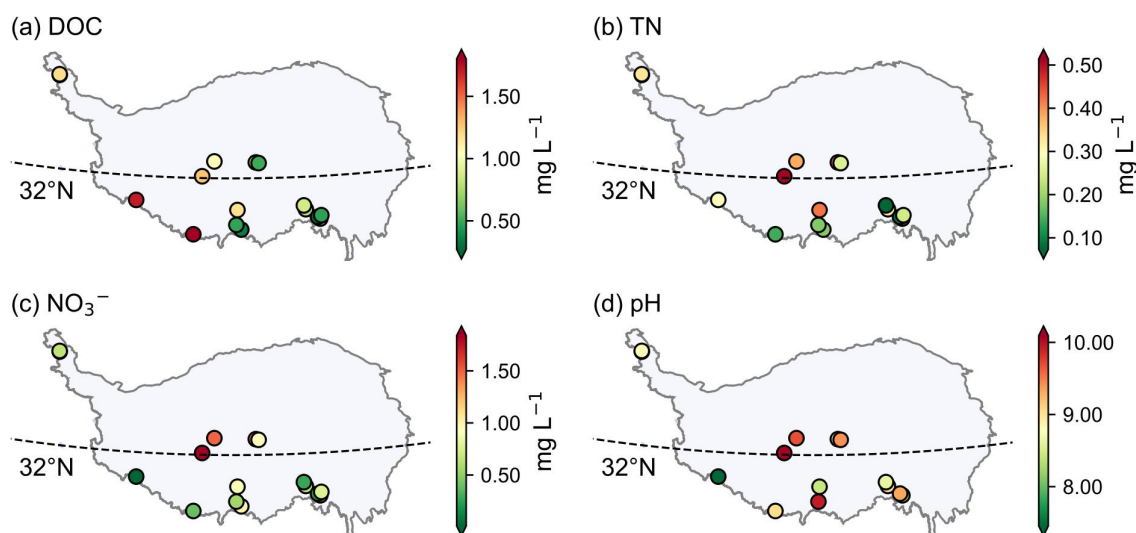
5.3 Runoff

The DOC concentrations ranged from 0.16 to 4.94 mg L^{-1} (0.79±0.68 mg L^{-1}) in proglacial runoff, which are consistent with the range from 42 glaciers worldwide (0.10 - 3.40 mg L^{-1}) summarized by Li et al. (2017). The TN concentration range from 0.05 to 2.3 mg L^{-1} (0.29±0.22 mg L^{-1}). The runoff water is alkaline with pH values ranging from 7.3 to 12.4 (9.10±0.88). Fig.

185   8 shows the spatial distribution of DOC, TN, NO$_3^-$ concentrations and pH for proglacial runoff on the TP. For the DOC concentrations of runoff, there is decreasing trend from west to east. There were significant differences in the TN and NO$_3^-$ concentrations of runoff between the westerly and monsoon regions with p values less than 0.01 according to the Mann

Whitney U test and the values in the westerly region were higher. The pH values in the westerly region were also larger in the westerly region than in the monsoon region although the Mann Whitney U test was not significant with a p value of 0.07.

190   Fig. 9 shows the diurnal variation of DOC, TN, NO_3^-, and conductivity in proglacial runoff of the Qiangyong glacier. There did not exist obvious patterns in the diurnal curve of DOC, while there were obvious unimodal patterns for TN, NO_3^-, and conductivity. These observations are very helpful for biogeochemical studies at fine temporal resolutions.

Figure 8: The spatial distribution of DOC (a), TN (b), NO_3^- (c) concentrations and pH (d) for proglacial runoff on the
195   **Tibetan Plateau.**
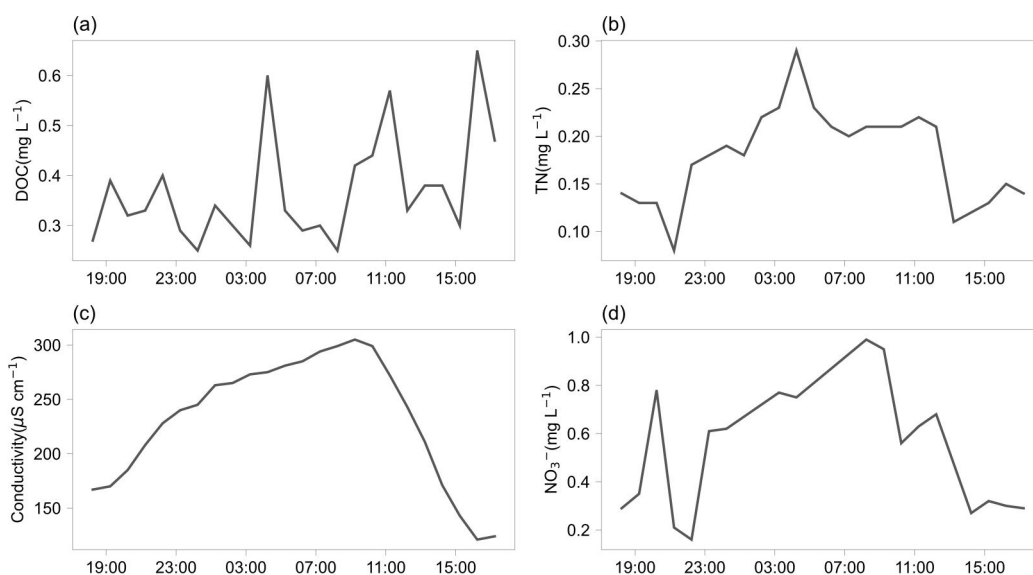

Figure 9: The 24-h time series of DOC (a), TN (b), conductivity (c), and NO_3^-(d) in proglacial runoff of the Qiangyong glacier.

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Discussions

5.4 Comparison among different habitats

Considering that observations of DOC and TN concentrations were available in all the five main glacial habitats (i.e. ice core,
200   snow pits, surface ice, surface snow, and proglacial runoff), differences among glacial habitats were compared. The boxplots
of different types of samples showed that the DOC and TN concentrations in ice cores were lower than those in surface ice
(Fig. 10). The Mann-Whitney-Wilcoxon test showed the difference was significant for DOC with a p value of 0.02 and not
significant for TN with a p value of 0.24. The DOC and TN concentrations were also lower in snow pits than in surface snow.
The difference was not significant for DOC with a p value of 0.19 and was significant for TN with a p value of 0.03. The
205   DOC and TN concentrations in proglacial runoff were similar to those in surface snow.
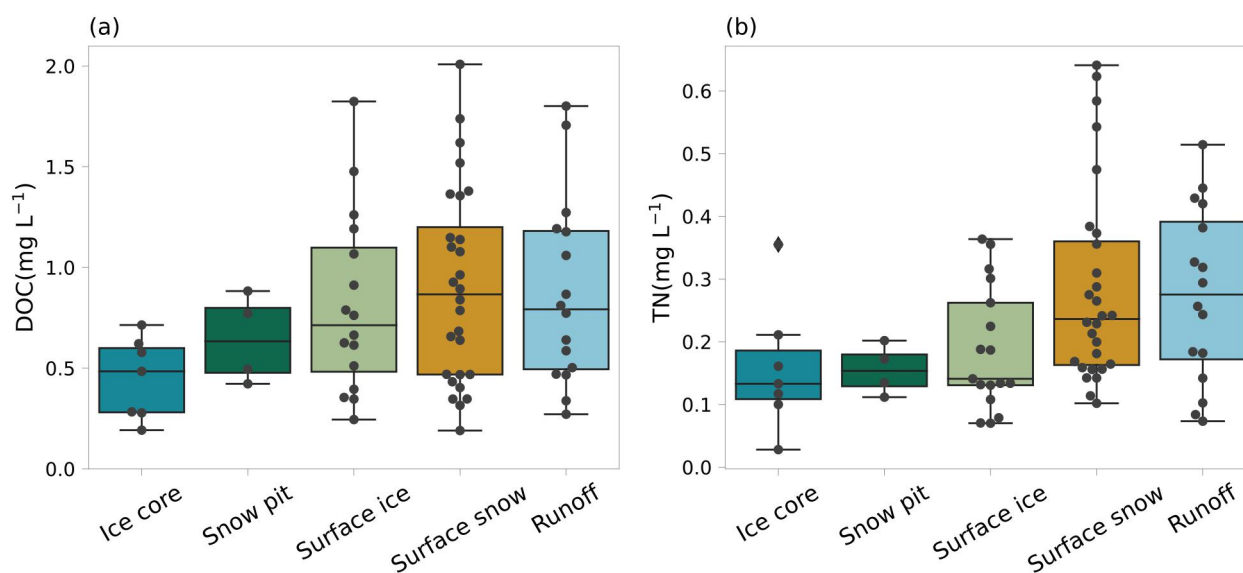
Figure 10: The boxplots of the DOC (a) and TN (b) concentrations in five main glacial habitats on the Tibetan Plateau.

6 Data availability

The dataset of microbial abundance, DOC and TN in ice core, snow pit, surface ice and snow from the Tibetan Plateau
210   glaciers are accessible at the National Tibetan Plateau/Third Pole Environment Data Center
(https://doi.org/10.11888/Cryos.tpdc.271841, Liu, 2021).

7 Conclusions

We constructed a new dataset of microbial abundance, DOC and TN for glaciers on the TP, comprising 5,409 microbial
abundance data from 12 glaciers and 2,556 DOC and TN data from 40 glaciers. The sampled glaciers cover diverse
215   geographic and climatic regions, which makes it qualified for large-scale researches across the TP. This systematic dataset

can provide important information for the studies on carbon and nitrogen cycle in glacial ecosystems, their response to global climate change, and their impact on downstream ecosystems such as glacier-feed streams and lakes. Furthermore, this dataset can be used as a basis for the global estimation of glacier microbial biomass, carbon, and nitrogen storage, which are fundamental information for forecasting the impact of warming on carbon cycle at a global scale.

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