



Chlorophyll-*a* growth rates and related environmental variables in global temperate and cold-temperate lakes

5 Hannah Adams¹, Jane Ye¹, Bhaleka Persaud¹, Stephanie Slowinski¹, Homa Kheyrollah Pour²,
Philippe Van Cappellen¹

¹ Ecohydrology Research Group, Department of Earth and Environmental Sciences and Water Institute, University of Waterloo, Waterloo, ON, Canada

10 ² ReSEC Research Group, Department of Geography and Environmental Studies, Wilfrid Laurier University,
Waterloo, ON, Canada

Correspondence to: Hannah Adams (hadams21@mun.ca)

15 **Keywords:** lake primary productivity, chlorophyll-*a*, growth window, growth rate, bottom-up controls, trophic state, ice phenology, solar irradiance



Abstract

Lakes are key ecosystems within the global biogeosphere. However, the bottom-up controls on the biological productivity of lakes, including surface temperature, ice phenology, nutrient loads and mixing regime, are increasingly altered by climate warming and land-use changes. To better understand the environmental drivers of lake productivity, we assembled a dataset on chlorophyll-*a* concentrations, as well as associated water quality parameters and surface solar irradiance, for temperate and cold-temperate lakes experiencing seasonal ice cover. We developed a method to identify periods of rapid algal growth from *in situ* chlorophyll-*a* time series data and applied it to measurements performed between 1964 and 2019 across 357 lakes, predominantly located north of 40°. Long-term trends show that the algal growth windows have been occurring earlier in the year, thus potentially extending the growing season and increasing the annual productivity of northern lakes. The dataset is also used to analyze the relationship between chlorophyll-*a* growth rates and solar irradiance. Lakes of higher trophic status exhibit a higher sensitivity to solar radiation, especially at moderate irradiance values during spring. The lower sensitivity of chlorophyll-*a* growth rates to solar irradiance in oligotrophic lakes likely reflects the dominant role of nutrient limitation. Chlorophyll-*a* growth rates are significantly influenced by light availability in spring but not in summer and fall, consistent with a switch to top-down control of summer and fall algal communities. The growth window dataset can be used to analyze trends in lake productivity across the northern hemisphere or at smaller, regional scales. We present some general trends in the data and encourage other researchers to use the open dataset for their own research questions.



40 **1 Introduction**

Lakes play an important role in the biogeochemical cycling of many elements (Battin et al., 2008; Cole et al., 2007; O’Connell et al., 2020; Rousseaux and Gregg, 2013; Schindler, 1971). With over 100 million documented lakes on earth (Verpoorter et al., 2014), evidence indicates that the majority of global lakes are shallow with enough light and nutrients available to make
45 them highly productive ecosystems (Downing et al., 2006; Wetzel, 2001). Lakes therefore represent active sites for the storage, transport, and transformation of carbon, nutrients (e.g., nitrogen, phosphorus, silicon, iron), and contaminants (e.g., mercury) along the freshwater continuum (Lauerwald et al., 2019; Tranvik et al., 2009).

There are multiple bottom-up controls on lake primary productivity, including water
50 temperature, ice phenology, nutrient concentrations, circulation, mixing regime, and solar radiation (Lewis, 2011). Stressors such as climate change and nutrient pollution can significantly impact these controls, altering the ecosystem structure and biogeochemical functioning of lakes (Jeppesen et al., 2020; Markelov et al., 2019). Changes affecting northern lakes include warmer water temperatures, enhanced stratification and hypoxia, nutrient enrichment, light attenuation
55 by chromophoric organic matter, and increases in the relative abundance of toxic cyanobacteria in the phytoplankton community (Deng et al., 2018; Huisman and Hulot, 2005; Jeppesen et al., 2003; Creed et al., 2018). For example, Lake Superior has seen an increase in primary production during the last century, together with increasing surface water temperatures and longer seasonal stratification and ice-free periods (O’Beirne et al., 2017). Other lakes are
60 similarly experiencing increases in productivity. According to Lewis (2011), the current mean primary production of lakes is $260 \text{ g C m}^{-2}\text{y}^{-1}$, which is 162% higher than earlier estimations under historical baseline conditions.

Phytoplankton (i.e., algae) are the main primary producers in lakes and generally make up the foundation of lentic food webs (Carpenter et al., 2016). Periods of high lake productivity
65 coincide with a rapid increase in phytoplankton biomass. In extreme cases, algal blooms can reach hundreds to thousands of cells per milliliter (Henderson-Seller and Markland, 1987). These bloom events produce large quantities of decomposing organic matter that cause the expansion of hypoxic conditions within the lake (Watson et al., 2016). In harmful algal blooms, certain algal species also release hepatotoxic and neurotoxic compounds (Codd et al., 2005). Thus,



70 identifying trends in the timing and intensity of seasonal algal growth, and linking them to changes in environmental stressors, can help predict the future of lake productivity and assess the risk of undesirable algal blooms.

Because it is challenging to measure algal population growth directly, chlorophyll-*a* is often used as a proxy for both the algae biomass and the associated primary production rate in lakes (Huot
75 et al., 2007). Although other proxies have been developed (Lyngsgaard et al., 2017), chlorophyll-*a* is the most common metric to characterize trends in algal biomass within and across lakes, especially in historical water quality records. Tett (1987) proposes a chlorophyll-*a* threshold of 100 μgL^{-1} to define “exceptional” blooms”, Jonsson et al. (2009) use a threshold of 5 μgL^{-1} to identify a bloom, while Binding et al. (2021) flags an algal bloom when the chlorophyll-*a*
80 concentrations extracted from satellite observations exceed 10 μgL^{-1} . Such threshold values, however, do not take into account the baseline (i.e., no-bloom) chlorophyll-*a* concentration specific to a given lake, or the lake’s trophic status (German et al., 2017). Furthermore, focusing on harmful and nuisance algal blooms alone may mask the impact that a changing climate or other stressors may have on a lake’s overall biological productivity.

85 Annual fluctuations in lake chlorophyll-*a* concentration are an indicator of the natural seasonal succession of algal species as a function of temperature, light, and nutrient availability (Lyngsgaard et al., 2017). For instance, in a dimictic lake algal growth in the spring tends to be controlled by these bottom-up controls, with light often being the primary limiting factor, while later in the summer or fall algal biomass may be more influenced by zooplankton grazing (i.e., a
90 top-down control), while nutrient availability may overtake solar radiation as the limiting resource for growth (Lewis et al., 2018; Lyngsgaard et al., 2017; Scofield et al., 2020).

A common approach for comparing chlorophyll-*a* trends across multiple lakes is to consider the maximum or mean annual chlorophyll-*a* concentrations. For example, Ho et al. (2020) used the Mann-Kendall trend test to analyze time series of annual maximum chlorophyll-*a* concentrations,
95 while Shuvo et al. (2021) used a random forest regression approach to assess the relative importance of climatic versus non-climatic controls on mean chlorophyll-*a* concentrations. However, these approaches do not specifically look at the periods of the year when algal biomass is primarily determined by bottom-up controls and exhibits rapid growth.



Alternatively, the rate of change in chlorophyll-*a* concentration can be used to capture the timing
100 of rapid increase in algal biomass associated with periods of high lake productivity. In this study,
we refer to these periods as “growth windows”. The weeks leading up to a growth window are
crucial to create the necessary environmental conditions that enable algal growth (Lewis et al.,
2018). Thus, to analyze trends in lake productivity one should consider environmental variables,
such as surface water temperature, solar radiation and nutrient concentrations, both during and
105 preceding the annual growth windows.

Although the rate of chlorophyll-*a* concentration growth has been used to detect algal blooms
within individual water bodies, for example in the San Roque reservoir (Germán et al., 2017), it
has rarely been used across large temporal (i.e., more than a few years) and spatial (i.e., regional
and up) scales. Here, we present a method for calculating seasonal chlorophyll-*a* growth rates
110 and then create a dataset of these rates derived from *in situ* chlorophyll-*a* concentrations obtained
in 357 lakes, most of which are at latitudes above 40° N. The entire dataset covers the period
from 1964 to 2019, and further contains data on coincident bottom-up environmental control
variables, including *in situ* surface solar radiation measurements. To illustrate the potential
applications of the dataset, we present some general trends of the chlorophyll-*a* rates and their
115 relationships with environmental variables. The dataset is made available as an open resource
that other researchers are encouraged to use in their own work.

2 Data and methods

All data processing, visualizations, and analyses were carried out with Python (ver. 3.7.6; Python
Software Foundation, 2021) using the pandas library (Reback et al., 2020), NumPy library
120 (Harris et al., 2020), and Dplython library (Riederer, 2015), while QGIS/PYQGIS was used for
all spatial data analyses (ver. 3.16; QGIS Development Team, 2021).

2.1 Data acquisition, compilation, and quality control

2.1.1 Lake data selection

In situ chlorophyll-*a* concentrations and other lake physico-chemical data were collected from
125 open source international, national, and regional databases. The data include surface water
temperature, Secchi depth and pH, as well as the concentrations of particulate organic carbon
(POC), total phosphorus (TP), soluble reactive phosphorus (SRP), total Kjeldahl nitrogen (TKN)
and dissolved organic carbon (DOC). We selected lakes from latitudes $\geq 40^\circ$ N to reduce the



latitude-dependent variability in mixing and thermal regimes, both of which exert a strong
130 control on lake productivity (Kirillin et al., 2012). At mid-to-high latitudes most lakes are
dimictic with seasonal ice cover while low-latitude lakes are typically meromictic or polymictic
(Woolway and Merchant, 2019). High-elevation lakes at lower latitudes can experience similar
effects from the transition from winter to spring, even without ice cover (Deng et al., 2020). We
therefore included the extensively monitored Lake Kasumigaura in Japan and Lake Taihu in
135 China in our study, although they are located at latitudes lower than 40°N.

Chlorophyll-*a* measurements are collected at variable water depths by different lake monitoring
agencies and researchers. For consistency, we only included measurements taken at ≤ 3 m depth.
When the sampling depth was not provided, we assumed the sample was taken from within the
top 0.5-3 m of the lake, given that this is standard sampling protocol (Dorset Environmental
140 Science Centre, 2010; United States Environmental Protection Agency, 2012).

We omitted all variable values below the corresponding analytical detection limit. Data from
different sources were individually reformatted to yield consistent (standard) units and headings.
Reported values were averaged to yield daily values mean before being combined into a single
csv file. When multiple chlorophyll-*a* data types were available (as, for example, in the
145 Laurentian Great Lakes data series), we selected the uncorrected data because most reported lake
chlorophyll-*a* concentrations have not been corrected for phaeophytin pigments. If no
coordinates were provided, we assigned those of the lake centroid in QGIS or estimated based on
the location name. Fifteen lakes had no known location and were removed from the final dataset.
We further restricted ourselves to lakes that were sampled at least 8 times per year. This was
150 found to be the minimum number of sampling points required to detect the growth windows. The
location of all lake sampling locations in the growth window dataset are shown in Figure 1.

After the above selection and quality assessments, the final dataset used for calculating the
growth windows contained 52116 unique data points (62% of the original data) for 357 lakes, all
 $\geq 40^\circ\text{N}$ (except Lake Kasumigaura and Lake Taihu), covering the period 1964-2019.

155 2.1.2 Surface solar radiation data

Open source *in situ* surface solar radiation (SSR) data for the period 1950-2020 were collected
from stations paired with the selected lakes. Each lake was paired with the closest SSR station
using the nearest neighbor function in QGIS, allowing for a maximum radius of three degrees



(Schwarz et al., 2018; Figure 1). In the dataset, the geodesic distance between each lake and its
160 paired SSR station is given, as well as the difference in elevation.

The SSR data temporal resolution varied from minutes to months. Hence, where needed, the SSR
data were resampled to yield monthly mean values. For the Experimental Lakes Area (ELA) in
Ontario, Canada, the data were converted from photosynthetically active radiation (PAR) to
SSR, where the PAR wavelength range (400-700 nm) was averaged to 550 nm.

165 2.1.3 Lake characteristics

For each lake, we calculated the trophic status index (TSI) based on the mean chlorophyll-*a*
concentration over the sampling period. This TSI value was used to assign the lake to the
corresponding trophic state category according to Carlson and Simpson (1996). The
HydroLAKES shapefile yielded the lake's surface area, mean depth, elevation, and volume
170 (Messenger et al., 2016). The climate zone of the lake was extracted from the HydroATLAS
shapefile (Linke et al., 2019).

2.2 Detecting seasonal growth windows

Growth windows were defined based on the rate of change in chlorophyll-*a* concentration at each
lake sampling point throughout the year. To locate the start and end of a growth window, we
175 smoothed the annual chlorophyll-*a* time series using a Savitzky-Golay filter (SciPy.signal
savgol_filter) and flagged optima in the smoothed data (SciPy.signal find_peaks) using functions
from the open source SciPy ecosystem (Virtanen et al., 2020). The procedure is illustrated in
Figure 2.

For each year, the spring growth window began when the daily rate of increase surpassed the
180 threshold of $0.05 \mu\text{gL}^{-1}\text{day}^{-1}$ for the first time. The $0.05 \mu\text{gL}^{-1}\text{day}^{-1}$ rate was chosen because it
corresponds to the median rate at which a distinct switch to a “rapid growth” period in the
mesotrophic-hypereutrophic lakes in the dataset was observed. The growth window ended at the
first “peak” in chlorophyll-*a* concentration. If a threshold rate of $0.05 \mu\text{gL}^{-1}\text{day}^{-1}$ was never
reached during a given year, the growth window began when the rate of change first became
185 positive. The summer (or fall) window was identified in the same way following the end of the
spring window. If there was only one peak in the smoothed data, only one growth window was
identified for that year. This year was then labelled as a “single growth window” year (i.e., only



one major algal growth window occurred within that year). Years with more than three chlorophyll-*a* peaks, or with no peaks, were not included in the growth window dataset.

190 Depending on data availability, the pre-growth window was defined as the one or two week period immediately preceding the growth window start date. For each pre-growth window, the mean surface water temperature, SSR, and TP concentration were calculated. These served as (simple) indicators of how favorable in-lake conditions were to initiate algal growth (Lyngsgaard et al., 2017). An example of a spring and summer growth window is shown in Figure 3. Note that
195 we use the label “summer” to indicate the second yearly growth window, although in many cases the summer growth window occurred after the fall equinox.

Once the growth window and pre-growth durations were determined, the mean values of the variables listed in Table 1 were calculated for both the growth window and the pre-growth window. This was done for each lake and for each year data were available. In the dataset, each
200 row represents a single growth window and includes the timing and duration, rate of increase of the chlorophyll-*a* concentration, and all other relevant lake variables, including SSR. Note that, along with the variables in Table 1, we included the total number of samples collected each year so the dataset can be filtered for sampling frequency. The reader is referred to the supplementary information included with the dataset for a more detailed explanatory table with additional
205 information on the organization carrying out the monitoring, physiological attributes of each lake, and years that data are available for a given sampling location.

3 Dataset: data distributions

3.1 Dataset characteristics

Most lakes in the dataset are located between 50 and 60° N as the majority of available open data
210 are from organizations within the United Kingdom, Sweden, Canada, and the United States. The years with available data in the dataset are unevenly distributed, however, with most detected growth windows falling in the period 2005-2019, likely due to a combination of increased lake monitoring efforts and a push in recent years towards greater accessibility of publicly funded data (Hallegraeff et al., 2021; Roche et al., 2020; Figure 4a).

215 The majority of growth windows recorded in the dataset fall in the eutrophic category (1.6% oligotrophic, 18.0% mesotrophic, 75.4% eutrophic, and 5.0% hypereutrophic). Single growth



windows dominate oligotrophic lakes where they make up 96% of all growth windows (Figure 4b). This may reflect the severe nutrient limitation in oligotrophic lakes, which prevents the occurrence of a second annual algal growth window (Rigosi et al., 2014). Oligotrophic lakes also
220 tend to occur at the higher latitudes (Figure 4c) where lower water temperatures and solar radiation may further limit algal growth (Lewis, 2011).

The growth window durations range from 2 to 251 days, with a median of 71 days across all lakes (Figure 5a). Summer growth windows tend to be shorter than those of spring and single growth windows, with the latter exhibiting the most variable start and end dates (Figure 5b).

225 **3.2 Environmental conditions during growth windows**

Chlorophyll-*a* rates during the growth windows exhibit log-normal distributions (Figure 6a). The mean chlorophyll-*a* rate is lowest in the single growth window category and highest in the summer growth windows. Mean surface water temperature has a distinct bimodal spring-summer distribution (Figure 6b), which is expected for northern temperate and cold-temperate lakes
230 where surface water temperature during the ice-free period follows the seasonal air temperature trend (Kirillin et al., 2012). For the single growth windows, temperature is evenly distributed across the annual range, which aligns with the large variability in the timing of single growth windows (Figure 5b). Total phosphorus concentrations are lowest during the spring growth windows, which likely reflects a greater control of P limitation on algal growth during spring
235 compared to summer and fall (Kirillin et al., 2012; 6c). Secchi depth during the growth windows ranges from 0.01 to 14.6 m, with summer growth windows experiencing the lowest mean Secchi depth, as turbidity generally increases after the spring bloom (Figure 6d).

4 Dataset: trend analyses

The growth window delineation and the estimation of chlorophyll-*a* rates can in principle be
240 applied to any lake for which time series chlorophyll-*a* concentration data are available. By creating a dataset comprising many lakes and covering multi-year time periods, it becomes possible to analyze global trends in lake productivity. Here, we provide a few illustrative examples of how the dataset can be interrogated, thereby setting the stage for its use by other researchers.



245 **4.1 Chlorophyll-*a* rates: trophic status and latitude**

When grouped by trophic status, mean and median chlorophyll-*a* growth rates show the expected increase from oligotrophic to hypereutrophic lakes (Figure 7a). The rates in the different trophic categories, however, cover very large and overlapping ranges. When grouped according to latitude, lakes between 40 and 50° N exhibit the widest range in chlorophyll-*a* rates (Figure 7b) that, in part, reflects the high proportion of lakes in this latitude range. The highest latitude lakes (60-70° N) tend to have the lowest chlorophyll-*a* rates, which is expected given the cooler temperatures and lower solar irradiance they experience (Lewis, 2011).

While differences in chlorophyll-*a* rates usually indicate comparable differences in algal biomass growth rates, it is important to note that the chlorophyll-*a* to biomass ratio varies within and among lakes. In particular, chlorophyll-*a* to biomass ratios are known to be sensitive to variations in solar irradiance and temperature (Behrenfeld et al., 2016). The summer ratio of chlorophyll-*a* to biomass (typically expressed as particulate organic carbon concentration) generally decreases with increasing latitude because the algae are adapted to the more variable daylight conditions, including longer summer photoperiods, at higher latitudes (Behrenfeld et al., 2016). By contrast, cooler temperatures at higher latitudes may result in higher chlorophyll-*a* to biomass ratios because of lower growth rates, at least when the algae are nutrient-replete (Behrenfeld et al., 2016).

4.2 Chlorophyll-*a* rates: temperature and climate warming

The start and end dates of the spring, single and summer growth windows show temporal trends towards occurrence earlier in the year (Figure 8a). The trends are most pronounced for the spring windows, which likely reflects a greater sensitivity of springtime algal activity to climate warming. The latter causes earlier ice break-up and produces earlier surface water temperature conditions favorable for algal growth (Markelov et al., 2019). This hypothesis is consistent with the correlations between the chlorophyll-*a* rates and water temperature (Figure 8b).

The start and end dates of the spring growth windows show a positive correlation with increasing temperature (Figure 8b). By contrast, little or even negative correlations are seen for the summer growth windows. Thus, all other conditions unchanged, a warmer climate would see earlier spring blooms, but little temporal shifts for the summer growth windows and, possibly, even a slight delay. For the spring and single growth windows, the duration of the window shows a



275 maximum around 10° C. Therefore, moderate temperatures close to 10° C should, on average,
produce the longest lasting algal growth events. No distinct trend is seen for summer growth
windows, presumably because they occur when water temperatures are already above 10° C.

4.3 Chlorophyll-*a* rates: solar irradiance

Solar radiation is essential for phytoplankton growth (Inomura et al., 2020). For example, at the
280 single lake scale, Tian et al. (2017) showed that SSR is a major predictor of growing-season
chlorophyll-*a* concentrations in the Western Basin of Lake Erie. A paleolimnological study of
Lake Tanganyika also provided evidence for a positive correlation between multi-centennial
oscillations of SSR and diatom productivity dating back to ~1000 CE (McGlue et al., 2020).
Nonetheless, the relationship between algal growth and SSR has yet to be compared across a
285 large set of lakes.

Solar radiation is used directly by photosynthetic organisms for carbon fixation (Melkozernov
and Blankenship, 2007). In addition, SSR exerts a strong control on lake surface water
temperature (Jakkila et al., 2009) and the timing of ice breakup in seasonally ice-covered lakes
(Kirillin et al., 2012b), both of which influence lake primary productivity. While the global
290 distribution of mean annual SSR is primarily a function of latitude (Kirillin et al., 2012b),
atmospheric controls (e.g., cloud cover) cause regional variability, as well as variability over
time (Alpert and Kishcha, 2008; Cutforth and Judiesch, 2007; Wild, 2009). It is important to note
that SSR is not related directly to global warming (Kirillin et al., 2012b), nor is it controlled by
the cycles in the sun's energy output (Wild, 2009).

295 To determine to what extent SSR explains variations in chlorophyll-*a* growth rates, we removed
the effect of temperature by normalizing the rates using the temperature dependency function
(which we refer to as "*f_{temp}*") proposed by Rosso et al. (1995). This function describes the non-
linear temperature dependence of cellular metabolic activity and requires that a minimum,
maximum, and optimum growing temperature be assigned. Dividing the *in situ* chlorophyll-*a*
300 rate during the growth window by the corresponding *f_{temp}* value corrects for the effect of
differences in temperature between growth windows.

The temperature-corrected chlorophyll-*a* growth rates indicate that the relationship between SSR
and algal growth is a function of the trophic status (i.e., nutrient availability), as seen in Figure 9.
Lakes of higher trophic status are more sensitive to SSR than lakes of lower trophic status. For



305 eutrophic lakes, the effect of SSR on the temperature-corrected chlorophyll-*a* rates is most pronounced in the low to moderate SSR range typical of the spring season (Figure 9a). The same effect is not seen when considering the rates without temperature correction (Figure 9b). Thus, the increasing SSR during spring is counterbalanced by cooler temperatures compared to the later summer growth window. Note that the summer chlorophyll-*a* growth rates show little
310 influence from SSR, whether corrected or not for temperature, supporting the theory of a greater top-down control on algal growth during the summer versus the spring as proposed, among others, by Lyngsgaard et al. (2017).

The chlorophyll-*a* growth rate data near or above 200 Wm⁻² remain low, with no clear dependence on SSR. This is likely indicative of a photoacclimation response of the algae, where
315 they produce less chlorophyll-*a* in proportion to their total biomass so they can allocate more resources to growth when nutrients – not light – are limiting growth (Lewis et al., 2018; Inomura et al., 2020). Furthermore, when light intensity during the summer months reaches damaging levels, algae may start producing additional photosynthetic pigments to protect their chlorophyll (so-called sunscreen pigments). However, nutrient availability may limit the amount of pigments
320 that can be synthesized, impeding the photoacclimation response (Lewis et al., 2018). This nutrient limitation of the photoacclimation response would explain the differences in the temperature corrected growth rate's sensitivity to SSR as a function of trophic status (Figure 9a). Lakes of higher trophic status (i.e., less nutrient limitation) show a larger response to changes in SSR, presumably because they have sufficient nutrients to produce additional chlorophyll-*a* in
325 response to an increase in SSR.

5 Key findings

The following points summarize the general trends that emerged from our analysis of the dataset.

1. Higher water temperatures and reduced ice-cover cause algal growth windows to start earlier in the year, extending the growing season and potentially increasing annual net primary
330 productivity of northern lakes under ongoing and future climate warming.
2. Chlorophyll-*a* growth rates increase with nutrient availability while they decrease at higher latitudes due to cooler temperatures and lower SSR.
3. Oligotrophic lakes tend to have the highest proportion of single annual growth windows, likely reflecting the dominant role of nutrient limitation.



- 335 4. Temperature-corrected chlorophyll-*a* growth rates suggest a relationship with SSR that depends on the trophic state of lakes:
- a. compared to mesotrophic and oligotrophic lakes, eutrophic lakes exhibit a higher sensitivity to SSR, especially in the low to moderate irradiance levels experienced during spring;
 - 340 b. at the upper end of SSR, chlorophyll-*a* growth rates remain low and independent of SSR, which may reflect a photoacclimation response of algae.
5. The low SSR sensitivity of chlorophyll-*a* growth rates during summer and fall suggests a stronger top-down control on algal growth compared to the earlier spring growth windows.
6. In summary, light limitation is an important control on chlorophyll-*a* growth rates during
345 spring, whereas lower nutrient availability and increased grazing from zooplankton tend to be more significant during summer.

6 Conclusions

We present a novel way to delineate periods of rapid algal growth, or growth windows, in lakes based on time series chlorophyll-*a* measurements. We apply this approach to derive the
350 chlorophyll-*a* growth rates occurring during the growth windows of 357 lakes from cold and cold-temperate regions in the northern hemisphere, using data collected between 1964 and 2019. The derived growth rates are assembled in an open-source dataset, together with additional information on the lakes including data on water quality, trophic state, and solar radiation. Note that the dataset can be paired with databases such as the [HydroLAKES](#), [HydroATLAS](#) and
355 GLCP databases to access additional lake and/or watershed attributes. Our dataset is designed to support comparative analyses of the controls on algal productivity within and between lakes. We present several examples of such analyses. We hope these will encourage others to use the dataset in their own research and to further expand the dataset's geographical reach and information content.

360 Code and data availability

All code is available in the project GitHub repository (https://github.com/hfadams/growth_window) and in Zenodo (<https://doi.org/10.5281/zenodo.5171442>). The growth window dataset and supplementary data



files are available in the Federated Research Data Repository at
365 <https://doi.org/10.20383/102.0488> (Adams et al., 2021).

Author contributions

All authors took part in development of the study. SS, BP, and HKP conceptualized the study, while HA and JY developed methods and carried out data collection and data post-processing. HA wrote the original manuscript with contributions from JY, BP, SS, HKP, and PVC. All
370 authors reviewed and edited the final paper.

Competing interests

The authors declare that they have no conflict of interest.

Acknowledgments

This work is funded by the Canada First Research Excellence Fund's Global Water Futures
375 Programme. We would also like to thank all institutions listed in the supplementary information for making their data open source and free to support this research.

380

385



References

- 390 Adams, H., Ye, J., Slowinski, S., Persaud, B., Kheyrollah Pour, H. and Van Cappellen, P.: Chlorophyll-a growth rates and related environmental variables in global temperate and cold-temperate lakes, *Fed. Res. Data Repos.*, <https://doi.org/10.20383/102.0488>, 2021.
- Alberta Climate Information Service: Alberta Weather Station Database, Available from: <https://agriculture.alberta.ca/acis/weather-data-viewer.jsp>, 2020.
- 395 Alberta Lake Management Society: Alberta Environment and Parks data repository, Available from: <http://environment.alberta.ca/apps/EdwReportViewer/LakeWaterQuality.aspx>, 2020.
- Alpert, P. and Kishcha, P.: Quantification of the effect of urbanization on solar dimming, *Geophys. Res. Lett.*, 35(8), 1–5, doi:10.1029/2007GL033012, 2008.
- 400 Battin, T. J., Kaplan, L. A., Findlay, S., Hopkinson, C. S., Marti, E., Packman, A. I., Newbold, J. D. and Sabater, F.: Biophysical controls on organic carbon fluxes in fluvial networks, *Nat. Geosci.*, 1(2), 95–100, doi:10.1038/ngeo101, 2008.
- Behrenfeld, M. J., O'Malley, R. T., Boss, E. S., Westberry, T. K., Graff, J. R., Halsey, K. H., Milligan, A. J., Siegel, D. A. and Brown, M. B.: Revaluating ocean warming impacts on global phytoplankton, *Nat. Clim. Chang.*, 6(3), 323–330, doi:10.1038/nclimate2838, 2016.
- 405 Carlson, R. E. and Simpson, J.: A coordinator's guide to volunteer lake monitoring methods., *North Am. Lake Manag. Soc.*, 96, 1996.
- Carpenter, S., Kitchell, J., Cole, J. and Pace, M.: Cascade Project at North Temperate Lakes LTER Core Data Process Data 1984 - 2016 [Data set], *Environ. Data Initiat.*, doi:<https://doi.org/10.6073/PASTA/6A658526E313DBCECBC0331A1F343C01>, 2017.
- 410 Carpenter, S. R., Kitchell, J. F., Hodgson, J. R.: Cascading trophic interactions and lake productivity, , 35(10), 634–639, 2016.
- Codd, G. A., Morrison, L. F. and Metcalf, J. S.: Cyanobacterial toxins: Risk management for health protection, *Toxicol. Appl. Pharmacol.*, 203(3 SPEC. ISS.), 264–272, doi:10.1016/j.taap.2004.02.016, 2005.
- 415 Cole, J., J., Prairie, Y. T., Caraco, N. F., McDowel, L. T., Tranvik, L. J., Striegel, C. M., Duarte, C. M., Kortelainen, P., Downing, J. A., Middleburg, J. J. and Melack, J.: Plumbing the Global Carbon Cycle: Integrating Inland Waters into the Terrestrial Carbon Budget, *Ecosystems*, 10(1), 171–184, doi:10.1007/s, 2007.
- 420 Creed, I. F., Bergström, A. K., Trick, C. G., Grimm, N. B., Hessen, D. O., Karlsson, J., Kidd, K. A., Kritzberg, E., McKnight, D. M., Freeman, E. C., Senar, O. E., Andersson, A., Ask, J., Berggren, M., Cherif, M., Giesler, R., Hotchkiss, E. R., Kortelainen, P., Palta, M. M., Vrede, T. and Weyhenmeyer, G. A.: Global change-driven effects on dissolved organic matter composition: Implications for food webs of northern lakes, *Glob. Chang. Biol.*, 24(8), 3692–3714, doi:10.1111/gcb.14129, 2018.
- Cutforth, H. W. and Judiesch, D.: Long-term changes to incoming solar energy on the Canadian Prairie, *Agric. For. Meteorol.*, 145(3–4), 167–175, doi:10.1016/j.agrformet.2007.04.011, 2007.
- 425 Deng, J., Paerl, H. W., Qin, B., Zhang, Y., Zhu, G., Jeppesen, E., Cai, Y. and Xu, H.: Climatically-modulated decline in wind speed may strongly affect eutrophication in shallow lakes, *Sci. Total Environ.*, 645, 1361–1370, doi:10.1016/j.scitotenv.2018.07.208, 2018.
- Deng, J., Zhang, W., Qin, B. Q., Zhang, Y., Salmaso, N. and Jeppesen, E.: Winter Climate Shapes Spring Phytoplankton Development in Non-Ice-Covered Lakes: Subtropical Lake Taihu as an Example, *Water*



- 430 Resour. Res., 56(9), 1–11, doi:10.1029/2019WR026680, 2020.
Department of Fisheries and Oceans: Lake Winnipeg Chemistry data from 2002 to 2004, Can. Watershed Inf. Netw., doi:10.25976/4t8h-zn16, 2004.
Department of Fisheries and Oceans: Lake Winnipeg Chemistry, 1994–2012, CanWIN Data HUB, doi:10.5203/ywqt-vp63, 2012.
- 435 Dorset Environmental Science Centre: Lakeshore Capacity Assessment Handbook: Protecting Water Quality in Inland Lakes on Ontario’s Precambrian Shield Appendix C, Dorset Environmental Science Centre Technical Bulletins., Toronto, Ontario., 2010.
Downing, J. A., Prairie, Y. T., Cole, J., J., Duarte, C. M., Tranvik, L. J., Striegl, R. G., McDowell, W. H., Kortelainen, P., Caraco, N. F., Melack, J., M. and Middelburg, J. J.: The global abundance and size distribution of lakes, ponds and impoundments, *Limnol. Oceanogr.*, 51(5), 2388–2397, doi:10.1016/B978-012370626-3.00025-9, 2006.
- 440 Driemel, A., Augustine, J., Behrens, K., Colle, S., Cox, C. J., Cuevas-Agulló, E., Denn, F. M., Duprat, T., Dutton, E. G., Fukuda, M., Grobe, H., Haeffelin, M., Hodges, G., Hyett, N., Ijima, O., Kallis, A., Knap, W., Kustov, V., Lanconelli, C., Long, C., Longenecker, D., Lupi, A., Maturilli, M., Mimouni, M., Ntsangwane, L., Ogihara, H., Olano, X., Olefs, M., Omori, M., Passamani, L., Pereira, E. B., Schmithüsen, H., Schumacher, S., Sieger, R., Tamlyn, J., Vogt, R., Vuilleumier, L., Xia, X., Ohmura, A. and König-Langlo, G.: Baseline surface radiation data (1992–2017), doi:10.1594/PANGAEA.880000, 2018.
- 445 Environment and Climate Change Canada: Great Lakes Water Quality Monitoring and Surveillance Data, Open Gov., doi:10.18164/495eb10d-d423-432a-980f-264ef287d45b, 2015.
Environment and Climate Change Canada: Hamilton Harbour Water Quality Data, Open Gov., doi:10.18164/05c4fa07-2a1e-46fa-a98a-1dc64bb97326, 2018.
ETH Zurich: Global Energy Balance Archive- Solar radiation data set, Available from: <https://geba.ethz.ch/>, 2020.
- 455 National Institute for Environmental Studies Biodiversity Area: GEMS database, Available from: <http://db.cger.nies.go.jp/gem/inter/GEMS/database/kasumi/contents/datalist.html>, 2020.
German, A., Tauro, C., Scavuzzo, M. C. and Ferral, A.: Detection of algal blooms in a eutrophic reservoir based on chlorophyll-a time series data from MODIS, *Int. Geosci. Remote Sens. Symp.*, 2017–July(April), 4008–4011, doi:10.1109/IGARSS.2017.8127879, 2017.
- 460 Gleick, P.: Water and conflict, *Int. Secur.*, 18(79), 112, doi:10.1016/S0262-4079(13)60875-1, 1993.
Hallegraeff, G. M., Anderson, D. M., Belin, C., Bottein, M.-Y. D., Bresnan, E., Chinain, M., Enevoldsen, H., Iwataki, M., Karlson, B., McKenzie, C. H., Sunesen, I., Pitcher, G. C., Provoost, P., Richardson, A., Schweibold, L., Tester, P. A., Trainer, V. L., Yñiguez, A. T. and Zingone, A.: Perceived global increase in algal blooms is attributable to intensified monitoring and emerging bloom impacts, *Commun. Earth Environ.*, 2(1), doi:10.1038/s43247-021-00178-8, 2021.
- 465 Harris, C. R., Millman, K. J., van der Walt, S. J., Gommers, R., Virtanen, P., Cournapeau, D., Wieser, E., Taylor, J., Berg, S., Smith, N. J., Kern, R., Picus, M., Hoyer, S., van Kerkwijk, M. H., Brett, M., Haldane, A., del Río, J. F., Wiebe, M., Peterson, P., Gérard-Marchant, P., Sheppard, K., Reddy, T., Weckesser, W., Abbasi, H., Gohlke, C. and Oliphant, T. E.: Array programming with NumPy, *Nature*, 585(7825), 357–362, doi:10.1038/s41586-020-2649-2, 2020.
Henderson-Seller, B. and Markland, H. R.: Decaying Lakes—The Origins and Control of Cultural



- Eutrophication, New York, NY., 1987.
- Huisman, J. and Hulot, F. D.: Population Dynamics of Harmful Cyanobacteria, *Harmful Cyanobacteria*, 143–176, doi:10.1007/1-4020-3022-3_7, 2005.
- 475 Huot, Y., Babin, M., Bruyant, F., Grob, C., Twardowski, M. S. and Claustre, H.: Relationship between photosynthetic parameters and different proxies of phytoplankton biomass in the subtropical ocean, *Biogeosciences*, 4(5), 853–868, doi:10.5194/bg-4-853-2007, 2007.
- IISD: IISD-ELA private database, Available from: <https://www.iisd.org/ela/science-data/our-data/data-requests/>, 2020.
- 480 Inomura, K., Omta, A. W., Talmy, D., Bragg, J., Deutsch, C. and Follows, M. J.: A Mechanistic Model of Macromolecular Allocation, Elemental Stoichiometry, and Growth Rate in Phytoplankton, *Front. Microbiol.*, 11(February), 1–22, doi:10.3389/fmicb.2020.00086, 2020.
- Jakkila, J., Leppäranta, M., Kawamura, T., Shirasawa, K. and Salonen, K.: Radiation transfer and heat budget during the ice season in Lake Pääjärvi, Finland, *Aquat. Ecol.*, 43(3), 681–692, doi:10.1007/s10452-009-9275-2, 2009.
- 485 Jeppesen, E., Søndergaard, M. and Jensen, J. P.: Climatic warming and regime shifts in lake food webs - Some comments, *Limnol. Oceanogr.*, 48(3), 1346–1349, doi:10.4319/lo.2003.48.3.1346, 2003.
- Jeppesen, E., Canfield, D. E., Bachmann, R. W., Søndergaard, M., Havens, K. E., Johansson, L. S., Lauridsen, T. L., Sh, T., Rutter, R. P., Warren, G., Ji, G. and Hoyer, M. V.: Toward predicting climate change effects on lakes: a comparison of 1656 shallow lakes from Florida and Denmark reveals substantial differences in nutrient dynamics, metabolism, trophic structure, and top-down control, *Inl. Waters*, 10(2), 197–211, doi:10.1080/20442041.2020.1711681, 2020.
- Jonsson, P. R., Pavia, H. and Toth, G.: Formation of harmful algal blooms cannot be explained by allelopathic interactions, *Proc. Natl. Acad. Sci. U. S. A.*, 106(27), 11177–11182, doi:10.1073/pnas.0900964106, 2009.
- 495 Kirillin, G., Leppäranta, M., Terzhevik, A., Granin, N., Bernhardt, J., Engelhardt, C., Efremova, T., Golosov, S., Palshin, N., Sherstyankin, P., Zdorovenнова, G. and Zdorovennov, R.: Physics of seasonally ice-covered lakes: A review, *Aquat. Sci.*, 74(4), 659–682, doi:10.1007/s00027-012-0279-y, 2012a.
- Kirillin, G., Leppäranta, M., Terzhevik, A., Granin, N., Bernhardt, J., Engelhardt, C., Efremova, T., Golosov, S., Palshin, N., Sherstyankin, P., Zdorovenнова, G. and Zdorovennov, R.: Physics of seasonally ice-covered lakes: A review, *Aquat. Sci.*, 74(4), 659–682, doi:10.1007/s00027-012-0279-y, 2012b.
- 500 Lauerwald, R., Regnier, P., Figueiredo, V., Enrich-Prast, A., Bastviken, D., Lehner, B., Maavara, T. and Raymond, P.: Natural Lakes Are a Minor Global Source of N₂O to the Atmosphere, *Global Biogeochem. Cycles*, 33(12), 1564–1581, doi:10.1029/2019GB006261, 2019.
- 505 Lewis, K. M., Arntsen, A. E., Coupel, P., Lowry, K. E., Dijken, G. L. Van, Selz, V., Arrigo, K. R., Matsuoka, A. and Mills, M. M.: Photoacclimation of Arctic Ocean phytoplankton to shifting light and nutrient limitation, *Limnol. Oceanogr.*, 63(3), 1–18, doi:10.1002/lno.11039, 2018.
- Lewis, W.: Global primary production of lakes: 19th Baldi Memorial Lecture, *Inl. Waters*, 1(1), 1–28, doi:10.5268/iw-1.1.384, 2011.
- 510 Linke, S., Lehner, B., Ouellet Dallaire, C., Ariwi, J., Grill, G., Anand, M., Beames, P., Burchard-Levine, V., Maxwell, S., Moidu, H., Tan, F. and Thieme, M.: Global hydro-environmental sub-basin and river reach characteristics at high spatial resolution, *Sci. Data*, 6(1), 1–15, doi:10.1038/s41597-019-0300-6, 2019.



- 515 LUBW water quality data, LUBW data map Serv. Available from: <https://udo.lubw.baden-wuerttemberg.de/public/index.xhtml>, 2020.
- Lyngsgaard, M. M., Markager, S., Richardson, K., Møller, E. F. and Jakobsen, H. H.: How Well Does Chlorophyll Explain the Seasonal Variation in Phytoplankton Activity?, *Estuaries and Coasts*, 40(5), 1263–1275, doi:10.1007/s12237-017-0215-4, 2017.
- 520 Maberly S.C.;Carter, H. T.;Clark. M. A.;D. V. M. M.;Fletche. J. M.;Jame. J. B.;Keena. P.;Kell. J. L.;Macka. E. B.;Parke. J. E.;Pate. M.;Pereir. M. G.;Rhode. G.;Tanna B.;Thackera. S. J.;Vincen. C.;Feuchtmay. H.: Surface temperature, surface oxygen, water clarity, water chemistry and phytoplankton chlorophyll a data from Bassenthwaite Lake, 1990 to 2013, doi:10.5285/91d763f2-978d-4891-b3c6-f41d29b45d55, 2017.
- 525 Maberly S.C.;Brierley, B.;Carte. H. T.;Clark. M. A.;D. V. M. M.;Fletche. J. M.;Jame. J. B.;Keena. P.;Kell. J. L.;Macka. E. B.;Parke. J. E.;Pate. M.;Pereir. M. G.;Rhode. G.;Tann. B.;Thackera. S. J.;Vincen. C. J.;Feuchtmay. H.: Surface temperature, surface oxygen, water clarity, water chemistry and phytoplankton chlorophyll a data from Windermere South Basin, 1945 to 2013, doi:10.5285/e3c4d368-215d-49b2-8e12-74c99c4c3a9d, 2017.
- 530 Maberly S.C.;Brierley, B.;Carte. H. T.;Clark. M. A.;D. V. M. M.;Fletche. J. M.;Jame. J. B.;Keena. P.;Kell. J. L.;Macka. E. B.;Parke. J. E.;Pate. M.;Pereir. M. G.;Rhode. G.;Tann. B.;Thackera. S. J.;Vincen. C. J.;Feuchtmay. H.: Surface temperature, surface oxygen, water clarity, water chemistry and phytoplankton chlorophyll a data from Windermere North Basin, 1945 to 2013, doi:10.5285/f385b60a-2a6b-432e-aadd-a9690415a0ca, 2017.
- 535 Dudley B. J.;May, L.;Spear. B. M.;Kirik. A.: Loch Leven long-term monitoring data: phosphorus, silica and chlorophyll concentrations, water clarity and temperature, 1985-2007, doi:10.5285/2969776d-0b59-4435-a746-da50b8fd62a3, 2013.
- 540 Maberly S.C.;Brierley, B.;Carte. H. T.;Clark. M. A.;D. V. M. M.;Fletche. J. M.;Jame. J. B.;Keena. P.;Kell. J. L.;Macka. E. B.;Parke. J. E.;Pate. M.;Pereir. M. G.;Rhode. G.;Tann. B.;Thackera. S. J.;Vincen. C.;Feuchtmay. H.: Surface temperature, surface oxygen, water clarity, water chemistry and phytoplankton chlorophyll a data from Grasmere, 1968 to 2013, doi:10.5285/b891c50a-1f77-48b2-9c41-7cc0e8993c50, 2017.
- 545 Maberly S.C.;Brierley, B.;Carte. H. T.;Clark. M. A.;D. V. M. M.;Fletche. J. M.;Jame. J. B.;Keena. P.;Kell. J. L.;Macka. E. B.;Parke. J. E.;Pate. M.;Pereir. M. G.;Rhode. G.;Tann. B.;Thackera. S. J.;Vincen. C.;Feuchtmay. H.: Surface temperature, surface oxygen, water clarity, water chemistry and phytoplankton chlorophyll a data from Esthwaite Water, 1945 to 2013, doi:10.5285/87360d1a-85d9-4a4e-b9ac-e315977a52d3, 2017.
- 550 Maberly S.C.;Carter, H. T.;Clark. M. A.;D. V. M. M.;Fletche. J. M.;Jame. J. B.;Keena. P.;Kell. J. L.;Macka. E. B.;Parke. J. E.;Pate. M.;Pereir. M. G.;Rhode. G.;Tanna B.;Thackera. S. J.;Vincen. C.;Feuchtmay. H.: Surface temperature, surface oxygen, water clarity, water chemistry and phytoplankton chlorophyll a data from Derwent Water, 1990 to 2013, doi:10.5285/106844ff-7b4c-45c3-8b4c-7cfb4a4b953b, 2017.
- 555 Maberly S.C.;Brierley, B.;Carte. H. T.;Clark. M. A.;D. V. M. M.;Fletche. J. M.;Jame. J. B.;Keena. P.;Kell. J. L.;Macka. E. B.;Parke. J. E.;Pate. M.;Pereir. M. G.;Rhode. G.;Tann. B.;Thackera. S. J.;Vincen. C.;Feuchtmay. H.: Surface temperature, surface oxygen, water clarity, water chemistry and phytoplankton chlorophyll a data from Blelham Tarn, 1945 to 2013, doi:10.5285/393a5946-8a22-4350-80f3-a60d753beb00, 2017.
- Magnuson, J., Carpenter, S. and Stanley, E.: North Temperate Lakes LTER: Chlorophyll - Madison Lakes



- Area 1995 - current, Environ. Data Initiat.,
doi:<https://doi.org/10.6073/pasta/f28e278afc34f1b7bd4f3cdc02b733a2>, 2019.
- 560 Magnuson, J., Carpenter, S. and Stanley, E.: North Temperate Lakes LTER: Chlorophyll - Trout Lake Area 1981 - current, Environ. Data Initiat.,
doi:<https://doi.org/10.6073/pasta/6c8ee65f6876a7274bfe7714ae7c3a70>, 2019.
- Markelov, I., Couture, R. M., Fischer, R., Haande, S. and Van Cappellen, P.: Coupling Water Column and Sediment Biogeochemical Dynamics: Modeling Internal Phosphorus Loading, Climate Change Responses, and Mitigation Measures in Lake Vansjø, Norway, *J. Geophys. Res. Biogeosciences*, 124(12), 3847–3866, doi:[10.1029/2019JG005254](https://doi.org/10.1029/2019JG005254), 2019.
- 565 McGlue, M. M., Ivory, S. J., Stone, J. R., Cohen, A. S., Kamulali, T. M., Latimer, J. C., Brannon, M. A., Kimirei, I. A. and Soreghan, M. J.: Solar irradiance and ENSO affect food security in Lake Tanganyika, a major African inland fishery, *Sci. Adv.*, 6(41), 1–9, doi:[10.1126/sciadv.abb2191](https://doi.org/10.1126/sciadv.abb2191), 2020.
- 570 Melkozernov, A. N. and Blankenship, R. E.: Photosynthetic Functions of Chlorophylls, in *Chlorophylls and Bacteriochlorophylls*, pp. 397–412, Springer Netherlands., 2007.
- Messenger, M. L., Lehner, B., Grill, G., Nedeva, I. and Schmitt, O.: Estimating the volume and age of water stored in global lakes using a geo-statistical approach, *Nat. Commun.*, 7, 1–11, doi:[10.1038/ncomms13603](https://doi.org/10.1038/ncomms13603), 2016.
- 575 O’Beirne, M. D., Werne, J. P., Hecky, R. E., Johnson, T. C., Katsev, S. and Reavie, E. D.: Anthropogenic climate change has altered primary productivity in Lake Superior, *Nat. Commun.*, 8, 15713, doi:[10.1038/ncomms15713](https://doi.org/10.1038/ncomms15713), 2017.
- O’Connell, D. W., Ansems, N., Kukkadapu, R. K., Jaisi, D., Orihel, D. M., Cade-Menun, B. J., Hu, Y., Wiklund, J., Hall, R. I., Chessell, H., Behrends, T. and Van Cappellen, P.: Changes in Sedimentary Phosphorus Burial Following Artificial Eutrophication of Lake 227, Experimental Lakes Area, Ontario, Canada, *J. Geophys. Res. Biogeosciences*, 125(8), doi:[10.1029/2020JG005713](https://doi.org/10.1029/2020JG005713), 2020.
- 580 Parks Canada: Clear Lake Water Quality - Riding Mountain, Open Gov. Available from: <https://open.canada.ca/data/en/dataset/2a55313f-26fc-4872-9a57-2a7bf2a4cc38>, 2017.
- Python Software Foundation: Python Language, Available from: <https://www.python.org/>, 2021.
- 585 QGIS Development Team: QGIS Geographic information system, Available from: <http://qgis.osgeo.org>, 2021.
- Reback, J., McKinney, W., jbrockmendel, Van den Bossche, J., Augspurger, T., Cloud, P., Gyoung, Sinhrks, Klein, A., Roeschke, M., Hawkins, S., Tratner, J., She, C., Ayd, W., Petersen, T., Garcia, M., Schendel, J. and Hayden, A.: pandas-dev/pandas: Pandas 1.0.3 (v1.0.3), Available from: <https://doi.org/10.5281/zenodo.3715232>, 2020.
- 590 Riederer, C.: Dplython, GitHub Repos. Available from: <https://github.com/dodger487/dplython>, 2015.
- Rigosi, A., Carey, C. C., Ibelings, B. W. and Brookes, J. D.: The interaction between climate warming and eutrophication to promote cyanobacteria is dependent on trophic state and varies among taxa, *Limnol. Oceanogr.*, 59(1), 99–114, doi:[10.4319/lo.2014.59.1.0099](https://doi.org/10.4319/lo.2014.59.1.0099), 2014.
- 595 Roche, D. G., Granados, M., Austin, C. C., Wilson, S. and Mitchell, G. M.: Open government data and environmental science : a federal Canadian perspective, (Appendix 1), 942–962, doi:[10.1139/facets-2020-0008](https://doi.org/10.1139/facets-2020-0008), 2020.
- Rosso, L., Lobry, J. R., Bajard, S. and Flandrois, J. P.: Convenient model to describe the combined effects



- of temperature and pH on microbial growth, *Am. Soc. Microbiol.*, (61), 610–616, doi:0099-2240, 1995.
- 600 Rousseaux, C. S. and Gregg, W. W.: Interannual variation in phytoplankton primary production at a global scale, *Remote Sens.*, 6(1), 1–19, doi:10.3390/rs6010001, 2013.
- Rudstam, L. G.: Limnological data and depth profile from Oneida Lake, New York, 1975 to present, *Knowl. Netw. Biocomplexity*, 2017.
- 605 Schindler, D. W.: A Hypothesis to Explain Differences and Similarities Among Lakes in the Experimental Lakes Area, Northwestern Ontario, *J. Fish. Res. Board Canada*, 28(2), 295–301, doi:10.1139/f71-039, 1971.
- Schwarz, M., Folini, D., Hakuba, M. Z. and Wild, M.: From Point to Area: Worldwide Assessment of the Representativeness of Monthly Surface Solar Radiation Records, *J. Geophys. Res. Atmos.*, 123(24), 13,857–13,874, doi:10.1029/2018JD029169, 2018.
- 610 Scofield, A. E., Watkins, J. M., Osantowski, E. and Rudstam, L. G.: Deep chlorophyll maxima across a trophic state gradient: A case study in the Laurentian Great Lakes, *Limnol. Oceanogr.*, 65(10), 2460–2484, doi:10.1002/lno.11464, 2020.
- Tett, P.: The ecophysiology of exceptional blooms, *Rapp. Proces-verbaux des Reun. Cons. Int. pour l'Exploration la Mer*, 3(187), 47–60, 1987.
- 615 Tian, D., Xie, G., Tian, J., Tseng, K. H., Shum, C. K., Lee, J. and Liang, S.: Spatiotemporal variability and environmental factors of harmful algal blooms (HABs) over western Lake Erie, *PLoS One*, 12(6), 1–16, doi:10.1371/journal.pone.0179622, 2017.
- 620 Tranvik, L. J., Downing, J. A., Cotner, J. B., Loiselle, S. A., Striegl, R. G., Ballatore, T. J., Dillon, P., Finlay, K., Fortino, K., Knoll, L. B., Kortelainen, P. L., Kutser, T., Larsen, S., Laurion, I., Leech, D. M., Leigh McCallister, S., McKnight, D. M., Melack, J. M., Overholt, E., Porter, J. A., Prairie, Y., Renwick, W. H., Roland, F., Sherman, B. S., Schindler, D. W., Sobek, S., Tremblay, A., Vanni, M. J., Verschoor, A. M., Von Wachenfeldt, E. and Weyhenmeyer, G. A.: Lakes and reservoirs as regulators of carbon cycling and climate, *Limnol. Oceanogr.*, 54(6 PART 2), 2298–2314, doi:10.4319/lno.2009.54.6_part_2.2298, 2009.
- 625 UK Environment Agency: All of England water quality data, Available from: <https://environment.data.gov.uk/water-quality/view/download>, 2020.
- United States Environmental Protection Agency: 2012 National Lakes Assessment. Field Operations Manual., Washington, DC., 2012.
- 630 Verpoorter, C., Kutser, T., Seekell, D. and Tranvik, L.: A global inventory of lakes based on high-resolution satellite imagery, *Geophys. Res. Lett.*, 41, 6396–6402, doi:10.1002/2014GL060641. Received, 2014.
- 635 Virtanen, P., Gommers, R., Oliphant, T. E., Haberland, M., Reddy, T., Cournapeau, D., Burovski, E., Peterson, P., Weckesser, W., Bright, J., van der Walt, S. J., Brett, M., Wilson, J., Millman, K. J., Mayorov, N., Nelson, A. R. J., Jones, E., Kern, R., Larson, E., Carey, C. J., Polat, İ., Feng, Y., Moore, E. W., VanderPlas, J., Laxalde, D., Perktold, J., Cimrman, R., Henriksen, I., Quintero, E. A., Harris, C. R., Archibald, A. M., Ribeiro, A. H., Pedregosa, F., van Mulbregt, P., Vijaykumar, A., Bardelli, A. Pietro, Rothberg, A., Hilboll, A., Kloeckner, A., Scopatz, A., Lee, A., Rokem, A., Woods, C. N., Fulton, C., Masson, C., Häggström, C., Fitzgerald, C., Nicholson, D. A., Hagen, D. R., Pasechnik, D. V., Olivetti, E., Martin, E., Wieser, E., Silva, F., Lenders, F., Wilhelm, F., Young, G., Price, G. A., Ingold, G. L., Allen, G. E., Lee, G. R., Audren, H., Probst, I., Dietrich, J. P., Silterra, J., Webber, J. T., Slavič, J., Nothman, J., Buchner, J., Kulick, J., Schönberger, J. L., de Miranda Cardoso, J. V., Reimer, J., Harrington, J.,



- 645 Rodríguez, J. L. C., Nunez-Iglesias, J., Kuczynski, J., Tritz, K., Thoma, M., Newville, M., Kümmerer, M., Bolingbroke, M., Tartre, M., Pak, M., Smith, N. J., Nowaczyk, N., Shebanov, N., Pavlyk, O., Brodtkorb, P. A., Lee, P., McGibbon, R. T., Feldbauer, R., Lewis, S., Tygier, S., Sievert, S., Vigna, S., Peterson, S., More, S., Pudlik, T., et al.: SciPy 1.0: fundamental algorithms for scientific computing in Python, *Nat. Methods*, 17(3), 261–272, doi:10.1038/s41592-019-0686-2, 2020.
- Water Information System Sweden (VISS): VISS database, Available from: <https://viss.lansstyrelsen.se/>, 2020.
- 650 Watson, S. B., Miller, C., Arhonditsis, G., Boyer, G. L., Carmichael, W., Charlton, M. N., Confesor, R., Depew, D. C., Höök, T. O., Ludsin, S. A., Matisoff, G., McElmurry, S. P., Murray, M. W., Peter Richards, R., Rao, Y. R., Steffen, M. M. and Wilhelm, S. W.: The re-eutrophication of Lake Erie: Harmful algal blooms and hypoxia, *Harmful Algae*, 56, 44–66, doi:10.1016/j.hal.2016.04.010, 2016.
- Wetzel, R. G.: *Limnology: Lake and River Ecosystems*, third edition, Academic press., 2001.
- 655 Wild, M.: Global dimming and brightening: A review, *J. Geophys. Res. Atmos.*, 114(12), 1–31, doi:10.1029/2008JD011470, 2009.
- Woolway, R. I. and Merchant, C. J.: Worldwide alteration of lake mixing regimes in response to climate change, *Nat. Geosci.* |, 12, 271–276, doi:10.1038/s41561-019-0322-x, 2019.

660

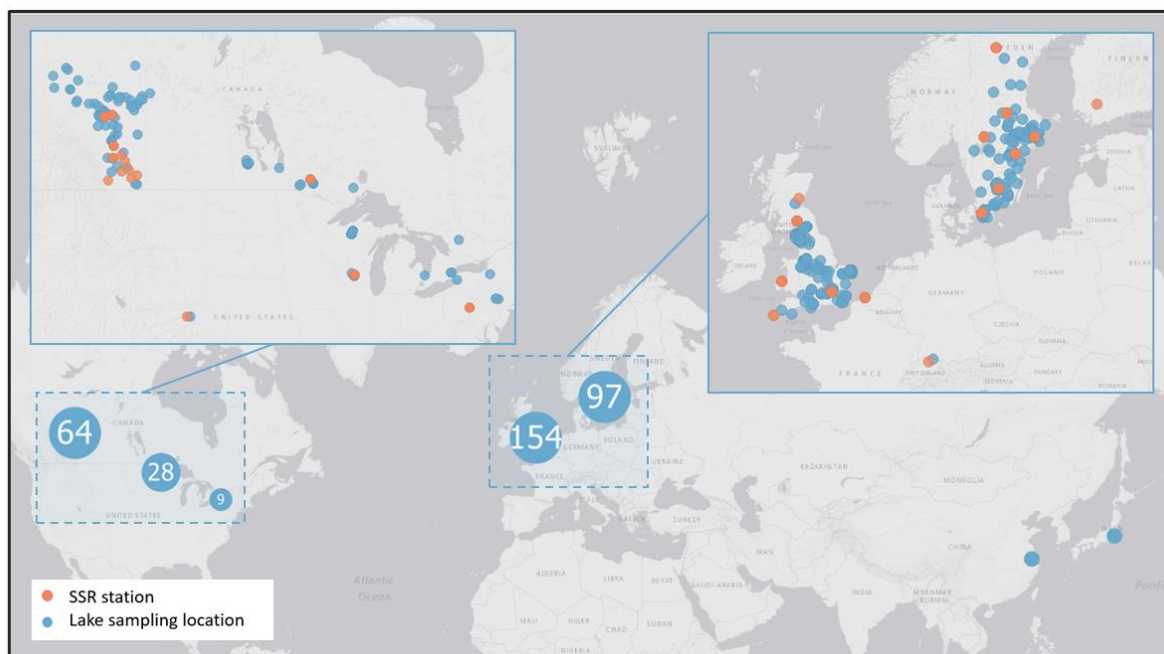
665

670



675 Table 1: Summary of variables in the derived growth window dataset.

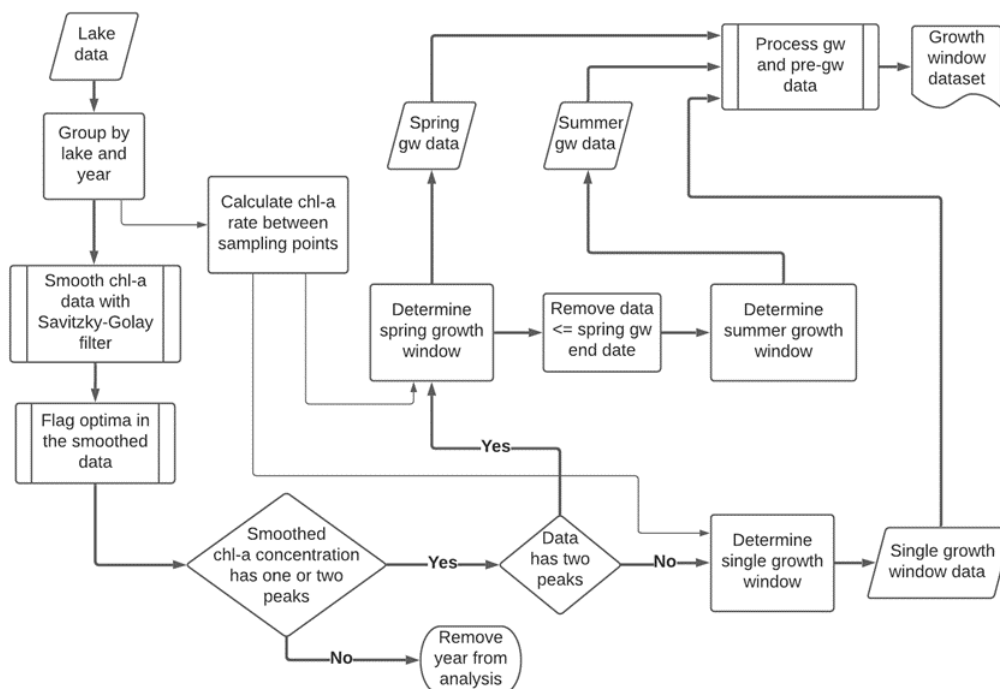
Variable	Units	Description	Comments
Season	NA	Time of year when the growth window is detected: Spring, Summer, or Single growth window	A single growth window occurs when there is no decrease in chlorophyll- <i>a</i> concentration between spring and summer
Chlorophyll-<i>a</i> growth rate	$\mu\text{g L}^{-1}\text{day}^{-1}$	Rate of increase in chlorophyll- <i>a</i> concentration between the start and end of the growth window	Chlorophyll- <i>a</i> rate of change between sampling times are used to define the growth window period
Specific chlorophyll-<i>a</i> rate	day^{-1}	Chlorophyll- <i>a</i> growth rate divided by initial concentration	Can be compared across lakes
Temperature-corrected specific chlorophyll-<i>a</i> rate	day^{-1}	Temperature correction function provided in supplementary data	Initial parameters: min temp=0°C, max temp=40°C, optimal temp=25°C
POC growth rate	$\text{mg L}^{-1}\text{day}^{-1}$	Rate of increase or decrease between consecutive sampling times	Representative metric for the rate of change in total algal biomass
Chlorophyll-<i>a</i> growth rate : POC growth rate	$\text{mg chlorophyll-}a : \text{mg POC}$	Ratio of the chlorophyll- <i>a</i> and POC rates of change	Can be used to see how the chlorophyll- <i>a</i> rate of production changes in proportion to total algal biomass
Mean surface water temperature	°C	Mean value across the growth window and the 14-day pre-growth window	
Surface solar radiation	Wm^{-2}	Mean value across the growth window and the 14-day pre-growth window	
TP	mg L^{-1}	Growth window mean values	(co-)limiting macronutrients
SRP	mg L^{-1}		
TKN	mg L^{-1}		
Secchi depth	m		Proxy for turbidity
pH	pH units		
Trophic Status Index (TSI)	Range: from 0-100	Calculated from the mean chlorophyll- <i>a</i> concentration across all years the lake was sampled	Used to assign trophic status
Trophic status	NA	Trophic status class assigned based on TSI: Oligotrophic, Mesotrophic, Eutrophic, or Hypereutrophic	TSI thresholds are those of the North American Lake Management Society



680 *Figure 1: Distribution of the 357 lake sampling locations in the growth window dataset. Sampling points are clustered by proximity, where marker size and value indicate the number of unique locations represented by each point. Enlarged sections show each lake sampling location and along with the location of the 322 paired SSR stations. Base map credit: ESRI, 2011.*

685

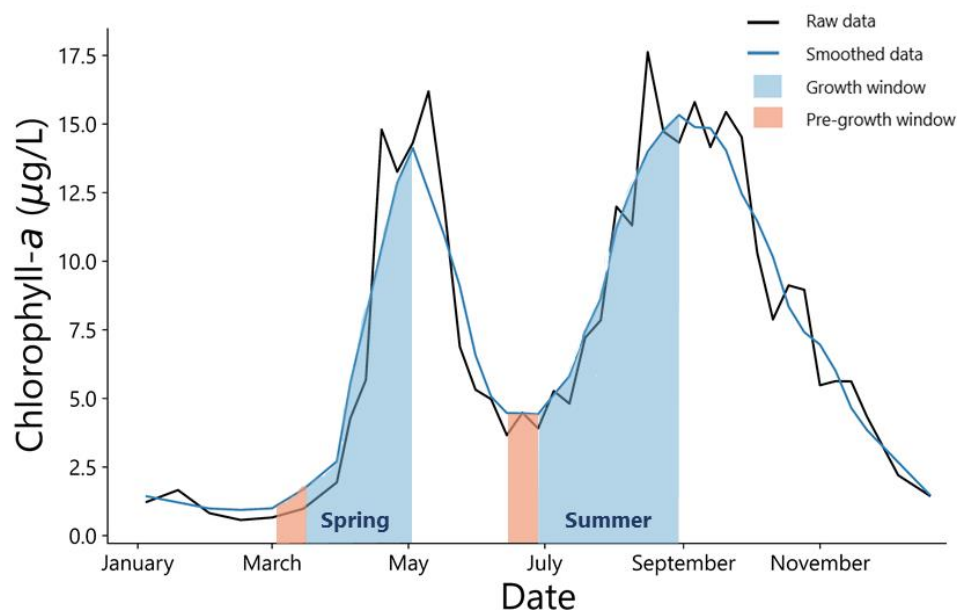
690



695 *Figure 2: Workflow for detecting and processing growth window data. For each lake sampling point, chlorophyll-a (Chl-a) data are smoothed with a Savitzky-Golay filter and then growth windows are detected based on peaks in the chlorophyll-a concentration. Growth windows are flagged as spring, summer, or single growth windows.*

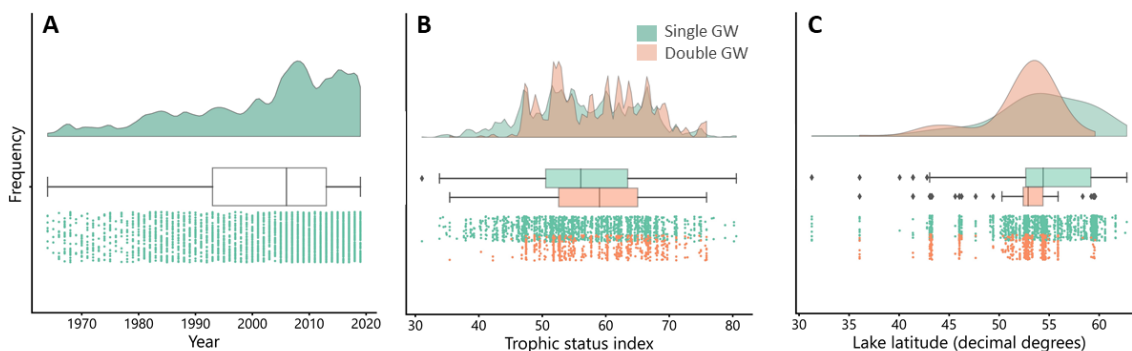
700

705



710 *Figure 3. Example of spring and summer growth windows in Lake Windermere’s north basin in 1988. Peaks in the smoothed data indicate the end of the growth window, and the window begins when the rate of increase in chlorophyll-a concentration surpasses a threshold of $0.05 \mu\text{gL}^{-1}\text{day}^{-1}$ (median rate for the distinct switch to a “rapid growth” period in mesotrophic-hypereutrophic lakes) for the first time. The growth window and pre-growth window (two weeks leading up to the growth window) are shown in blue and orange shading respectively.*

715



720 *Figure 4. Distributions of (a) year of occurrence, (b) lake trophic status index, and (c) lake latitude for each growth window in the dataset. Data are grouped by “double GW” or “single GW” year. The data is skewed toward more recent years and higher latitudes. Lakes in the oligotrophic category (TSI < 40) have the highest proportion of single growth windows.*

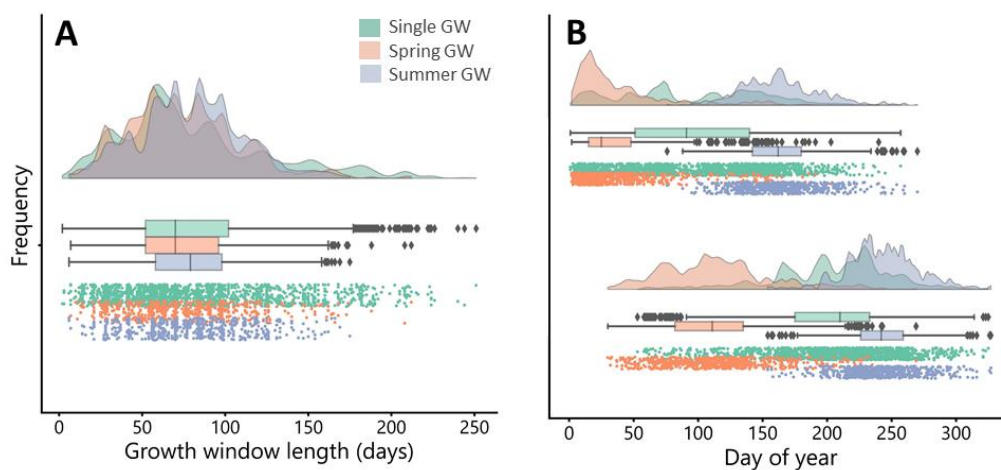
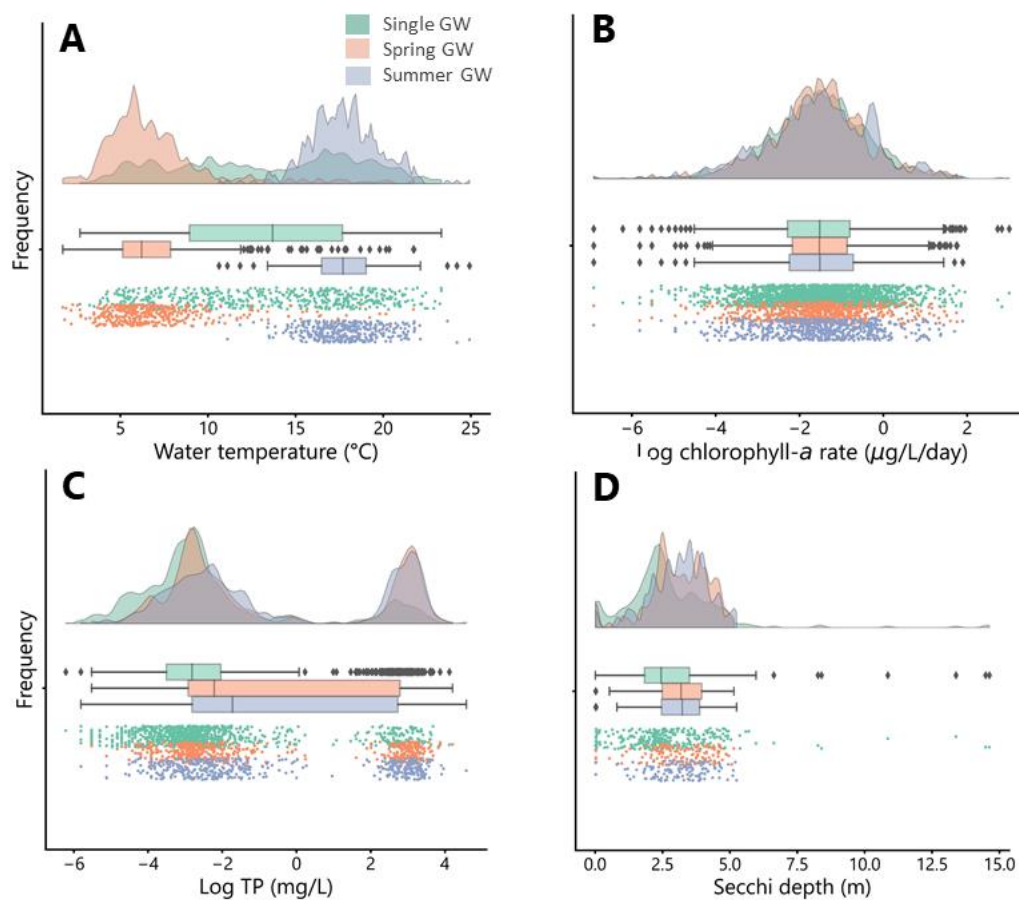


Figure 5. Distributions of (a) duration and (b) timing of the growth windows, grouped by growth window type. Single growth windows have both the longest range in length and the most even distribution of start and end dates.

725

730

735



740 Figure 6. Distributions of selected water quality variables during the growth window period: (a) log chlorophyll-a rate, (b) mean
745 water temperature, (c) log mean total phosphorus (TP), and (d) mean Secchi depth.

745

750

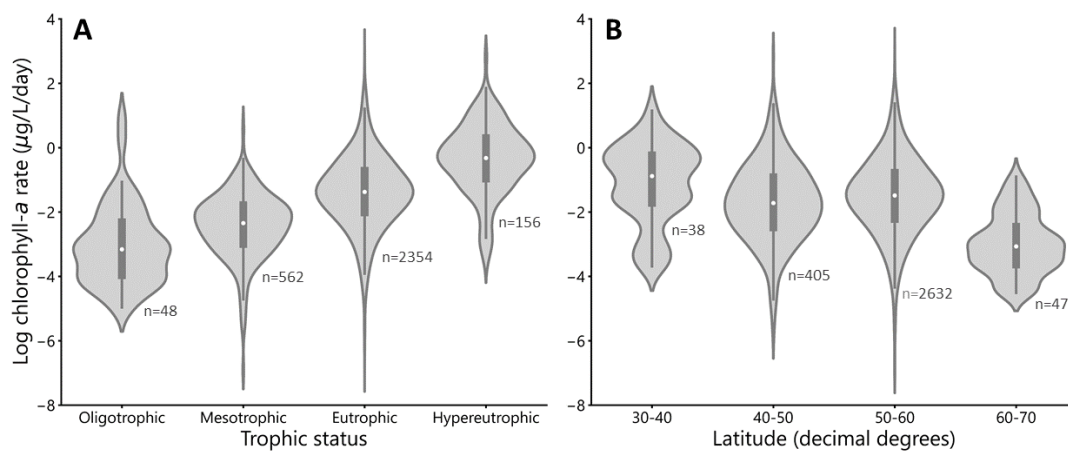


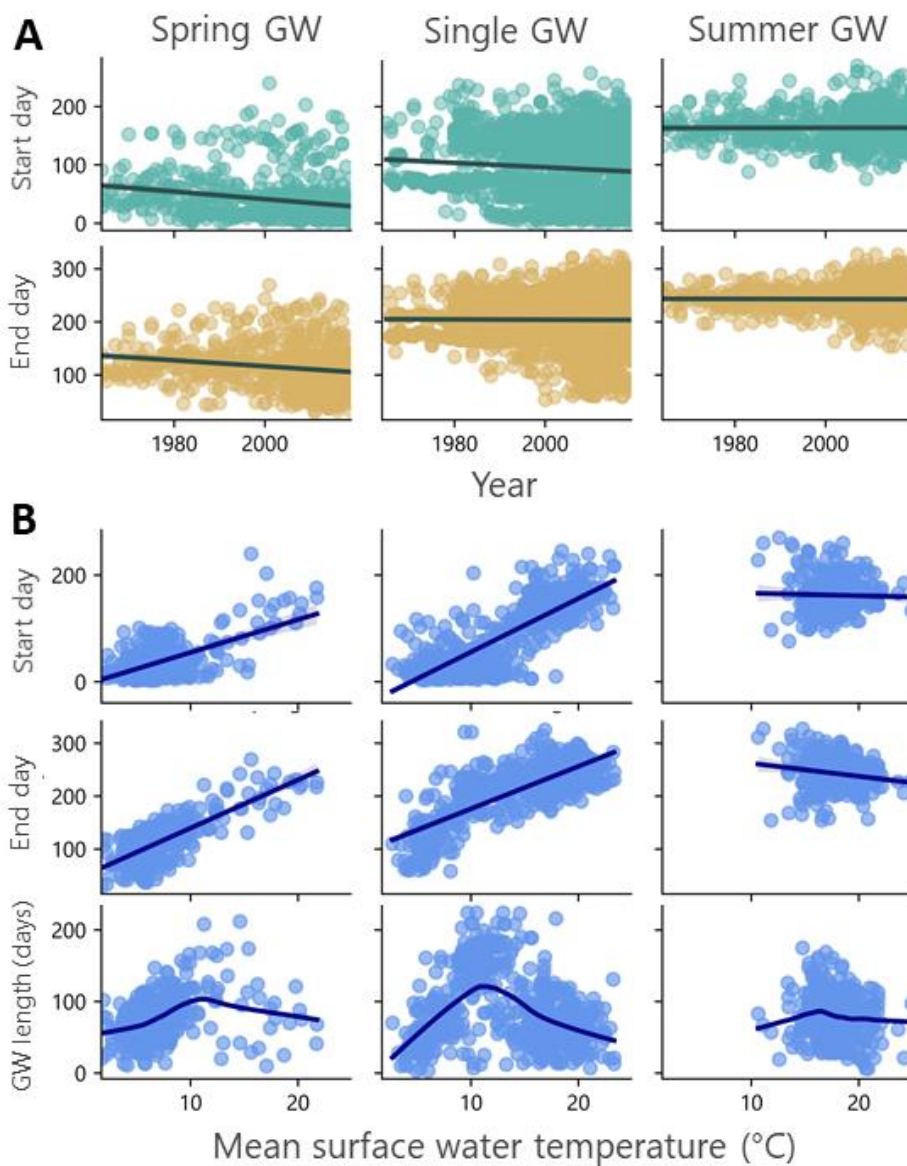
Figure 7. Chlorophyll-a growth rate trends in the dataset: grouped by (a) trophic status and (b) latitude. Lakes of a higher trophic status have higher mean chlorophyll-a growth rates and lakes at higher latitudes have lower chlorophyll-a growth rate during the growth windows. The number of lakes represented by each violin is shown in text on the panels.

755

760

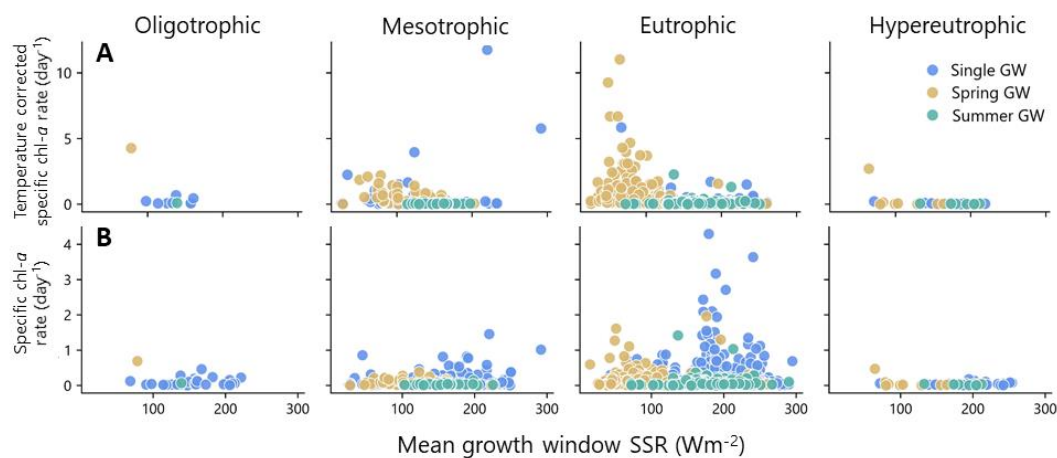
765

770



775

Figure 8. (a) time series of the start and end dates for the spring, summer, and single growth windows for all the lakes in the dataset; all growth window categories trend toward earlier start and end dates, especially in the spring. (b) Start and end dates of the growth windows as a function of temperature (regression line in dark blue) suggest a positive relationship between growth window timing and surface water temperature in the spring and a negative relationship in the summer. Growth window length (dark blue trendline shows locally weighted scatterplot smoothing) shows that longer growth windows occur at moderate surface water temperatures that aren't seen in the summer months.



780 *Figure 9. Comparison of trends in the relationship between mean growth window SSR with (a) temperature corrected*
chlorophyll-a rate and (b) specific chlorophyll-a rate without temperature correction. Data are grouped by trophic status, and
hue indicates growth window type. Lakes of a higher trophic status show an increased sensitivity to solar radiation, especially
during the spring (panel A) while summer growth windows do not show sensitivity to solar radiation or water temperature,
suggesting top-down control from zooplankton grazing. Low chlorophyll growth rates at SSR near or greater than 200 Wm⁻²
785 *indicate a photoacclimation response in the algae.*