Optical and biogeochemical properties of Belgian inland and coastal waters

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Abstract. From 2017 to 2019, an extensive sampling campaign was conducted in Belgian inland and coastal waters, aimed at providing paired data of optical and biogeochemical properties to support research into optical monitoring of aquatic systems. The campaign was focused on inland waters, with sampling of four lakes and a coastal lagoon along the growth season, in addition to samples of opportunity of other four lakes. Campaigns also included the Scheldt estuary over a tidal cycle and two sampling campaigns in the Belgian coastal zone. Measured parameters include inherent optical properties (absorption, scattering and attenuation coefficients, near-forward volume scattering function, turbidity), apparent optical properties (Secchi disk depth, substrate and water-leaving Lambert-equivalent bi-hemispherical reflectance), and biogeochemical properties (suspended particulate matter, mineral fraction of particle mass, particle size distribution, pigment concentration, DNA metabarcoding, flow microscopy counts, and bottom type classification). The diversity of water bodies and environmental conditions covered a wide range of system states. The chlorophyll a concentration varied from $0.63 \ \mathrm{mg \, m^{-3}}$ to $382.72 \ \mathrm{mg \, m^{-3}}$, while the suspended particulate matter concentration varied from $1.02 \ \mathrm{g \, m^{-3}}$ to $791.19 \ \mathrm{g \, m^{-3}}$, with mineral fraction varying from 0 to 0.95. Depending on system and season, phytoplankton assemblages were dominated by cyanobacteria, green algae (Mamiellophyceae, Pyramimonadophyceae) or diatoms. The dataset is available from Castagna et al. (2022), https://www.pangaea.de/tok/c67200d99ea9bbbeadd9edec9690f937b5bacbff.

1 Introduction

Datasets of paired optical and biogeochemical properties are essential for developing and validating the interpretation of optical signals captured with *in situ* instrumentation or remote sensors. Though the data gathered in the last 50 years provide a large collection of conditions across a diverse set of environments, three major caveats are observed in the freely accessible datasets:

1. The majority of the data concerns open oceans and coastal waters, with little representation of inland water systems;

2. The majority of the data concerns multispectral measurements, particularly at wavebands typical of ocean colour sensors;

3. The majority of the paired biogeochemical data includes only broad features (*e.g.*, chlorophyll *a* concentration) deemed suitable for operational retrievals with multispectral instruments.

In order to fully exploit the potential of hyperspectral satellite missions, hyperspectral datasets paired with detailed composition information of aquatic systems are required (Dierssen et al., 2020). In addition, more extensive data is necessary to develop and validate regional optical retrievals over complex optical systems such as lakes, lagoons, estuaries and rivers.

The data presented here were gathered and processed druring three projects funded by the Belgian Science Policy Office (BELSPO) and one project funded by the Research Foundation – Flanders (FWO). The PONDER project (BELSPO SR/00/325) focused on developing tools for spaceborne remote sensing of inland water systems using high spatial resolution ($\leq$ 30 m) sensors, of which current global coverage and open access data is only available for multispectral missions. The HYPERMAQ project (BELSPO SR/00/335) focused on exploring hyperspectral data for detailed biogeochemical retrievals in support of the new generation of hyperspectral spaceborne remote sensing missions. The PHYTOBEL project (BELSPO SR/02/213) provided funding for data curation and publication. The Flemish part of the LifeWatch BE, a long-term research project funded by FWO, provided biogeochemical data and infrastructure for sampling the Belgian coastal zone (BCZ). Samples were taken from 2017 to 2019 and cover eight lakes, the Spuikom lagoon, the Scheldt estuary and the BCZ.

The goal of this data report is to provide a detailed description and validation of the methods used during the research, present a summary of the observations and briefly discuss aspects of the data that might be relevant for potential users.

2 Methods and data description

Measurements are presented in three groups: (1) Inherent optical properties (IOPs) consisting of absorption, scattering and attenuation coefficients, near-forward volume scattering function and turbidity; (2) Apparent optical properties (AOPs) consisting of Secchi disk depth, water-leaving and substrate Lambert-equivalent bi-hemispherical reflectance; and (3) biogeochemical properties consisting of suspended particulate matter concentration, mineral fraction of particle mass, particle size distribution, pigment concentration, DNA metabarcoding, flow imaging microscopy counts, and bottom type classification. Table 1 presents the description of relevant acronyms, symbols, constants and subscripts used in this study. The studied aquatic systems are listed in Table 2. The measured parameters and the number of samples are listed in Tables 3, 4 and 5. All coordinates are relative to the WGS84 datum and times are reported in UTC.

Linear models used for consistency check between parameters were fitted using robust linear regression. For selected parameters, the data summary is provided in the form of a violin plot, which in addition to the median point, 25th and 75th percentiles, presents the empirical distribution of the data as the variable width of the planar shape. All analyses were performed in R (version 3.6.3, R Core Team, 2020) with aid of packages 'MASS' (version 7.3-51.5, Venables and Ripley, 2002), 'vioplot' (version 0.3.6, Adler and Kelly, 2020), 'dada2' (version 1.12.1, Callahan et al., 2016) and 'decontam' (version 1.6.0, Davis et al., 2018).

2.1 Study sites

The majority of the measurements were performed in Belgian inland waters, in particular four lakes and a coastal lagoon. The sampling frequency along the growth season varied with the sampling year. The field campaigns were performed seasonally throughout 2017 (fortnightly) and 2018 (monthly), spanning a broad range of conditions. Data for four additional lakes with sporadic sampling during 2017 are also included in the dataset. Those systems were previously described in Castagna et al. (2020) and are reintroduced here for convenience. A summary of the study sites is presented in Table 2.

The Spuikom (51°13'41.0"N 2°57'09.5"E) is a shallow, brackish-marine coastal lagoon (typical salinity between 27 PSU and 33 PSU), with a surface area of 0.82 km^2 and an average depth of 1.5 m. It is connected to the sea through the Ostend harbor by a lock system. The lagoon exchanges water with the harbor continuously, resulting in median daily renewal of 3.7 % of its volume. It is used for recreational activities and shellfish aquaculture. During the observation period (2017 to 2018) it experienced a cycle of diatom blooms in spring followed by a transition to transparent waters in autumn, with extensive bottom coverage by macroalgae. It is subject to strong coastal winds that can cause sediment resuspension, adding bottom sediments and benthic diatoms to the water column. The chlorophyll a (Chl a) concentration typically varied from 2 mg m^{-3} to 25 mg m^{-3}, with the exception of an intense nanoflagellate bloom that reached a Chl a concentration of ≈130 mg m^{-3}. The Secchi disk depth ranged from 0.62 m to bottom.

The Hazewinkel (51°03'57.4"N 4°23'27.1"E) is a mesotrophic lake with an area of 0.66 km^2 and a maximum depth of 20 m. It is used for recreational and sport activities. During the observation period (2017) it presented a peak of phytoplankton abundance in spring, with Chl a concentration reaching 20 mg m^{-3}, after which the concentration stayed around 5 mg m^{-3}. Secchi disk depth varied from 1 m to 6 m. Submerged macrophytes were confined to near-shore locations due to a steep basin slope.

The Donkmeer (51°02'23.4"N 3°58'47.2"E) is the second largest lake in Flanders, with a surface area of 0.86 km^2 and an average depth of 2 m. It experiences recurrent cyanobacterial blooms of *Anabaena spp.* and *Planktothrix agardhii* (Descy et al., 2011). It is used for recreational activities and fishing. During the observation period (2017 to 2018) it experienced cyanobacterial blooms from summer to autumn, reaching a Chl a concentration of ≈400 mg m^{-3} and a Secchi disk depth of 0.2 m. Its northern and southern portions are connected through a narrow and shallow passage, with the northern portion experiencing a shorter period of cyanobacterial blooms due to management actions.

The Dikkebus (50°49'07.4"N 2°50'39.5"E; 0.36 km^2, 2.5 m deep) and the Zillebeke (50°50'15.4"N 2°54'34.3"E; 0.28 km^2, 2 m deep) are two lakes created in the 13th century for water supply, a function that remains to date along with recreational activities. Both lakes exhibit annual blooms of the cyanobacteria *Microcystis aeruginosa* and *Aphanizomenon flos-aquae* (Descy et al., 2011). During the observation period (2018), intense cyanobacterial blooms were observed with the Chl a concentration reaching 105 mg m^{-3} and in the Zillebeke those blooms were associated with fish mortality. The Secchi disk depth ranged from 0.5 m to 2 m. Additional stations were included from four other Flemish lakes: Bocht (51°04'24.0"N 4°23'19.8"E; 0.35 km^2 and 18 m deep), Nieuwdonk (51°02'04.1"N 3°58'32.2"E; 0.26 km^2 and 22 m deep) and two unnamed adjacent

85 lakes (51°06'06.1"N 4°01'44.1"E; ≈0.16 km² each and 10 m deep), here referred to as "DelMare", a sand extraction lake with water sports activities, and "Molsbroek", a nature reserve.

Other inland water systems included were the Scheldt estuary and the Leuven-Dijle canal. The Scheldt is a rain-fed low-land river, with an estuary environment subjected to tides from the mouth at Vlissingen (The Netherlands) to Ghent (Belgium, 160 km upstream), where a system of locks prevent further propagation of the tide (Meire et al., 2005). It has a large eco-

90 nomical importance as a transport waterway, connecting the harbours of Antwerp and Ghent to the North Sea. The Scheldt estuary was sampled in mid October 2019 at two locations near the city of Sint-Amands (51°03'18.0"N 4°11'59.6"E and 51°04'24.1"N, 4°11'24.0"E), with a time series including a full tidal cycle (tidal range of 6 m). During the observation period the Chl a concentration reached 55.7 $\mathrm{mg\,m^{-3}}$ at high tide, while the suspended particulate matter (SPM) concentration reached 791.2 $\mathrm{g\,m^{-3}}$. In the same campaign performed in the Scheldt, two samples were taken at the Leuven-Dijle canal, a

95 highly transparent artificial waterway running parallel to the Dijle river, connecting Leuven to the Zenne-Dijle confluence, and ultimately to the Scheldt. The sampling position was close to the lock of Zennegat (51°03'46.7"N 4°25'50.0"E).

The stations sampled in the BCZ are part of the regular LifeWatch sampling campaigns (Mortelmans et al., 2019). The April and July 2018 campaigns were augmented to include spectroscopic measurements. The BCZ is a shallow part of the North Sea (1 m to 40 m), experiencing high tidal fluctuations (average of 4 m) and strong tidal currents (1 $\mathrm{m\,s^{-1}}$). Those conditions,

100 combined with limited freshwater discharge into the region, result in a well mixed-water column (van Beusekom and Diel-Christiansen, 1993). It experiences high turbidity (SPM concentration from 1 $\mathrm{g\,m^{-3}}$ to 200 $\mathrm{g\,m^{-3}}$) with large influence from particulate material imported through the Strait of Dover (Fettweis and Van den Eynde, 2003). And develops a turbidity maximum zone near Zeebrugge, also influenced by the decreasing magnitude of the residual transport vectors from the East border (Fettweis and Van den Eynde, 2003). The inflow of the Yser and Scheldt rivers influence the availability of nutrients,

105 with an increase of nitrogen and phosphorus since the second half of the 20th century, followed by a de-eutrophication phase during which nitrogen and especially phosphorus decreased (Desmit et al., 2020). The phytoplankton seasonal dynamics are well described (*e.g.*, Reid et al., 1990), with an early spring diatom bloom followed by a mixed bloom of the haptophyte *Phaeocystis globosa* and diatoms. A recent review of the phytoplankton seasonal dynamic was provided by Castagna et al. (2021). The IOPs in this region were extensively studied by Astoreca et al. (2006, 2009, 2012).

110 ## 2.2 Sampling

Sampling was performed from a diverse set of platforms. For the inland water campaigns, samples were taken from pontoons or an inflatable boat, depending on the system and date. The water was sampled just below the surface, taking care not to draw in materials floating at the surface (*e.g.*, pollen, debris, etc). Field samples were stored in 5 L semi-transparent plastic carboys, and kept in the dark and cold during the transport to the laboratory. Sampling for the coastal campaigns was performed from

115 the Research Vessel (RV) Simon Stevin (Flanders Marine Institute, VLIZ). For most marine samples, water was sampled just below the surface using Niskin bottles (General Oceanics, Inc.), attached to a rosette system. The exception were subsurface samples for flow imaging microscopy, taken with a bucket from the side of the ship. Filtrations were performed on board and water subsamples were kept in the dark and cold during transport to the laboratory for spectrophotometric measurements.

Macrophytes, sediment and biofilm were sampled in the Spuikom lagoon. Macrophytes were sampled during 2017 and 2018, by collecting floating specimens or recovering specimens from the bottom using a rake. Specimens were stored in transparent plastic bags containing water from the lagoon, and kept in the dark and cold during transport to the laboratory. Sediments and biofilm were sampled in April 2018 using polymethyl methacrylate (PMMA) tubes attached to a short corer. The cores were retrieved with care not to disturb the surface of the sediment, sealed and transported to the laboratory in a vertical position. In July 2018, biofilm patches had detached from the bottom of the Spuikom and were sampled floating at the surface and stored in plastic bags for microscopic examination. The cores and macrophytes were stored in a climate room at 4°C for up to 3 days until analysis.

2.3 Inherent Optical Properties

Most of the IOP measurements were performed *ex situ*, with a benchtop spectrophotometer (Lambda 650S, PerkinElmer) equipped with a integrating sphere (150 mm internal diameter). The interior of the integrating sphere is made of highly reflective (nominal 99 %) sintered polytetrafluoroethylene (PTFE). The spectral IOP measurements made with the spectrophotometer include particle absorption (a_p), chemically decomposed into *in vivo* pigment absorption (a_ϕ) and depigmented particle absorption (a_d), chromophoric dissolved organic matter absorption (a_g), particle attenuation (c_p) and scattering (b_p) neperian coefficients. Methods followed recommendations from Pegau et al. (2002) and IOCCG (2018). Measurements covered the range from 250 nm to 850 nm in 1 nm steps, with a 2 nm integration slit and 0.24 s integration time (250 nm min^{-1}). Data is provided in the range of 380 nm to 850 nm, and include a single pass of a smoothing function to reduce noise (rectangular filter, 10 nm window moving average).

Turbidity was measured in discrete samples with a portable turbidimeter (2100P ISO, HACH). Additional IOP data were measured from *in situ* instrumentation for a subset of water systems and stations. These include the diffraction peak ($\psi < 15°$) of the particle volume scattering function (VSF; $\beta_p(670)$) and non-water attenuation at an acceptance angle of 0.018° measured at 670 nm ($c_{nw}(670, 0.018)$; LISST, Sequoia Scientific).

2.3.1 Absorption coefficient from dissolved components (< 0.45 μm)

Absorption due to chromophoric dissolved organic matter (CDOM) was determined in the laboratory from fresh and undiluted filtered subsamples, using a 5 cm pathlength quartz cuvette. Subsamples were filtered with 0.45 μm polyamide (nylon) fiber syringe filters. Between samples, the syringe and cuvette were rinsed with deionized water. The filters were first rinsed by filtrating 15 mL of deionized water before use with samples. The first sample filtration volume was used for final rinse of the quartz cuvette and the volume discarded. The cuvette was then gently filled with the second filtration volume to avoid the formation of bubbles, and allowed to rest for at least 3 min before measurements. The blank was determined with deionized water, using the same cuvette and at the same temperature (room temperature, ≈20°C). Quality control evaluated the presence of absorption peaks at 676 nm as an indication of cell leakage and the offset from zero absorption at 750 nm as a indication of the presence of hydrosols.

The use of deionized water as blank for all measurements resulted in a mismatch of salinity for marine and brackish samples. Dissolved salts change the complex refractive index of water (Quan and Fry, 1995; Röttgers et al., 2014b), affecting reflection and refraction interactions with the cuvette wall, water scattering and absorption. In the wavelength range reported here, the scattering effects and the absorption of salts are negligible, however the dissolved salts form what is known as salt-solvated

155   water, mixed with pure water (Max and Chapados, 2001). Salt-solvated water can have higher or lower absorption than pure water, depending on the wavelength, with a magnitude defined by the salt type (Max and Chapados, 2001; Röttgers et al., 2014b). This difference is particularly important in the NIR range, due to the spectral shape of a_g. The effect of the mismatch in salinity was investigated by comparing the effects of salinity on blank readings for a_g determinations from deionized water and artificial seawater at a salinity of 35 (NaCl at 35 $g\,kg^{-1}$). The results are shown in Fig. S1 (supplementary material) and

160   the resulting difference spectrum was subtracted from a_g of the Spuikom lagoon and the BCZ for an approximate correction. Dissolved molecular oxygen (O_2) also contributes to absorption in the UV range (Jonaz and Fournier, 2007, and references therein), however in the wavelength range reported here CDOM dominates the signal.

    The Lambda 650S was not available in 2017 and a_g was measured for a subset of samples using another spectrophotometer (UV-1601, Shimadzu). Sample preparation was the same, except that a_g was only measured at selected wavelengths (380 nm,

165   400 nm, 433 nm, 550 nm and 750 nm). To retrieve the full spectrum for these samples, an hyperbolic model was fitted to the available wavelengths (Twardowski et al., 2004). The hyperbolic model fit was also applied to all measurements in the range of 380 nm to 850 nm to provide a smooth, fitted version of a_g and to calculate the CDOM absorption spectral hyperbolic slope coefficient, S_{h_g}:

$$a_g(\lambda) = a_g(443)\left(\frac{\lambda}{443}\right)^{-S_{h_g}}. \tag{1}$$

170   Most samples had negligible absorption in the NIR ($< |0.1|$ $m^{-1}\,nm^{-1}$), fluctuating around zero due to instrument noise or residual blank salinity difference effect. Four stations had positive or negative offsets that were larger than $|0.1|$, and the negative offsets were attributed to the presence of bubbles in the blank of the day. All spectra were subtracted by the average a_g between 750 nm and 850 nm. Five freshwater samples had a pronounced Chl a absorption peak at 676 nm, and since NIR signals were close to zero, those peaks likely result from cell leakage. The a_g for those stations were flagged.

175   The magnitudes of $a_g(412)$ varied between 0.547 $m^{-1}\,nm^{-1}$ and 4.185 $m^{-1}\,nm^{-1}$, with values above 2.25 $m^{-1}\,nm^{-1}$ generally observed only in freshwater systems. The exception was a sample in the Spuikom collected during a nanoflagellate bloom, when $a_g(412)$ reached 3.160 $m^{-1}\,nm^{-1}$. The range of the S_{h_g} was between 5.60 and 8.81. The observed values of a_g and fitted S_{h_g} are presented in Fig. 1.

2.3.2   Absorption coefficient from particulate components (> 0.7 μm)

180 Particle absorption was measured using the filterpad method, with particles concentrated onto a glass fiber filter (GF/F, effective mesh of 0.7 μm). To improve the homogeneity of particle deposition over the filtration area, two stacked filters were used and samples with high abundance of particles were diluted in the filtration funnel with deionized water. Immediately after filtration,
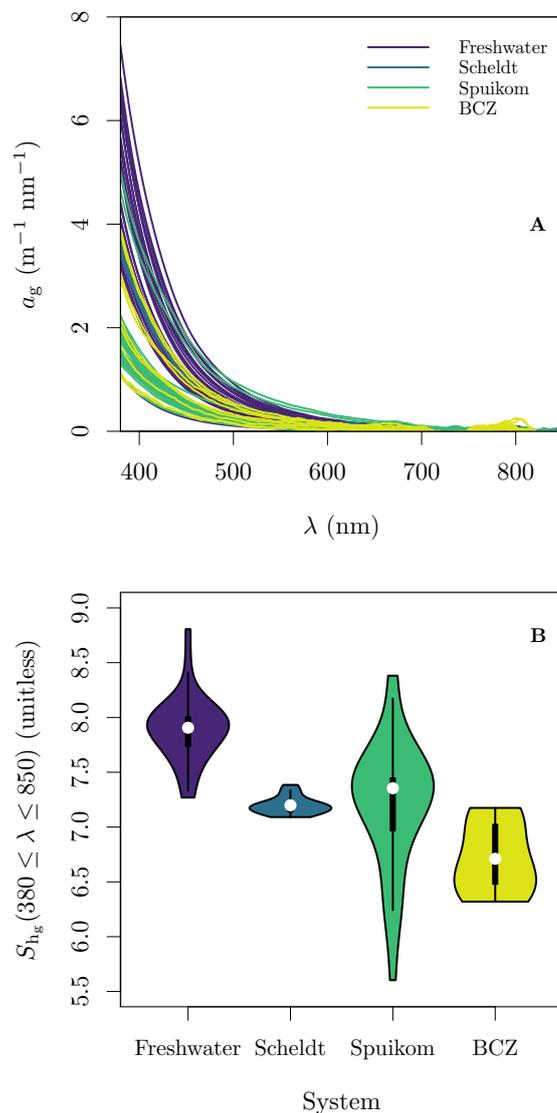
Figure 1. Chromophoric dissolved organic matter (CDOM) absorption in the Belgian water systems. (A) CDOM spectral absorption coefficient, a_g; and (B) hyperbolic slope of a_g, S_{hg}, fitted in the range of 380 nm to 850 nm.

filters were transferred to PetriSlides (Merck), wrapped in aluminum foil and frozen in liquid nitrogen. Filters were then stored at -80 °C until analysis.

185   Before analysis, the filters were allowed to thaw to room temperature and kept hydrated with deionized water. To avoid dislocating large particles deposited on the filter fibers, hydration was performed by raising the filter, adding a droplet of water to the PetriSlide base and gently lowering the filter onto the droplet, resulting in water spreading by capillarity. The "inside sphere" variant of the quantitative filterpad method was used (Stramski et al., 2015), using a center mount coated with PTFE.

Filters were read twice, with a 90° rotation between reads to average small deviations from homogeneous deposition. This was

190 especially important to account for large cyanobacteria colonies when these occurred in low densities (Fig. S2, supplementary material). Pigments were then oxidized by treating the filters with sodium hypochlorite (NaClO; Ferrari and Tassan, 1999), using the same approach as for hydration. Filters were read after 15 min or complete oxidation, following the same orientations as of the pigmented readings. The pathlength amplification correction was taken from Stramski et al. (2015) as recommended in IOCCG (2018). The *in vivo* pigment absorption coefficient (a_ϕ) was calculated as the total particulate absorption coefficient

195 (a_p) subtracted by the depigmented particle absorption coefficient (a_d). The chemical oxidation step was not performed for the Scheldt samples.

Sodium hypochlorite has an absorption peak at ≈300 nm, with residual absorption up to 500 nm. One method to remove the oxidant's signal is to rinse the filter with pure water before the determinations of a_d and this method was used for a subset of samples to evaluate the impact of sodium hypochlorite and rinsing. We observed that some samples had an apparent loss

200 of a_d observed between determinations with sodium hypochlorite and after rinsing (Fig. 2). This effect was dependent of lake, with Spuikom, Donkmeer, Hazewinkel and Bocht suffering little to no loss between 550 nm and 850 nm, while Dikkebus and Zillebeke showed large reductions in a_d after rinsing. This effect might provide additional information on the nature of the particles in a given system, but needs to be further explored to understand its causes. A direct consequence for measurements of a_d and a_ϕ is that rinsing might underestimate a_d.

205 Therefore we developed a statistical method to remove the oxidant's signal, instead of applying rinsing. We observed that when there was no loss of signal with rinsing, the minimum difference between a_p and rinsed a_d in the UV was at 305 nm, with $a_d(305)$ typically between 80 % and 90 % of $a_p(305)$ (Fig. 2A). Based on these observations, we fitted an exponential function to a_d with sodium hypochlorite, using the range of 550 nm to 850 nm, and including an estimate of rinsed $a_d(305)$ as $0.8a_p(305)$. The additional data point at 305 nm helps to set the curvature of the exponential model. Finally, an offset was

210 included as necessary to match the a_p absorption in the NIR end. The exponential model with an offset was:

$$a_d(\lambda) = \alpha_d + (a_d(550) - \alpha_d)e^{-S_{e_d}(\lambda-550)}, \tag{2}$$

where α_d (m^{-1} nm^{-1}) is a spectrally flat offset (Estapa et al., 2012) and S_{e_d} (nm^{-1}) is the spectral exponential slope of a_d.

The treatment with sodium hypochlorite can also introduce another artifact to the measurements. For a set of samples we observed a baseline offset between a_p and a_d in the NIR even before rinsing, propagating to increased baseline absorption of

215 a_ϕ. This likely results from sodium hypochlorite removing the adsorbed organic layer over particles (Binding et al., 2008), with the magnitude of the effect proportional to the concentration of particles and organic matter. In our samples this effect was larger in the maximum turbidity zone of the BCZ. This loss of absorbing material was not observed in a study by Röttgers et al. (2014a) including samples from a diverse set of environments, though the authors did not apply NaClO to the North Sea or Baltic Sea samples. The baseline effect was compensated for by adding the minimum offset necessary to match a_p and a_d in the

220 range of 800 nm to 850 nm without generating negative a_ϕ values. This constant offset likely underestimates the contribution of organic absorption in the blue end due to the exponential spectral shape of this component (*e.g.*, Cael and Boss, 2017).
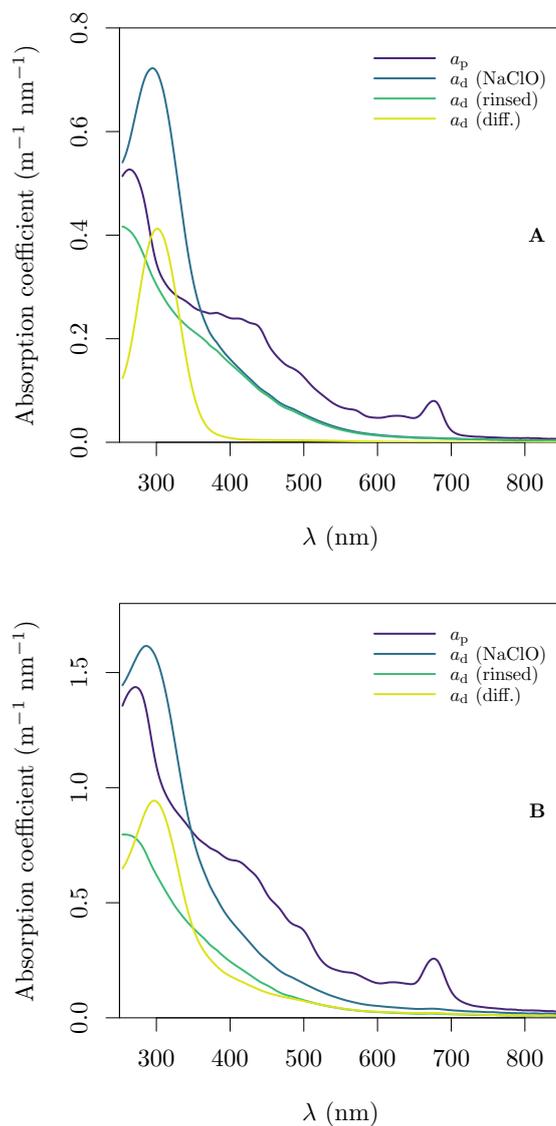
Figure 2. Evaluation of the treatment of the filters with sodium hypochlorite (NaClO) and subsequent rinse on the determinations of a_d. (A) A sample from the Bocht (BL_08) and (B) a sample from the Dikkebus (DK_01). The sample in (B) show loss of absorbing material after rinsing.

Similar to what was observed for $a_\mathrm{g}(412)$, the magnitudes of $a_\mathrm{d}(412)$ varied between $0.055\,\mathrm{m^{-1}\,nm^{-1}}$ and $4.570\,\mathrm{m^{-1}\,nm^{-1}}$, with values above $2.79\,\mathrm{m^{-1}\,nm^{-1}}$ generally observed only in freshwater systems. The baseline absorption coefficient in the NIR (a_p or a_d, and equal to a_α) ranged from $0.003\,\mathrm{m^{-1}\,nm^{-1}}$ to $7.933\,\mathrm{m^{-1}\,nm^{-1}}$, with only Scheldt samples presenting values higher than $0.565\,\mathrm{m^{-1}\,nm^{-1}}$ (Fig. 4A). The range of the $S_\mathrm{e_d}$ was between $0.0064\,\mathrm{nm^{-1}}$ and $0.0194\,\mathrm{nm^{-1}}$. The fitted

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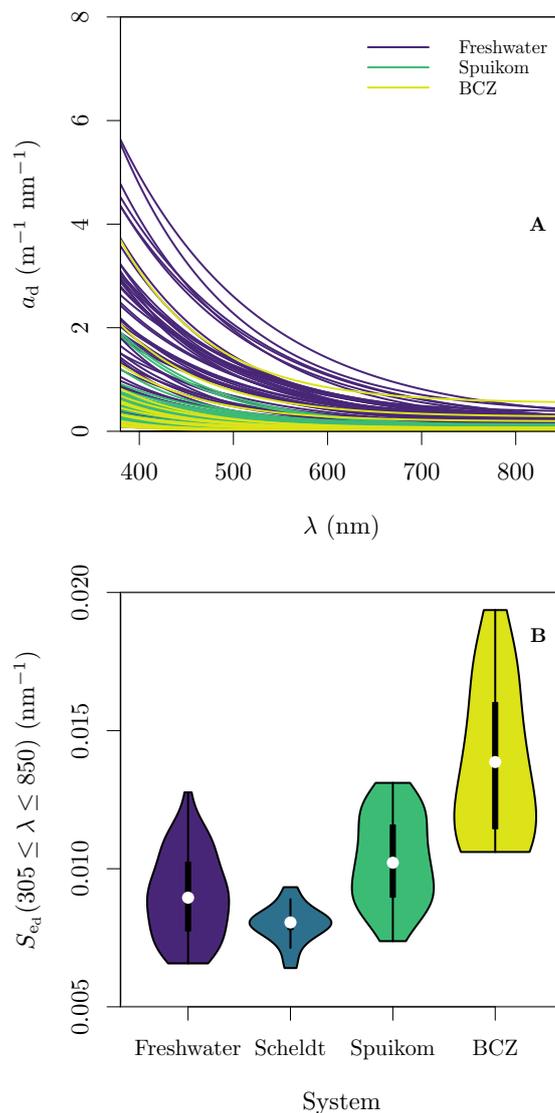
Figure 3. Depigmented particle absorption in the Belgian water systems. (A) Fitted depigmented particle absorption coefficient, a_d; and (B) spectral exponential slope of a_d, $S_{\mathrm{e_d}}$, fitted in the range of 380 nm to 850 nm. Note that for the Scheldt samples, the exponential model was fitted directly to a_p.

values of a_d and $S_{\mathrm{e_d}}$ are presented in Fig. 3. Since particle absorption samples from the Scheldt were not depigmented but were dominated by a_d (Fig. 4A), an estimate of $S_{\mathrm{e_d}}$ is provided by fitting Eq. 2 directly to a_p (Fig. 3B).

The integral-normalized a_ϕ presented a diversity of pigment absorption peaks (Fig. 4B), showing spectral shapes associated with dominance of cyanobacteria, green and red lineages of algae. Considering that a_ϕ was retrieved from fitted a_d, an indepen-

230   dent validation was performed against the total Chl a concentration (described later). In the blue end of the spectrum, we used
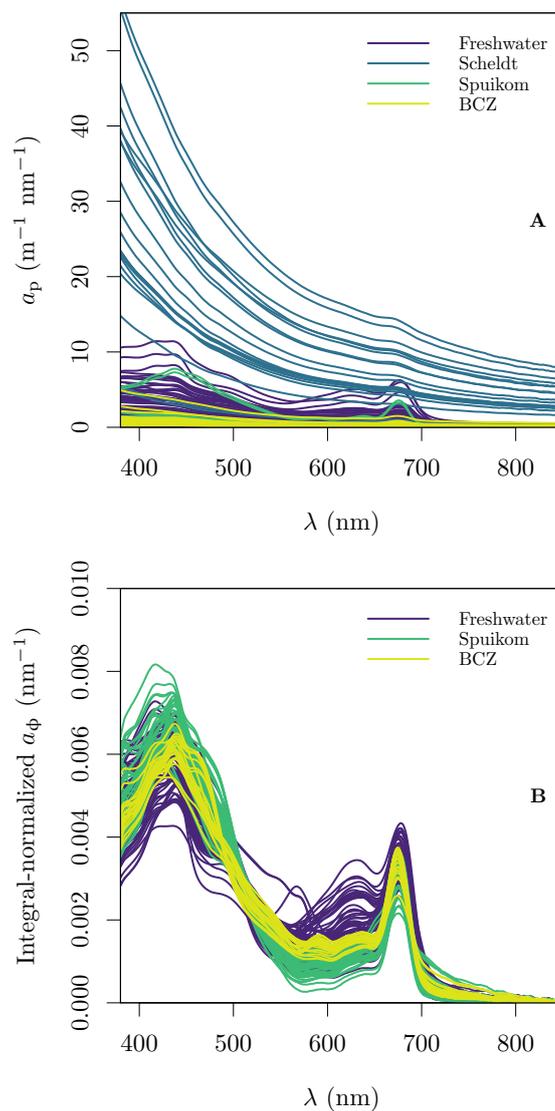
Figure 4. Particle and *in vivo* pigment absorption in the Belgian water systems. (A) Particle absorption coefficient, a_p; and (B) *In vivo* pigment absorption coefficient, a_ϕ. The a_ϕ presented here was calculated with the fitted a_d.

Gaussian decomposition (Hoepffner and Sathyendranath, 1991; Chase et al., 2013) to extract the magnitude associated with the Soret band of Chl *a in vivo* (435 nm; Fig. 5A). In the red end of the spectrum, we compared the $a_\phi(676)$ against models fitted to global datasets (Nardelli and Twardowski, 2016, Fig. 5B). We note that our observations are similar to those reported by Hoepffner and Sathyendranath (1991), while the value of Nardelli and Twardowski (2016) are lower likely due a combination of their lower Chl *a* concentration range and their estimation of $a_\phi(676)$ from the $a_{nw}(650, 676, 715)$ line height (Roesler and Barnard, 2013). The median spectral mass-specific *in vivo* pigment absorption coefficient at 676 nm, $^*a_\phi(676)$, was equal to

235

0.022 $\mathrm{m^2\,mg^{-1}\,nm^{-1}}$ (Fig. 5C), similar to the value of 0.020 $\mathrm{m^2\,mg^{-1}\,nm^{-1}}$ presented by Hoepffner and Sathyendranath (1991). The comparisons in the blue and red spectral regions we observe a linearity in log scale, with larger spread in the comparison $a_{\mathrm{Gauss}}(435)$ likely due to a combination of the uncertainty in the estimate of the fitted a_{d} and the variable pigment
240   packaging effect (Morel and Bricaud, 1981; Latimer, 1983), both larger in the blue spectral range.

2.3.3   Near-foward ($< 15°$) particle volume scattering function

The near-forward particle volume scattering function (VSF; β_{p}) at 670 nm was measured with two Laser In-Situ Scattering and Transmissometery (LISST, Sequoia Scientific Inc.) instruments, models 200X and 100X, equipped with a detector type C. The LISST-100X has a set of 32 concentric detectors measuring β_{p} between 0.038° and 7.519°, while the LISST-200X has
245   a set of 36 concentric detectors, the first 32 detectors measuring β_{p} between 0.081° and 15.394°. Single depth measurements were taken at 1 m depth, with the instrument deployed horizontally (LISST-200X) or vertically (LISST-100X), having the optical sensor end oriented down. The vertical deployment, preferentially made on the shady side of the platform, was chosen to reduce the interference of environmental light since the model 100X does not compensate for external light. Considering those deployment conditions and that our systems were mostly turbid, we expect negligible influence of environmental light
250   on the measurements. The instruments were used in a subset of stations. Calibrations with deionized water were performed monthly and the most recent calibration used for blank values. The VSF was retrieved from the raw LISST data file, using the procedures described in Agrawal (2005) and the instrument calibration files.

Quality control was performed by flagging entries in which the single scattering transmittance (T_{b}) was lower than 30 %, to avoid artifacts produced by multiple scattering:

255    $$T_{\mathrm{b}}(670) = e^{-b_{\mathrm{t}}(670,0.018)l_{\mathrm{LISST}}},\tag{3}$$

where b_{t} is the total scattering coefficient and l_{LISST} is the pathlength of the LISST instrument. The non-water beam attenuation coefficient at 670 nm measured by the LISST transmittance sensor with an acceptance angle of 0.018°, $c_{\mathrm{nw}}(670,0.018)$, was subtracted by the $a_{\mathrm{nw}}(670)$ measured with a benchtop spectrophotomenter to calculate $b_{\mathrm{t}}(670,0.018)$. The VSF is an important parameter for a tentative correction of the spectral particle attenuation and scattering coefficients measured with the benchtop
260   spectrophotomenter.

2.3.4   Attenuation and scattering from particulate components (> 0.45 μm)

In addition to the $c_{\mathrm{nw}}(670,0.018)$ measured by the LISST instruments, the spectral particle attenuation (c_{p}) and scattering (b_{p}) coefficients were calculated from the c_{nw} measured on fresh samples with the Lambda 650S spectrophotometer, by subtracting a_{g} and $a_{\mathrm{g}}+a_{\mathrm{p}}$, respectively. To reduce the acceptance angle of the spectrophotometer (Boss et al., 2009; Leymarie et al., 2010),
265   a black barrier with a central circular aperture of 2.4 mm diameter was placed in the entry port of the integrating sphere. With a distance of 69.5 cm from the center of the sample cell to the integrating sphere (due to system of mirrors extending the pathlength), the detector acceptance angle in water for this configuration is 0.074°. The integration time of the instrument was
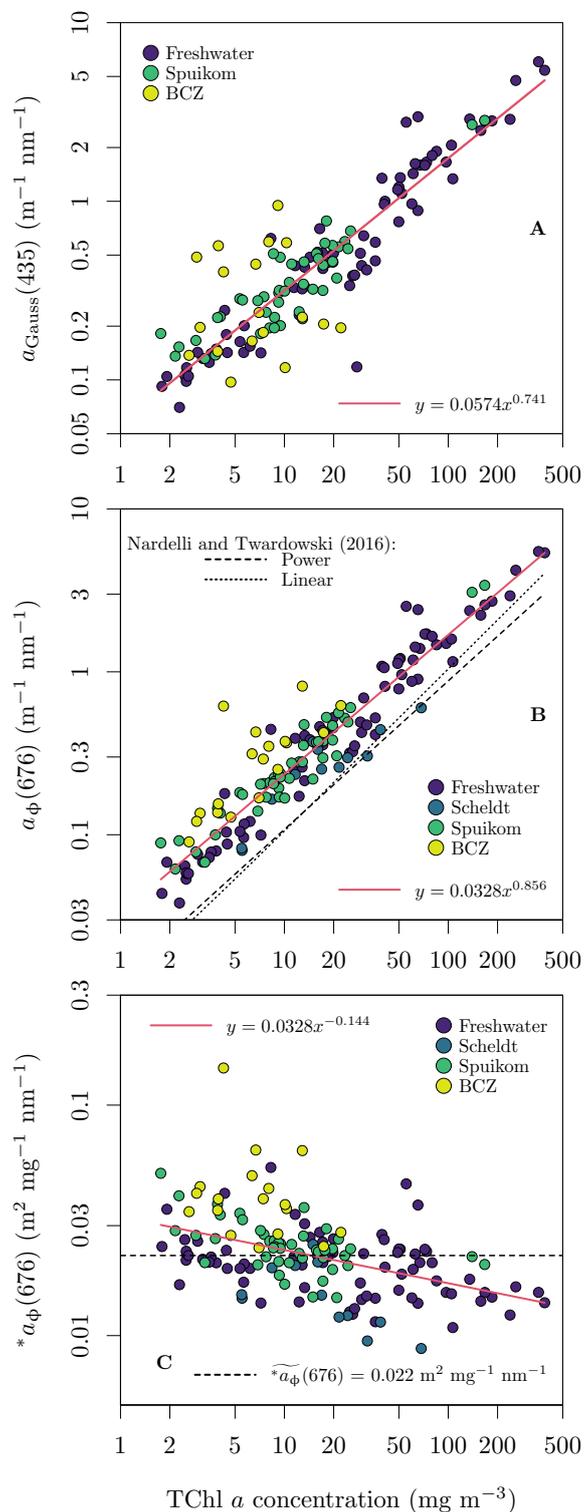
Figure 5. Relation between the total chlorophyll a (monovinyl chlorophyll a + chlorophyllide a) and *in vivo* pigment absorption. (A) Magnitude of the Gaussian peak centered at 435 nm: (B) the *in vivo* pigment absorption at 676 nm, $a_\phi(676)$; and (C) mass-specific *in vivo* pigment absorption at 676 nm, $^*a_\phi(676)$. The $a_\phi(676)$ for the Scheldt samples were estimated as the $a_{\rm nw}(650, 676, 715)$ line height (Roesler and Barnard, 2013).

increased to 0.4 s (150 nm min^{-1}) to compensate for the reduced signal, but noise levels were still noticed throughout the spectra. On later samples, the reference beam was partially blocked with a 1 % transmittance filter to reduce the dynamic range

270 imposed by the lack of a similar aperture in the reference beam.

As with the a_{g}, c_{nw} was measured in undiluted water subsamples, using 1 cm, 5 cm or 10 cm quartz cuvettes, depending on the turbidity. Blank readings were measured with deionized water, resulting in mismatched salinity for brackish and marine samples. The effect of the mismatched salinity was evaluated by comparing the attenuation between deionized water and artificial seawater at a salinity of 35 (NaCl at 35 g L^{-1}) in the same experimental conditions from sample measurements. In the

275 range from 380 nm to 850 nm, the difference in beam attenuation coefficient was ≈0.12 m^{-1} nm^{-1} (Fig. S3, supplementary material). The difference spectrum was subtracted from all brackish and marine samples for an approximate correction.

The measurements of $c_{\mathrm{p}}(670, 0.074)$ made with the benchtop spectrophotometer were tested against the LISST determinations of $c_{\mathrm{p}}(670, 0.018)$, with a similar method as described in Boss et al. (2009). Measurements of c_{nw} from both instruments were converted to c_{p} by removing the absorption by CDOM at 670 nm. The difference between $c_{\mathrm{p}}(670)$ estimated by each

280 instrument should be accounted for by the additional scattering signal captured by the larger acceptance angle of the spectrophotometer, given by the integration of the VSF between 0.018° and 0.074°. The VSF between the lowest measured angle by the LISST (0.038° or 0.081°) and its transmittance acceptance angle (0.018°) was calculated with linear extrapolation in $\log_{10}$ space from the two lowest measured angles. A detailed demonstration for sample DN_29 is presented in Fig. S4 (supplementary material). Fig. 6A shows the result for all samples with paired data available, with data closely following the 1:1

285 line, and variation possibly caused by artifacts related to *in situ* versus *ex situ* measurements. This analysis validates the c_{p} measurements with the spectrophotometer.

The additive error due the contamination with the near-forward scattered signal within the acceptance angle can be accurately corrected for only when the VSF of each sample is known. However, an approximate correction is possible. Fig. 6B shows that the scattering signal in the range of 0.018° to 0.074° represents an average of 43.2 % of the particle scattering coefficient, with

290 a stable relation across systems and seasons. Assuming that the angular shape of the VSF presents minor spectral variability and that the VSFs of different systems and seasons in our dataset are well represented by the subset for which LISST data is available, it is possible to use the approximate correction:

$$b_{\mathrm{p}}(\lambda, 0.074) = c_{\mathrm{p}}(\lambda, 0.074) - a_{\mathrm{p}}(\lambda), \tag{4}$$

295 $$b_{\mathrm{p}}(\lambda, 0.018) = \frac{b_{\mathrm{p}}(\lambda, 0.074)}{1 - 0.432}, \tag{5}$$

$$c_{\mathrm{p}}(\lambda, 0.018) = a_{\mathrm{p}}(\lambda) + b_{\mathrm{p}}(\lambda, 0.018). \tag{6}$$

The LISST-equivalent (acceptance angle of 0.018°) version of the spectral b_{p} and c_{p} are also provided in conjunction with the measured values (acceptance angle of 0.074°), as recommended by IOCCG (2018).
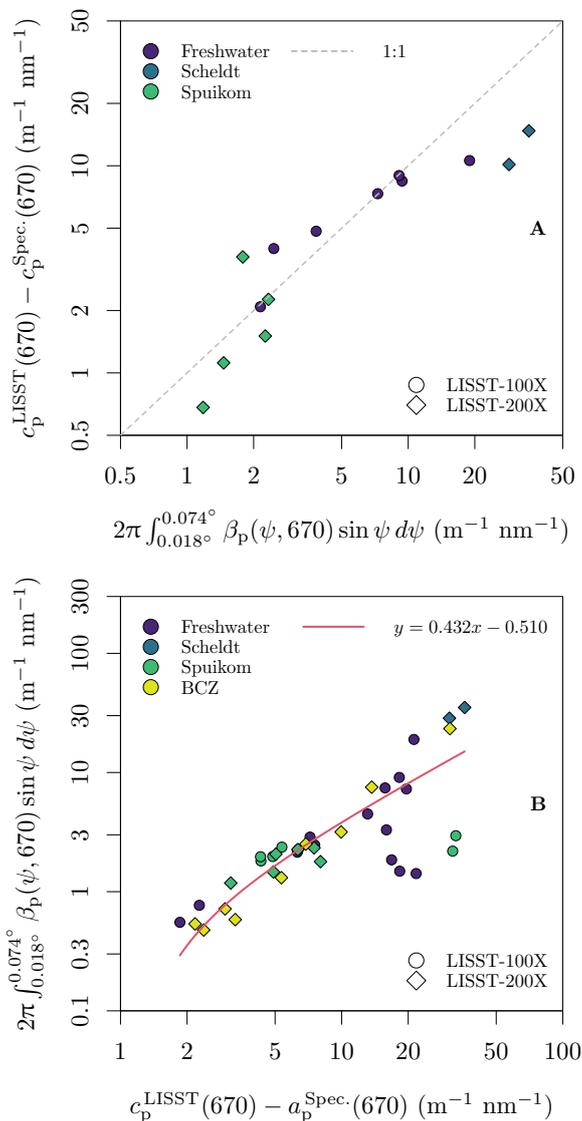
Figure 6. Evaluation of c_p measurement bias due to the finite acceptance of the spectrophotometer. (A) Relation between the difference of the c_p measured with the spectrophotometer and the LISST, and the VSF integral in the range of solid angles defined by the difference of their acceptance angles. (B) Relation between the scattering coefficient ($b_p = c_p - a_p$) and the magnitude of scattering between $0.018°$ and $0.074°$ scattering angles (ψ).

300   Quality control included the flagging of c_p spectra showing irregular behavior. Particle attenuation is a smooth spectral function (Roesler and Boss, 2003) due to anomalous diffraction that cause a complementary pattern between scattering and absorption (Zaneveld and Kitchen, 1995). Disturbances can be present in the form of small peaks caused by pigment absorption or oscillations caused by anomalous dispersion when there is a large contribution of small particles to the attenuation, but their

shape and position are characteristic. Large, abrupt and otherwise irregular spectral behavior likely arises from motion of large
particles in the beam cross sectional area. Six samples from Belgian lakes were flagged by this procedure, all with presence of
large cyanobacteria aggregates. All BCZ samples of April, taken during a major *Phaeocystis* bloom, were also discarded due
to similar effects of the large colonies. Beam attenuation remains elusive for standard bentchtop spectrophotometric analysis
(sequential spectral scanning) under these conditions. As an example, cyanobacteria aggregates were observed floating on the
surface of the cuvette at the end of the spectral run, even though the samples were mixed at the start of the measurement.

Another source of bias related to attenuation measurements in a spectrophotometer is that total internal reflection will occur
for a fraction of the scattered light, dependent on the incident angles on the water side of the quartz wall. Some fraction
of the internally reflected light could scatter back into directions in the detector's acceptance angle and artificially decrease
attenuation measurements (IOCCG, 2018). Our experimental procedure did not include a dark baffle inside the cuvette to
reduce this potential effect. Finally, our measurements show an oscillation centered at ≈500 nm, regardless of the system,
particle type or concentration. This suggests that this oscillation is an artifact of the measurement procedure, though we could
not identify its source.

2.3.5 Turbidity

Turbidity was quantified as formazin nephelometric units (FNU), a NIR side-scattering (90°) VSF magnitude relative to
formazin standards. For the BCZ and Scheldt campaigns, turbidity was measured just after water sampling, while for the lakes
and lagoon the turbidity was measured after transportation to the laboratory. The transportation might result in changes in
particle aggregation due to changes in turbulence. However, we found a single linear relation between turbidity and suspended
particulate matter (described later) across all systems and seasons (Fig. 11).

2.4 Apparent Optical Properties

The AOP measured *in situ* were the Secchi disk depth (d_{Secchi}) and the Lambert-equivalent water-leaving bi-hemispherical
reflectance ($\rho_{\mathrm{wl}}^{\mathrm{L}}$). $\rho_{\mathrm{wl}}^{\mathrm{L}}$ was measured with the above water or on-water protocols, depending on the system. The Lambert-
equivalent bi-hemispherical reflectance ($\rho_{\mathrm{s}}^{\mathrm{L}}$) measurements of substrate samples (sediments, macroalgae) were performed *ex
situ*, underwater and in air, using natural illumination.

The "Lambert-equivalent" qualification indicates that the bi-directional reflectance distribution function (BRDF) of the tar-
gets is assumed to be well represented by the Lambert model, and the hemispherical-directional measurement is converted to
bi-hemispherical by scaling it with the cosine-weighted solid angle integral of an hemisphere, π sr. The water-leaving signal
is not strictly Lambertian, however this approximation is commonly used for remote sensing purposes (*cf.* Frouin et al., 2019).

2.4.1 Secchi disk depth

A standard quadrant Secchi disk (black and white, 30 cm diameter) was used to measure transparency. The Secchi disk was
deployed from the shady side of the sampling platform, recording the depths of disappearence and reappearance. The Secchi
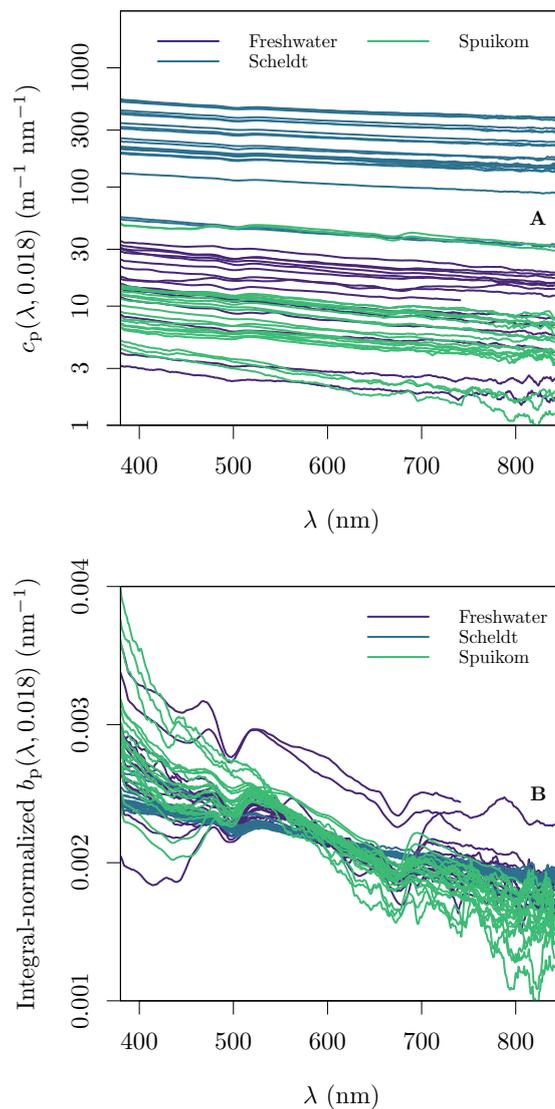
Figure 7. LISST-equivalent (acceptance angle of $0.018°$) spectral particle beam attenuation coefficient, c_p (A), and integral-normalized particle scattering coefficient, b_p (B).

335   disck depth (d_{Secchi}) was recorded as the average of the two depths. A tentative correction for the effect of sun zenith angle (described later) at the time of measurement (Lee et al., 2015) was applied following the formulation of Verschuur (1997). This correction normalizes the Secchi disk depth measurements to the Sun in the zenith. Measured and corrected values are provided.

2.4.2 Lambert-equivalent water-leaving bi-hemispherical reflectance

340 Reflectance spectroscopy measurements were performed in Belgian lakes and the coastal lagoon using the "on-water" method, also known as the skylight-blocked approach (Lee et al., 2013, 2019; Ruddick et al., 2019b). Measurements were made with a hand held spectrometer (FieldSpec HH, Analytical Spectral Devices) equipped with a 7.5° Field-of-View (FOV) foreoptics. The instrument has bands with 3.6 nm full width at half maximum (FWHM) and a spectral sampling of 1.6 nm, covering the range from 325 nm to 1075 nm. Water-leaving radiance (L_{wl}) was recorded at 0.5 m horizontal distance from the deployment

345 platform, aligned with the Sun azimuth, and with the opening of the lens' cylindrical shield at 2.5 cm below the water surface. The global downwelling plane irradiance just above the surface ($E_{dn}(0^+)$) was estimated from near-coincident measurements of the exitant radiance of a sintered PTFE reference target (nominal reflectivity of 12 %), held parallel to the surface (Castagna et al., 2019; Ruddick et al., 2019a). An example of the measurement approach is presented in Fig. S5 (supplementary material). The Lambert-equivalent water-leaving bi-hemispherical reflectance (ρ_{wl}^L) was estimated according to:

350
$$\rho_{wl}^L(\theta, \Delta\phi, \lambda) = \pi \frac{L_{wl}^{Meas}(\theta, \Delta\phi, \lambda)}{(1-\epsilon)E_{dn}(\lambda)}, \tag{7}$$

where ϵ is the estimated fractional shadowing error from the instrument and platform, θ is the nadir angle (0°) and $\Delta\phi$ here refers to the platform-sensor system relative azimuth to the Sun (0°). The superscript "Meas" for L_{wl} in Eq. 7 refers to the measured water-leaving signal, biased due to shadowing of the water system. Measurements were resampled to a regular 1 nm interval before Eq. 7 was applied. The shadowing error ϵ was calculated for the range of IOPs observed in the lakes and

355 lagoon and for the deployment from the inflatable boat and sensor as used in the campaigns, using a backward Monte Carlo radiative transfer code (unpublished). Formally, ϵ is a function of the deployment setup, Sun zenith angle, diffuse fraction of E_{dn}, a_t and c_t. However, the main IOP defining ϵ is a_t (Fig. S6, supplementary material). The correction was applied using field observations of the required parameters. For stations without measurements of IOPs, spectral c_p was estimated from a multivariate linear regression against turbidity calculated over all available data and a_g was estimated as the median a_g of given

360 water system. The diffuse fraction of E_{dn} was simulated from the Sun zenith angle for clear skies (Castagna et al., 2019) and was set to 1 when field observations recorded clouds covering the direct Sun illumination.

The same spectrometer used in the lakes and lagoon campaigns was used in the Scheldt campaign, with the above water approach (Mobley, 1999; Ruddick et al., 2019b, a). For this measurement, $\Delta\phi$ refers to the view direction relative to the Sun, set at 135°. The water-system radiance (L_{ws}) was recorded at a nadir angle of 40°, the sky radiance (L_{sky}) was measured at the

365 nominal specular angle from L_{ws}, 140°, and E_{dn} estimated from the exitant radiance of the reference sintered PTFE plaque. ρ_{wl}^L was estimated according to:

$$\rho_{wl}^L(\theta, \Delta\phi, \lambda) = \pi \frac{L_{ws}(\theta, \Delta\phi, \lambda) - \tilde{\rho}_f L_{sky}(180° - \theta, \Delta\phi, \lambda)}{E_{dn}(\lambda)}, \tag{8}$$

where $\tilde{\rho}_f$ is the effective Fresnel reflectance.

For the BCZ campaigns of 2018, reflectance spectroscopy measurements were also made with the above water approach,
370 using a set of three spectroradiometers (VIS-ARC RAMSES, TriOS) fixed on the bow of the RV Simon Stevin (Castagna et al.,
2021). The effective Fresnel reflectance was estimated from wind speed and L_{sky} according to Ruddick et al. (2006). Further
details of processing and quality control are described in Ruddick et al. (2006). The RAMSES instruments have a typical
bandwidth of 10 nm FWHM with spectral sampling every ≈ 3 nm. Radiance and irradiance measurements were resampled to a
regular 2.5 nm interval before Eq. 8 was applied, and $\rho_{\text{wl}}^{\text{L}}$ was interpolated to 1 nm interval to match the other reflectance data.
375 The above water measurements were not corrected for possible disturbances by the platform in the spectroscopic measurements
(Shang et al., 2020) and were not corrected for the non-nadir viewing geometry (Gleason et al., 2012).

The available $\rho_{\text{wl}}^{\text{L}}(0°)$ and $\rho_{\text{wl}}^{\text{L}}(40°, 135°)$ are presented in Fig. 8. As expected from the wide range of SPM concentrations,
$\rho_{\text{wl}}^{\text{L}}$ at 810 nm varied over three orders of magnitude, from 0.00036 to 0.10560. Similarly, the large diversity in terms of particle
composition (*e.g.*, mineral fraction, taxonomy, pigment) and relative contribution of a_ϕ translate to a diversity of $\rho_{\text{wl}}^{\text{L}}$ spectral
380 shapes. A validation of $\rho_{\text{wl}}^{\text{L}}$ and the shadowing correction for the on water approach is provided in Fig. 9, by estimating turbidity
from $\rho_{\text{wl}}^{\text{L}}(730)$ following the algorithm proposed by Nechad et al. (2010). The Nechad et al. (2010) algorithm was calibrated
with data between ≈ 1 g m^{-3} and ≈ 100 g m^{-3} and the comparison is restricted to that range, covering all systems with the
exception of the Scheldt.

2.4.3 Lambert-equivalent bi-hemispherical reflectance of bottom substrates

385 Measurements of reflectance spectroscopy of the surface of the sediment cores were made with the portable spectrometer
described previously (FieldSpec HH, Analytical Spectral Devices). The PMMA tubes were cut 5 cm above the sediment's
surface level, and measurements made in nadir view, with 2.5 cm of water above the sediment. Measurements were performed
with the on-water approach as described previously, *i.e.* an extension to the shield of the foreoptics was submerged to 2.5 cm
below the water surface. A circular NIST-traceable Munsel card was used as a submerged reference, gently placed over the
390 sediment. An example of the measurement setup is shown in Fig. S7 (supplementary material). The bottom depth at the
sampled stations varied between 1.3 m to 1.93 m and all but the station SP_43 (1.3 m deep) had the Secchi disk depth above
the bottom (average Secchi disk depth of 1.4 m). In July 2018, reflectance spectroscopy measurements were also made of
floating biofilm mats that had detached from the bottom of the Spuikom, presumably due to enhanced buoyancy caused by
bubbles trapped in the mucilage matrix following several days of high irradiance and heat. Measurements were performed
395 from a boat, without disturbing the floating biofilm mats. A sample taken for microscopy examination, revealed an assemblage
composed of representatives of the benthic diatom genera *Pleurosigma*, *Gyrosigma* and *Navicula* (Fig. S8, supplementary
material).

Reflectance spectroscopy measurements of the most conspicuous macrophyte species occurring in the Spuikom (*Cladophora
glomerata*, *Ulva sp.* and *Sargassum muticum*) were performed in air, with an hyperspectral camera (SOC710-VP, Surface
400 Optics Coorporation). The macrophyte reflectance measurements were made in nadir view with natural illumination. The
hyperspectral camera was set up at 1 m above the samples, using a f/2.8 aperture following the observation of lower spatial
variability in the visible to red-edge spectral region. The 12 % (nominal) reflectivity sintered PTFE plaque was used to estimate
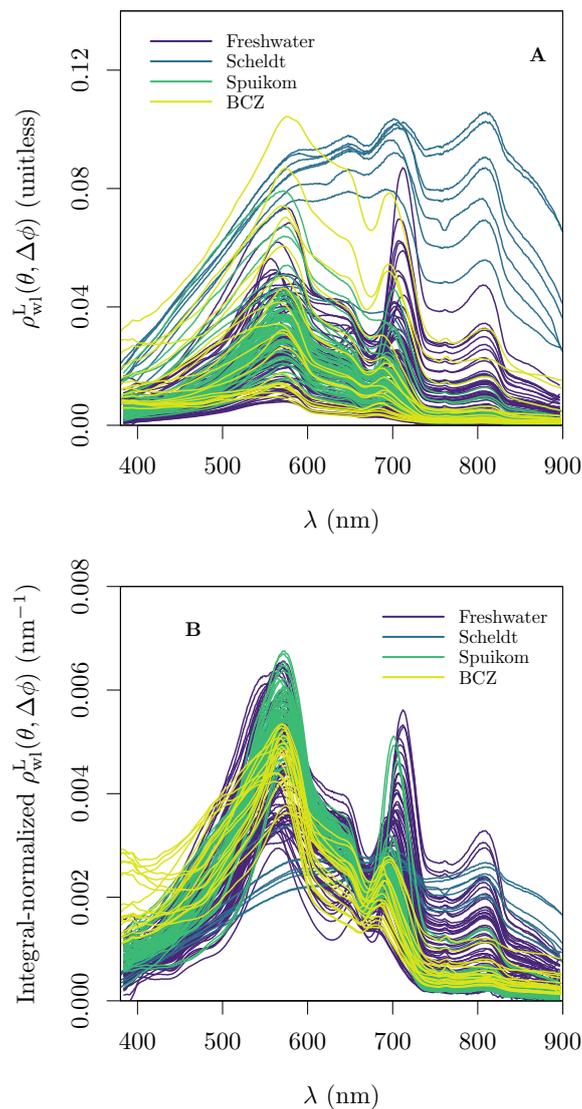
Figure 8. Lambert-equivalent water-leaving bi-hemispherical reflectance, ρ_{wl}^L, estimated on Belgian water systems. The view geometry is dependent on the measurement approach, being nadir view ($\theta = 0°$) for lakes and the Spuikom, and $\theta = 40°$ $\Delta\phi = 135°$ for the Scheldt and the BCZ. (A) Magnitude of ρ_{wl}^L and (B) its integral-normalization with respect to wavelength. $\rho_{wl}^L(0°)$ values measured with the on water approach presented here were corrected for shadowing.

E_{dn} and a spectrally flat 5 % reflectivity sheet was used as background. Specimens were folded over a supporting petridish to avoid signal from the background and the reflectance averaged over circular areas to average over the three dimensional

405 structures (Fig. S9, supplementary material). As the macroalgae were washed free of the sediment before the measurements, the determined ρ_s^L represent pure end members of substrate reflectance.
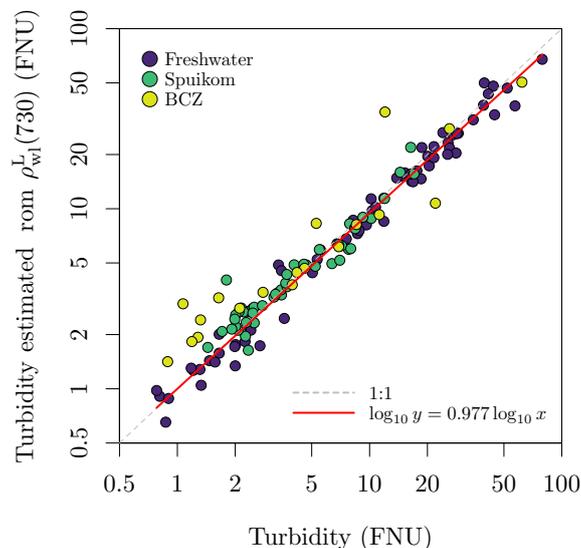
Figure 9. Assessment of ρ_{wl}^{L} data quality by comparing the turbidity estimated from reflectance using the algorithm of Nechad et al. (2010) to the measured turbidity.

The average ρ_{s}^{L} spectra of macroalgae and the spectra of sediment samples are presented in Fig. 10. ρ_{s}^{L} spectra of macroalage reached values as high as 0.4 in the NIR range, while sediments, independent of biofilm thickness, had an average ρ_{s}^{L} in the NIR of ≈ 0.11. In the visible range, the spectral shape of *Sargassum muticum* seems indistinguishable from the sediments with thickest biofilms, as expected due to similar pigment composition of brown algae and diatoms. The similarity in pigment composition also explains the similarity between the average ρ_{s}^{L} of healthy specimens of the green algae *Ulva sp.* and *Cladophora glomerata* (*cf.* Kotta et al., 2014).

2.5 Biogeochemical data

2.5.1 Suspended particulate matter and mineral fraction

Determinations of suspended particulate matter (SPM) concentration $(\mathrm{g\,m^{-3}})$ were performed gravimetrically, after filtration of the suspensions with pre-treated glass fiber filters (GF/F, Whatman, nominal mesh size of $0.7\,\mathrm{\mu m}$). For brackish and marine samples, the filters were rinsed with distilled water to remove salt from the filtration area and rim of the filter (Strickland and Parsons, 1968). The filters were then dried overnight at $60°\mathrm{C}$ and cooled to room temperature before mass determinations. The pre-treatment of filters involved combustion for one hour at $450°\mathrm{C}$ to eliminate organic components, followed by washing to remove loose glass fibers and blank mass determinations. To calculate the mineral fraction (f_{m}) of the SPM, the filters were heated to $500°\mathrm{C}$ for one hour for thermal oxidation of organic matter and cooled to room temperature before new mass determinations. For some filters, the last mass determinations were lower than the blank mass, indicating a combination of
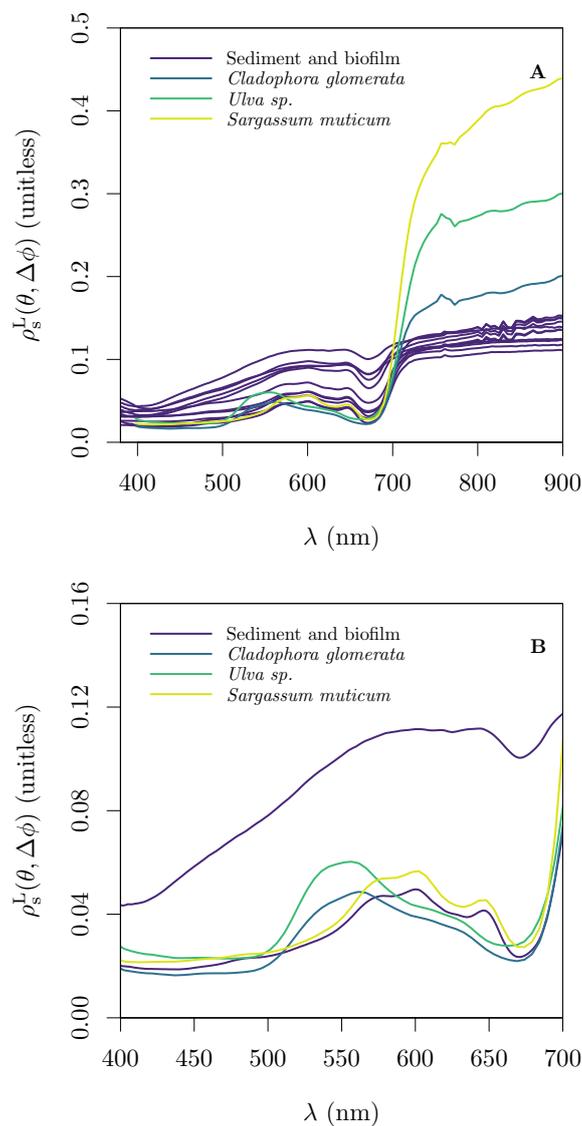
Figure 10. Lambert-equivalent substrate bi-hemispherical reflectance, ρ_s^L, estimated from samples on the Spuikom. The view geometry was nadir view ($\theta = 0°$, $\Delta\phi$ undefined) for all measurements. (A) The magnitude of ρ_s^L in the visible to NIR spectral range and (B) the magnitude in the visible spectral range. In (B) only the sediment samples with the thinnest and thickest biofilm mats are shown, as judged by the reflectance magnitude at 676 nm.

very low mineral fraction and loss of glass fibers during manipulation. Those samples had the mineral fraction set to zero. The observed range of SPM across all systems was from 1.02 ± 0.09 g m^{-3} to 791.19 ± 0.10 g m^{-3} and the range of mineral fraction was from 0 ± 0.00 to 0.95 ± 0.08. A comparison between SPM and turbidity is presented in Fig. 11.
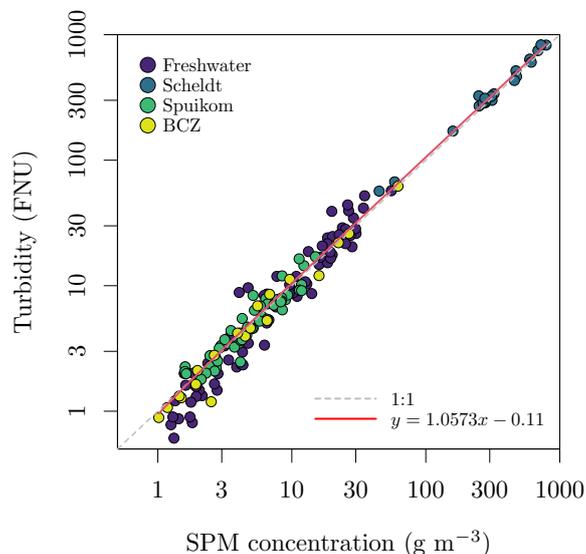
Figure 11. Relation between suspended particulate matter (SPM) concentration and turbidity in Belgian water systems.

2.5.2 Particle size distribution

The particle size distribution (PSD) was inverted from the LISST measurements of VSF, using the random shaped particle VSF kernels provided by the manufacturer. The LISST-100X with random shaped particle inversion provides the PSD between 1.8 μm and 381 μm in 32 log-spaced intervals, while the LISST-200X provides the PSD between 1 μm and 500 μm in 36 log-spaced intervals. The LISST software automatically flags measurements with transmittance lower than 30 %. For further details of the measurements with the LISST, see Sec. 2.3.3. Values are provided as volume concentration PSD ($cm^3\,m^{-3}$, ppm). The PSDs converted to particle number concentration, using the volume of a sphere with the median diameter of each retrieved bin size range (Buonassissi and Dierssen, 2010), are shown in Fig. 12. Assessing differences in the PSD measured with the two instruments is under further investigation, but beyond the scope of this data report.

2.5.3 Pigment concentration

Pigment mass concentrations were determined using High Performance Liquid Chromatography (HPLC), following the method of Van Heukelem and Thomas (2001). Cells were broken with sonication and the suspension was cleared by filtration through a 0.22 μm syringe filter. The HPLC was equipped with a reverse-phase column (Eclipse XDB C_8) and the detection was performed with spectral absorption (Agilent 1100 series, Diode Array Detector). Pigment standards were acquired form the Danish Hydrographic Institute (DHI) and quantified pigments are listed in Table 6. For the BCZ samples, the measurements are part of the regular marine LifeWatch BE sampling campaigns (Flanders Marine Institute, 2021a), described in Mortelmans
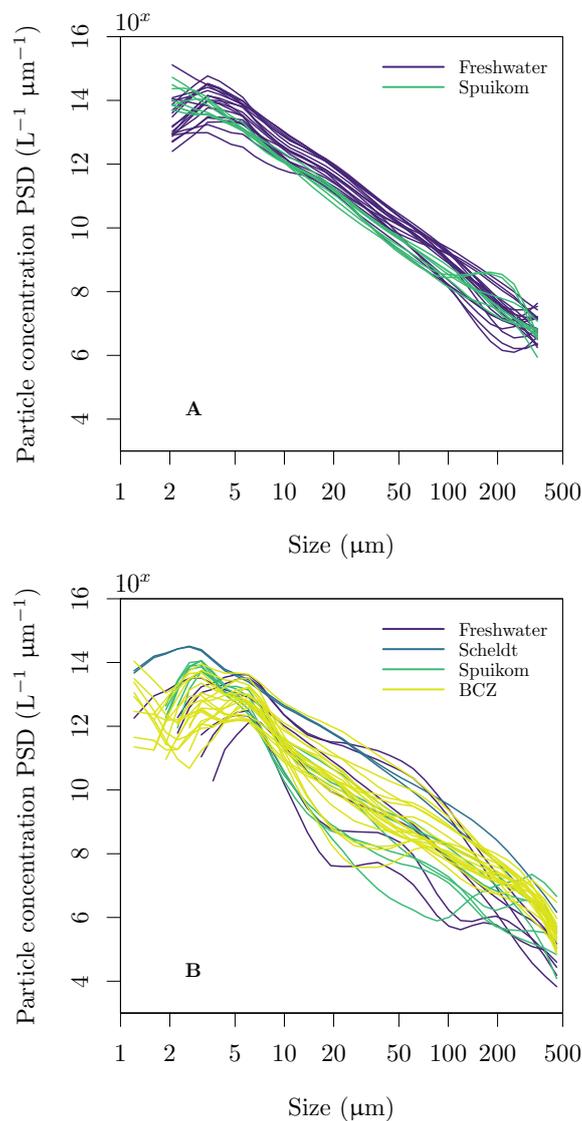
Figure 12. Particle size distribution (PSD) in the Belgian water systems. (A) Retrieved with the LISST-100X; and (B) retrieved with the LISST-200X. The continuous particle number concentration PSD was calculated from the volume concentration PSD assuming spherical particles of median diameter in each size bin and normalizing by the width of each logarithmically spaced size bin.

et al. (2019). The observed range of total Chl a (Chl a + Chlide a) across all systems was from 0.63 mg m^{-3} to 387.53 mg m^{-3}. Fig. 13 presents the distributions of total Chl a per system and the spread of Chl b to total Chl c (Chl c_1c_2 + Chl c_3).
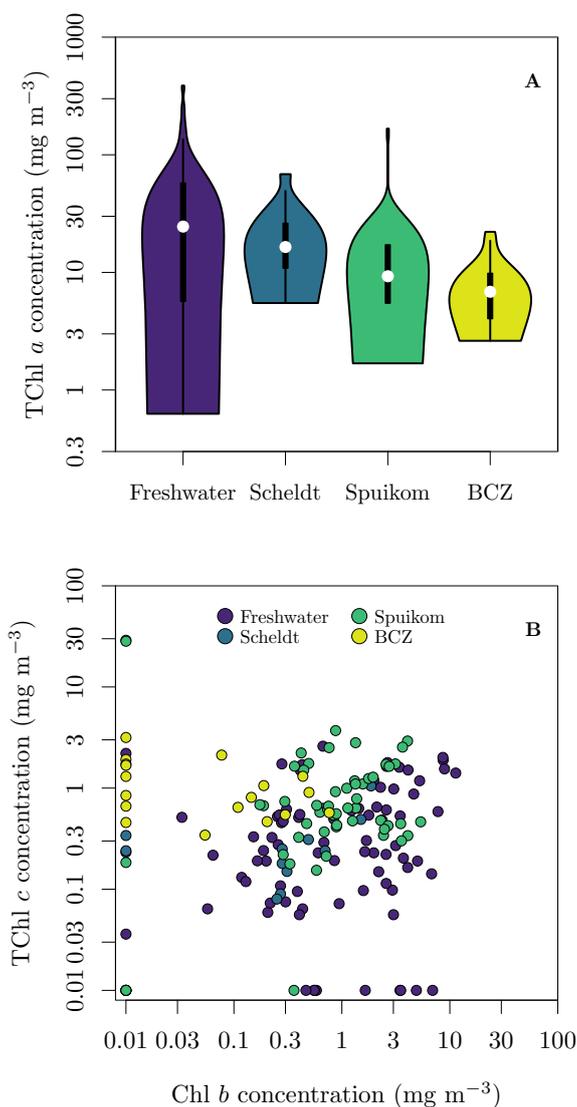
Figure 13. Pigment concentrations in Belgian water systems. (A) Total chlorophyll a (TChl a) concentration; and (B) relation between monovinyl chlorophyll b (Chl b) and total chlorophyll c (TChl c) concentrations. TChl a = monovinyl chlorophyll a + chlorophyllide a, TChl c = chlorophyll c_1c_2 + divinyl chlorophyll c_3 (*cf.* Table 6). A value of 0.01 mg m^{-3} was added to represent in the log-log plot the samples with concentrations below the detection limit.

2.5.4 DNA metabarcoding

445 DNA metabarcoding (amplicon sequencing) was performed only for samples from the Spuikom and the BCZ. The molecular analysis was based on replicate filters collected for pigment analysis. DNA extraction was performed with the DNeasy Plant

Mini Kit (Qiagen), with the polymerase chain reaction (PCR) amplification targeting the variable region 4 (V4) of the nuclear 18S ribossomal RNA gene. The 18S rRNA V4 primers were the TAReuk454FWD1 (5' CCAGCASCYGCGGTAATTCC 3') and the TAReukREV3 (5' ACTTTCGTTCTTGATYRA 3'; Stoeck et al., 2010). Paired-end (2 x 300 base pairs) sequencing was performed with the Illumina MiSeq technology (Illumina, San Diego, US) by Genewiz (Leipzig, Germany). The primers were trimmed from the sequenced reads using the FASTX-Toolkit (Gordon and Hannon, 2010). The resulting base-pair sequences were processed with the DADA2 algorithm (Callahan et al., 2016) to resolve amplicon single variants (ASVs). Probable contaminant sequences were removed using negative controls, following the method of Davis et al. (2018). Taxonomic assignment to the ASVs was based on the Protist Ribosomal Reference database (PR^2 version 4.12; Guillou et al., 2012). The raw molecular data can be found at the Sequence Read Archive (SRA) of the National Center for Biotechnology Information (NCBI) under the accession number PRJNA778668 (https://www.ncbi.nlm.nih.gov/sra/PRJNA778668).

The raw data was further processed to an aggregation level that is relevant for optical monitoring. The assigned taxonomy with PR^2 was updated to follow the taxonomy of the World Register of Marine Species (WoRMS Editorial Board, 2021), aggregated to species rank and filtered to remove non-pigmented organisms. Heterotrophic organisms were filtered at division rank, with the exception of exclusively heterotrophic dinoflagellates, which were filtered at the lowest rank possible based on reference sources (Hasle et al., 1997; WoRMS Editorial Board, 2021). The data was further annotated to indicate: (1) the pigmentation group (*sensu* Jeffrey et al., 2011) of each species based on our pigment ratio database; and (2) the toxicity, based on the IOC-UNESCO HAB reference list (Moestrup et al., 2021). More than 200 different species from 19 classes of phytoplankton were identified.

2.5.5 Flow imaging microscopy

As part of the regular LifeWatch BE samples in the BCZ, organisms in the range of 55 μm to 300 μm were counted and identified to the lowest taxonomic unit possible using flow imaging microscopy (FlowCam VS IV, Fluid Imaging Technologies, Inc.; Amadei Martínez et al., 2020; Flanders Marine Institute, 2021b). Fresh water samples were fixed with lugol and stored in the dark until analysis. The images were analyzed with the VisualSpreadsheet software (Fluid Imaging Technologies, Inc.) for automatic taxonomic identification. Data was visually inspected to validate the classification. The fixation with lugol results in the disaggregation of *Phaeocystis globosa* colonies and single-cell *P. globosa* were not detected due to the lower size range of the instrument configuration. In total, 30 different species were detected, the majority of which were diatoms. A comparison of the spatio-temporal pattern of relative abundance of *Rhizosolenia spp.* cells between amplicon sequencing and FlowCam is shown in Fig. 14.

2.5.6 Bottom cover

For systems in which the bottom can be visible from the surface, the bottom type was described. Bottom cover was classified based on visual inspection from the surface or from images taken with a submersible camera (Fig. 15). For some stations the bottom cover was classified based on sampling of bottom material. The discrete classes were: (1) Sediment (potentially with biofilm); (2) Shells; (3) *Cladophora*; (4) *Sargassum*; (5) Brown algae (not specified); and (6) Heterogeneous. The last class
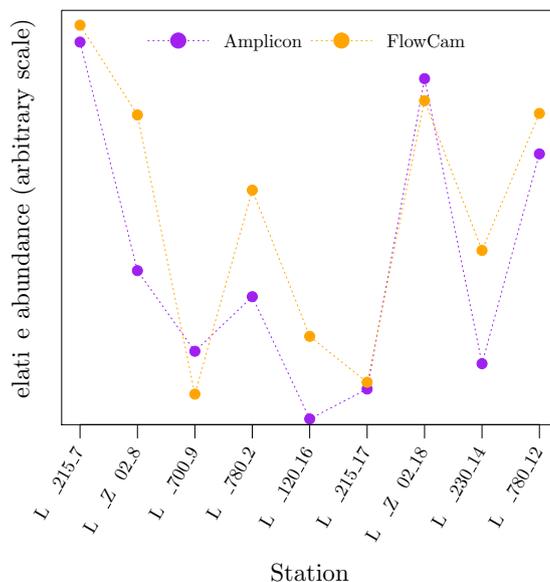
Figure 14. Relative abundance of *Rhizosolenia spp.* cells as estimated by DNA metabarcoding (amplicon sequencing) and microscopy (FlowCam). Data normalized using the counts of common species of diatoms in each dataset.

480   was used when the bottom was covered by a complex mixture of other classes. This classification is qualitative in the sense that describes the major composition of the bottom at a given station, but does not provide fractional cover.

2.6   Ancillary parameters

In addition to the optical and biogeochemical parameters, a series of ancillary parameters were also determined: Time, position, local depth, sampling platform, Sun zenith angle, and visual descriptions of the sky and water system.

485   For the inland water campaigns, time and position of each station were recorded from a handheld Global Navigation Satellite System (GNSS) receiver (GPSmap 62s, Garmin), which is enabled to receive corrections from the European Geostationary Navigation Overlay Service (EGNOS). The local depth at the time of sampling was estimated with a handheld single beam echosounder (Echotest II, Plastino) at the start of each station. The exception was one very shallow station (0.2 m), where a folding ruler was used. For marine stations, position, time and local depth were taken from the ship's navigation data, acquired

490   with a differential GPS and a mounted single beam ecosounder (JFE 380-25, Japan Radio Co., Ltd.). Sun zenith angles were calculated with the HORIZONS system (Jet Propulsion Laboratory, NASA; https://ssd.jpl.nasa.gov/horizons/). All ancillary parameters, with the exception of the Sun zenith angle, were combined into a single metadata file.
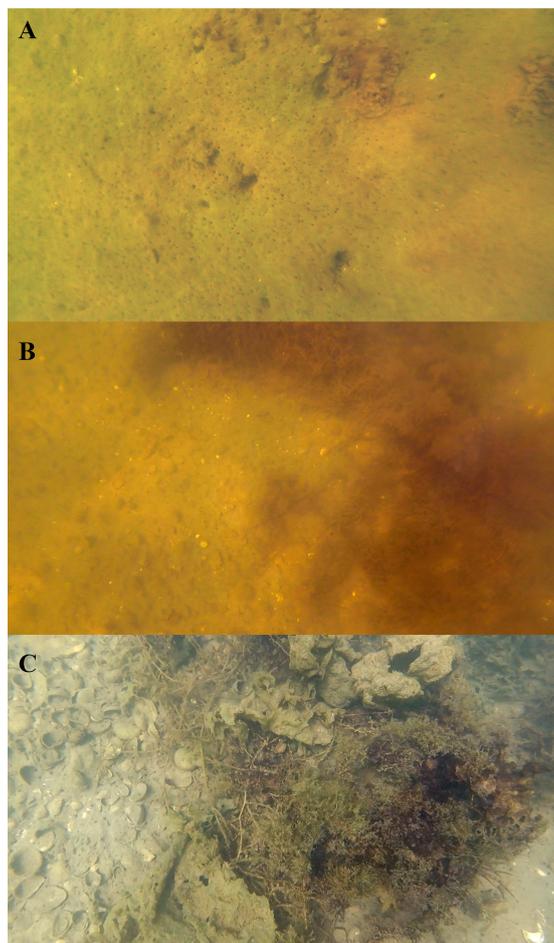
Figure 15. Example of bottom cover classes in the Spuikom. (A) Sediment (with biofilm); (B) Sediment, shells and brown algae; (C) Heterogeneous.

3 Conclusions and recommendations

This study described in detail the first open dataset of paired optical and biogeochemical measurements in a diverse set of water
495 systems located in Belgium. The wide range of observed conditions and the relative scarcity of similar open datasets in inland
waters make this a relevant contribution to the community. Potential users of this dataset are encouraged to contact the authors
for further inquiries concerning the data and an updated status of studies in development using the dataset.

Although the raw, measured IOPs and AOPs are provided, their fitted or corrected versions are recommended for use. For
example, the fitted a_g removes noise and oscillation in the NIR due to salinity mismatch between blanks and brackish or
500 marine samples. The fitted a_d was found as the least biased way to remove the absorption signal of NaClO, providing the best
estimate of a_d and $a_\mathrm{\phi}$. The b_p, and consequentially c_p, corrected to an acceptance angle of 0.018° (equivalent to the LISST

instruments) provides a more accurate estimate of the scattering and attenuation. The d_{Secchi} was corrected for the Sun zenith angle, normalizing all measurements to the Sun in the zenith, and $\rho_{\mathrm{wl}}^{\mathrm{L}}$ measured with the on-water method were corrected for platform and instrument shadowing. The results of the fitting and corrections were evaluated through consistency checks

505 between different data types and instruments as presented in the text.

4 Data availability

Data is available from Castagna et al. (2022), hosted at PANGAEA (http://www.pangaea.de) under the doi: https://www.pangaea.de/tok/c67200d99ea9bbbeadd9edec9690f937b5bacbff

Author contributions. AC, RD, and ID performed field campaigns in Belgian lakes. AC, HL, KR, MB, AD, and DD participated on field
510 campaigns in the Spuikom, Scheldt and BCZ. LA joined the Sheldt and BCZ campaigns, JM joined BCZ campaigns and HD joined Spuikom campaigns. AC performed laboratory measurements of IOPs, field and laboratory measurements of AOP, processing of LISST data, measurements of turbidity, SPM and mineral fraction, classification of the bottom cover, curated the data and wrote the manuscript. MB performed reflectance spectroscopy measurements in the BCZ. ID and RD performed HPLC analysis of pigments. RB and AC performed analysis of DNA metabarcoding. LA performed flow microscopy counts. HD, KS and WV provided infrastructure, planning and methodological
515 discussion. All authors reviewed the manuscript.

Competing interests. The authors declare that they have no conflict of interest.

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Table 1. Description of relevant acronyms, symbols, constants and subscripts used in this manuscript.

		Description
Acronyms	AOP	Apparent optical properties
	BCZ	Belgian coastal zone
	BRDF	Bidirectional reflectance distribution function
	CDOM	Chromophoric dissolved organic matter
	FNU	Formazin nephelometric units
	FWHM	Full-Width-at-Half-Maximum
	IOP	Inherent optical properties
	NIR	Near-infrared range of the electromagnetic spectrum (700 nm to 1000 nm)
	PMMA	Polymethyl methacrylate
	PSD	Particle size distribution
	PTFE	Polytetrafluoroethylene
	SPM	Suspended particulate matter
	UV	Ultraviolet range of the electromagnetic spectrum (10 nm to 400 nm)
Symbols	a	Spectral neperian absorption coefficient ($\mathrm{m^{-1}\,nm^{-1}}$)
	b	Spectral neperian scattering coefficient ($\mathrm{m^{-1}\,nm^{-1}}$)
	c	Spectral neperian beam attenuation coefficient ($\mathrm{m^{-1}\,nm^{-1}}$)
	d_{Secchi}	Secchi disk depth (m)
	$E_{\mathrm{dn}}(0^+)$	Spectral global (direct + diffuse) downwelling plane irradiance just above the water surface ($\mathrm{W\,m^{-2}\,nm^{-1}}$)
	f_{m}	Mineral fraction of SPM (1)
	l	Metric pathlength (m)
	L	Spectral radiance ($\mathrm{W\,m^{-2}\,sr^{-1}\,nm^{-1}}$)
	S_{h}	Spectral hyperbolic slope coefficient (1)
	S_{e}	Spectral exponential slope coefficient ($\mathrm{nm^{-1}}$)
	T	Transmittance (1)
	α_{d}	Spectral neperian depigmented particle absorption coefficient offset ($\mathrm{m^{-1}\,nm^{-1}}$)
	β	Spectral neperian volume scattering function ($\mathrm{m^{-1}\,sr^{-1}\,nm^{-1}}$)
	$\Delta\phi$	Relative azimuth angle (°)
	ϵ	Fractional error caused by shadowing of the downwelling light for on water spectroscopy measurements (1)
	θ	Nadir angle (°)
	λ	Wavelength (nm)
	ρ^{L}	Lambert-equivalent bi-hemispherical reflectance (1)
	$\tilde{\rho}_{\mathrm{f}}$	Effective Fresnel reflectance (1)
	ψ	Polar scattering angle (° or rad)
Constants	e	Euler's number (≈ 2.718282)
	π	The ratio of a circle's circumference to its diameter (≈ 3.141593)
Subscripts	b	Relative to backscattering
	d	Relative to the depigmented particle components
	g	Relative to the CDOM component
	nw	Relative to all non-water components
	p	Relative to the particle components
	s	Relative to the substrate (macroalgae, sediments)
	t	Relative to all components (total)
	wl	Relative to water-leaving
	ws	Relative to water-system (water-leaving + interface)
	φ	Relative to the *in vivo* pigments component

Table 2. Summary of Belgian water systems sampled in this study.

Type	System	Platform	Period
Fresh	Bocht	Boat, Pontoon	2017
	Donkmeer	Boat, Pontoon	2017-2018
	DelMare	Boat	2017
	Dikkebus	Pontoon	2018
	Hazewinkel	Boat, Pontoon	2017
	Leuven-Dijle	Pontoon	2019
	Molsbroek	Boat	2017
	Nieuwdonk	Pontoon	2017
	Zillebeke	Boat	2018
Brackish	Scheldt	Pontoon	2019
	Spuikom	Pontoon, Boat	2017-2019
Marine	BCZ	RV Simon Stevin	2018

Table 3. Summary of measured and derived inherent optical properties per water system. Values indicate the number of valid, quality checked, measurements.

Type	System	a_g	a_p	a_d	a_Φ	c_p	b_p	β_p	Turb
				$(\text{m}^{-1}\,\text{nm}^{-1})$				$(\text{m}^{-1}\,\text{sr}^{-1}\,\text{nm}^{-1})$	(FNU)
Fresh	Bocht	1	7	7	7	-	-	2	8
	Donkmeer	18	33	32	32	8	8	10	38
	DelMare	-	2	2	2	-	-	1	3
	Dikkebus	7	7	7	7	3	3	5	7
	Hazewinkel	1	17	17	17	-	-	1	18
	Leuven-Dijle	2	2	-	-	2	2	-	2
	Molsbroek	-	1	1	1	-	-	-	1
	Nieuwdonk	-	1	1	1	-	-	-	2
	Zillebeke	8	8	8	8	3	3	6	8
Brackish	Scheldt	21	21	-	-	21	21	2	21
	Spuikom	35	51	46	46	21	21	15	60
Marine	BCZ	9	19	19	19	-	-	19	19
Total		**102**	**169**	**140**	**140**	**58**	**58**	**61**	**187**

Table 4. Summary of measured apparent optical properties per water system.

Type	System	ρ_{wl}^{L} (1)	ρ_{s}^{L} (1)	d_{Secchi} (m)
Fresh	Bocht	7	-	8
	Donkmeer	34	-	37
	DelMare	-	-	4
	Dikkebus	7	-	7
	Hazewinkel	20	-	21
	Leuven-Dijle	-	-	-
	Molsbroek	-	-	1
	Nieuwdonk	-	-	2
	Zillebeke	7	-	7
Brackish	Scheldt	8	-	-
	Spuikom	62	14	68
Marine	BCZ	19	-	18
Total		**164**	**14**	**173**

Table 5. Summary of measured and derived biogeochemical parameters per water system.

Type	System	SPM (g m^{-3})	f_m (1)	PSD (cm^3 m^{-3})	Pigments (mg m^{-3})	Amplicon (counts)	FlowCam (counts)	Bottom (type)
Fresh	Bocht	8	8	2	8	-	-	-
	Donkmeer	34	34	10	36	-	-	-
	DelMare	2	2	1	3	-	-	-
	Dikkebus	7	7	5	7	-	-	-
	Hazewinkel	18	18	1	18	-	-	-
	Leuven-Dijle	2	2	-	-	-	-	-
	Molsbroek	1	1	-	1	-	-	-
	Nieuwdonk	2	2	-	2	-	-	1
	Zillebeke	8	8	6	8	-	-	-
Brackish	Scheldt	21	21	2	12	-	-	-
	Spuikom	56	48	15	53	48	-	26
Marine	BCZ	19	19	19	19	15	13	-
Total		**178**	**170**	**61**	**167**	**63**	**13**	**27**

Table 6. List of quantified pigments. Names and abbreviations follow Roy et al. (2011). Known co-elutions according to Van Heukelem and Thomas (2001) are described in the table footnotes.

Name	Abbreviation	Name	Abbreviation
(monovinyl) Chlorophyll a	Chl a	Diadinoxanthin	Diadino
(monovinyl) Chlorophyll b	Chl b	Diatoxanthin	Diato
Chlorophyll c_1c_2[†]	Chl c_1c_2[†]	Echinenone	Echin
(divinyl) Chlorophyll c_3	Chl c_3	Fucoxanthin	Fuco
Chlorophyllide a	Chlide a	19′-Butanoyloxyfucoxanthin	But-fuco
Pheophorbide a	Pheide a	19′-Hexanoyloxyfucoxanthin	Hex-fuco
Pheophytin a	Phe a	Lutein	Lut
α-Carotene	βε-Car	Mutatoxanthin	Mutato
β-Carotene	ββ-Car	Myxoxanthophyll	Myxo
Lycopene	Lyco	9′-cis-Neoxanthin[††]	c-Neo[††]
Alloxanthin	Allo	Peridinin	Peri
Antheraxanthin	Anth	Prasinoxanthin[†††]	Pras[†††]
Astaxanthin	Asta	Violaxanthin	Viola
Canthaxanthin	Cantha	Zeaxanthin	Zea
Crocoxanthin	Croco		
α-Cryptoxanthin	α-Cypto		
β-Cryptoxanthin	β-Cypto		

[†] Chl c_1c_2 = Chl c_1 + Chl c_2 + magnesium 2,4-divinylpheoporphyrin a_5 monomethyl ester (MgDVP). MgDVP is present in trace concentrations in most algae and cyanobacteria, but is a major component of PRASINO 3A and 3B pigmentation groups (*sensu* Jeffrey et al., 2011);

[††] c-Neo and 19′-Hexanoyloxy-4-ketofucoxanthin (Hex-kfuco) co-elute and are not know to co-occur in the same organism or pigmentation group (*sensu* Jeffrey et al., 2011);

[†††] Pras and micromonol (Microl) co-elute and co-occur in the PRASINO 3B pigmentation group (*sensu* Jeffrey et al., 2011).