A global map of root biomass across the world's forests

Yuanyuan Huang[1,2], Phillipe Ciais[1], Maurizio Santoro[3], David Makowski[4,5], Jerome Chave[6], Dmitry Schepaschenko[7,8,9], Rose Z. Abramoff[1], Daniel S. Goll[1], Hui Yang[1], Ye Chen[10], Wei Wei[11], Shilong Piao[12,13,14]

[1]Laboratoire des Sciences du Climat et de l'Environnement, LSCE/IPSL, CEA-CNRS-UVSQ, Université Paris-Saclay, 91191 Gif-sur-Yvette, France.

[2]Commonwealth Scientific and Industrial Research Organisation, Aspendale, 3195, Victoria, Australia.

[3]Gamma Remote Sensing, 3073 Gümligen, Switzerland.

[4]INRA, AgroParisTech, University Paris-Saclay, UMR 211, F-78850 Thiverval-Grignon, France.

[5]CIRED, 45bis Avenue de la Belle Gabrielle, 94130 Nogent-sur-Marne, France.

[6]Laboratoire Evolution et Diversite Biologique UMR 5174, CNRS, Universite Paul Sabatier, 118 route de Narbonne, Toulouse, 31062 France.

[7]International Institute for Applied Systems Analysis (IIASA) Schlossplatz 1, A-2361 Laxenburg, Austria.

[8]Center of Forest Ecology and Productivity of the Russian Academy of Sciences, Moscow 117997, Russia

[9]Siberian Federal University, Krasnoyarsk, 660041, Russia

[10]Department of Mathematics and Statistics, Northern Arizona University, 86001, Flagstaff, AZ, US.

[11]State Key Laboratory of Urban and Regional Ecology, Research Center for Eco-environmental Sciences, Chinese Academy of Sciences, Beijing, 100085, China.

[12]Sino-French Institute for Earth System Science, College of Urban and Environmental Sciences, Peking University, Beijing, China

[13]Key Laboratory of Alpine Ecology and Biodiversity, Institute of Tibetan Plateau Research, Chinese Academy of Sciences, Beijing, China

[14]Center for Excellence in Tibetan Earth Science, Chinese Academy of Sciences, Beijing, China

Correspondence to: Yuanyuan Huang (yuanyuanhuang2011@gmail.com)

Abstract. As a key component of the Earth system, ~~root plays the~~roots play a key role in linking Earth's lithosphere, hydrosphere, biosphere, and atmosphere. Here we combine 10307 field measurements of forest root biomass worldwide with global observations of forest structure, climatic conditions, topography, land management and soil characteristics to derive a spatially explicit global high-resolution (~ 1km) root biomass dataset, including fine and coarse roots. In total, 142 ± 25 (95% CI) Pg of live dry matter biomass is stored below-ground, representing a global average root:shoot biomass ratio of 0.25 ± 0.10. Our estimations of total root biomass in tropical, temperate and boreal forests are 44-226% smaller than earlier studies (Jackson et al., 1997; Robinson, 2007; Saugier et al., 2001)~~.~~, and the total global forest root biomass is 32% smaller than a recent estimate (Spawn et al., 2020). The smaller estimation is attributable to the updated forest area, spatially explicit above-ground biomass density used to predict the patterns of root biomass, new root measurements and upscaling methodology. We show specifically that the root shoot allometry is one underlying driver that leads to methodological overestimation of root biomass in previous estimations. Raw datasets and global maps generated in this study are deposited at the open access repository Figshare (https://figshare.com/articles/Supporting_data_and_code_for_A_global_map_of_root_biomass_across_the_world_s_forests/ 12199637)

1 Introduction

Roots act as a hub that connects complex feedbacks among biomes, soil, water, air, rocks and nutrients. Roots mediate nutrient and water uptake by plants, below-ground organic carbon decomposition, the flow of carbohydrates to mycorrhizae, species competition, soil stabilization and plant resistance to windfall(Warren et al., 2015). The global distribution of root biomass is related to how much photosynthates plants must invest below-ground to obtain water, nitrogen and phosphorus for sustaining photosynthesis, leaf area and growth. Root biomass and activity also control the land surface energy budget through plant transpiration(Wang et al., 2016; Warren et al., 2015). While Earth Observation data combined with field data enables the derivation of spatially explicit estimates of above-ground biomass with a spatial resolution of up to 30 meters over the whole globe(GlobalForestWatch, 2019; Santoro, 2018b), the global carbon stock and spatial details of the distribution of below-ground root biomass (fine + coarse) rely on sparse measurements and coarse extrapolation so far, therefore remaining highly uncertain.

More than twenty years ago, Jackson et al, 1996, 1997 [1,2] provided estimates of the average biomass density (weight per unit area) and vertical distribution of roots for 10 terrestrial biomes. Multiplying their average root biomass density with the area of each biome results in a global root biomass pool of 292 Pg, with forests accounting for ~68%. Saugier, et al. (2001) estimated global root biomass to be 320 Pg by multiplying biome-average root to shoot ratios (*R:S*) by shoot biomass density and the land area of each biome. Mokany, et al. (2006) argued that the use of mean *R:S* values at the biome scale is a source of error because root biomass measurements are performed at small scales, but root distributions are highly spatially heterogeneous and their size distribution spans several orders of magnitude, fine roots being particularly difficult to

sample(Jackson et al., 1996; Taylor et al., 2013). With updated *R:S* and broader vegetation classes, Mokany, et al. (2006) gave a higher global root biomass of 482 Pg. Robinson (2007) further suggested that *R:S* was underestimated by 60%, which translated into an even higher global root biomass of 540-560 Pg. These studies provided a first order estimation of the root biomass for different biomes, but not of its spatial details. Further, it is worth noting that estimations of the global total root biomass have increased with time., likely associated with improved methods in excavating roots that reduce under-sampling.

An alternative approach to estimate root biomass is through allometric scaling, dating back to West, Brown and Enquist (1997, 1999)[6][7] and Enquist and Niklas (2002). The allometric scaling theory assumes that biological attributes scale with body mass, and in the case of roots, an allometric equation verified by data takes the form of $R \propto S^{\beta}$ where R is the root mass, S the shoot mass and β a scaling exponent. In contrast to the studies listed above assuming the *R:S* ratio to be uniform, this equation implies that the *R:S* ratio varies with shoot size when β is not equal to one (Cairns et al., 1997; Enquist and Niklas, 2002; Jiang and Wang, 2017; McCarthy and Enquist, 2007; Niklas, 2005; Robinson, 2004; Zens and Webb, 2002) . Allometric equations also predict that smaller trees generally have a larger *R:S* with $\beta < 1$, which is well supported by measurement of trees of different sizes (Cairns et al., 1997; Jiang and Wang, 2017; Niklas, 2005; Robinson, 2004; Zens and Webb, 2002). The allometric equation approach was applied for various forest types, and the scaling exponent β was observed to differ across sites(Yunjian Luo et al., 2018), species(Cheng and Niklas, 2007), age(Cairns et al., 1997), leaf characteristics(Luo et al., 2012), elevation(Moser et al., 2011), management status(Ledo et al., 2018), climatic conditions, such as temperature(Reich et al., 2014), soil moisture and climatic water deficit (Ledo et al., 2018), as well as soil nutrient content and texture(Jiang and Wang, 2017). Despite successful application of allometric equations for site- and species-specific studies(Yunjian Luo et al., 2018), their use to predict global root biomass patterns appears to be limited and challenging.

2 Methods

2.1 Overview

We use a new approach to upscale root biomass of trees at the global scale (Supplementary Figure 1) based on machine learning algorithms trained by a large dataset of *in-situ* measurements (n=10307) [14,30,31]of root and shoot biomass for individual woody plants (see Methods: Field measurement, Supplementary Data), covering 465 species across 10 biomes defined by The Nature Conservancy(Olson and Dinerstein, 2002) (Supplementary Figure 2). We compared the allometric upscaling and tested three machine learning techniques (the random forest, the artificial neural networks and multiple adaptive regression splines), searched through a pool of 47 predictor variables that include shoot biomass and other vegetation, edaphic, topographic, anthropogenic and climatic variables (Supplementary Table 1)). After comparing the results of all three machine learning techniques and selectedthe allometric upscaling, we chose the random forest model (RF) that performsbecause it performed best on cross validation samples (see section Building predictive models below). and Supplementary Table 8), and we only used the RF for subsequent mapping and analysis. Using this RF model, we mapped the root biomass of an average tree over an area of ~1km x1 km across the globe using as predictors gridded maps of shoot biomass (weight per area) (Santoro, 2018a), tree height(Simard et al., 2011), soil nitrogen(Shangguan et al., 2014),

pH(Shangguan et al., 2014), bulk density(Shangguan et al., 2014), clay content(Shangguan et al., 2014), sand content(Shangguan et al., 2014), base saturation(Shangguan et al., 2014), cation exchange capacity(Shangguan et al., 2014), water vapor pressure(Fick and Hijmans, 2017), mean annual precipitation(Fick and Hijmans, 2017), mean annual temperature(Fick and Hijmans, 2017), aridity(Trabucco and Zomer, 2019) and water table depth(Fan et al., 2013) (Supplementary Figures 10,11,12). Combining our map of root biomass per tree with the tree density (number of trees per area)(Crowther et al., 2015) at the global scale, we quantified the global forest root biomass.

2.2 Field measurements

Our dataset (available at our Figshare repository) was compiled from literature and existing forest biomass structure or allometry databases(Falster et al., 2015) (Ledo et al., 2018; Schepaschenko et al., 2018; Schepaschenko et al., 2017). We included studies and databases that reported georeferenced location, root biomass and shoot biomass. Note here we focus on studies that measured biomass of individual trees instead of stand level biomass. We included studies that measured biomass directly, instead of estimated indirectly via allometric equations. For example, Ref(Poorter et al., 2015) is not included due to lack of georeferenced location and Ref(Iversen et al., 2017) in not used as we also need measurements of other plant compartments like shoot biomass. Repeated entries from existing databases were removed. One of the databases(Falster et al., 2015) reported data on woody plants which also include shrub species. We kept the shrub data partly because the remote sensing products we used to generate our root map do not clearly separate trees from shrubs. Around 82% of the extracted entries also recorded plant height and management status. Height was identified as an important predictor in our model assessment, and entries were discarded when height was missing (18% of data). As woody plant age was reported in 19% of the entries only, the values of this variable was determined from another source of information, i.e. from a composite global map introduced in the next section. Species names were systematically reported, but biotic, climatic, topographic and soil information were missing for a substantial proportion of entries and values of these variables were thus extracted from independent observation-driven global maps as explained in the next section. Our final dataset includes biomass measurements collected in 494 different locations from 10307 individual plants, which cover 465 species across 10 biomes as defined by The Nature Conservancy(Olson and Dinerstein, 2002) (Supplementary Figure 2; Supplementary Data).

2.3 Preparing predictor variables

We used 47 predictors that broadly cover 5 categories: vegetative, edaphic, climatic, topographic and anthropogenic (Supplementary Table 1). These variables are chosen due to their relevance to root dynamics based on field studies and their availability at the global scale. Vegetative variables include shoot biomass, height, age, maximum rooting depth, biome class and species. Edaphic predictors cover soil bulk density, organic carbon, pH, sand content, clay content, total nitrogen, total phosphorus, Bray phosphorus, total potassium, exchangeable aluminium, cation exchange capacity, base saturation (BS), soil moisture and water table depth (WT). Climatic predictors are mean annual temperature (MAT), mean annual precipitation (MAP), the aridity index that represents the ratio between precipitation and the reference evapotranspiration, solar radiation, potential evapotranspiration (PET), vapor pressure, cumulative water deficit (CWD=PET - MAP), wind speed, and mean diurnal range of temperature (BIO2, BIO is the abbreviation for WorldClim bioclimatic indicators), isothermality

(BIO2/BIO7) (BIO3), temperature seasonality (BIO4), max temperature of warmest month (BIO5), min temperature of coldest month (BIO6), temperature annual range (BIO7), mean temperature of wettest quarter (BIO8), mean temperature of driest quarter (BIO9), mean temperature of warmest quarter (BIO10), mean temperature of coldest quarter (BIO11), precipitation of wettest month (BIO13), precipitation of driest month (BIO14), precipitation seasonality (BIO15),

135 precipitation of wettest quarter (BIO16), precipitation of driest quarter (BIO17), precipitation of warmest quarter (BIO18), precipitation of coldest quarter (BIO19). The topographic variable is elevation. We did not have the slope and weaspect as topographic factors after a preliminary test which showed minor effect of elevation from our database. We take the management status (managed or not) as the anthropogenic predictor. All references are given in Supplementary Table 1. For each variable, we collected multiple datasets whenever possible only for uncertainty quantification. For the "central"

140 estimate, we stick to only one set of these predictors considering the quality and coverage of the databases. We favoured the database that is known to have higher quality data or covered more predictors that are relevant to our study for consistency.

As *in-situ* field measurements of above-ground biomass (AGB) do not offer a full global coverage, gridded shoot biomass data were derived from satellite AGB products to predict root biomass at the global scale with a 1km by 1 km spatial resolution. The gridded global shoot biomass dataset used in our study has been extensively calibrated with *in-situ*

145 observations and is currently the most reliable source of information on shoot biomass offering a global coverage (Baccini et al., 2017; Santoro, 2018a).(Santoro, 2018a; Spawn et al., 2020). To derive the shoot or AGB per tree (in unit of weight per tree) to generate spatially explicit global root biomass, we combined the GlobBiomass-AGB satellite data product(Santoro, 2018a) (in unit of weight per unit area) with a tree density map (number of trees per unit area)(Crowther et al., 2015). The GlobBiomass dataset was based on multiple remote sensing products (radar, optical, LiDAR) and a large pool of *in-situ*

150 observations of forest variables(Santoro et al., 2015; Santoro, 2018b). The original GlobBiomass-AGB map was generated at 100 m spatial resolution; for this study, the map was averaged into a 1 km pixel by considering only those pixels that were labeled as forest (Santoro, 2018b). A pixel was labeled as forest when the tree canopy density was larger than 15% according to Hansen et al. (2013)'s dataset (hereafter Hansen2013) averaged at 100 m. The 1-km resolution global tree density map was constructed through upscaling 429,775 ground-based tree density measurements with a predictive regression model for

155 forests in each biome(Crowther et al., 2015). The forest canopy height map took advantage of the Geoscience Laser Altimeter System (GLAS) aboard ICESat (Ice, Cloud, and land Elevation Satellite).) (Simard et al., 2011). Forest definitions are slightly different among these three maps. Forest area of the tree density map was based on a global consensus land cover dataset that merged four land cover products (Tuanmu and Jetz, 2014), and which gave a total tree count equal to the Hansen et al. (2013) land cover product (Crowther et al., 2015). The canopy height map used the Globcover land cover

160 map. Crowther et al. (2015) showed the total tree count from tree density map based on the Tuanmu and Jetz (2014) land cover is the same as from the Hansen2013 land cover product. The canopy height map used the Globcover land cover map (Hagolle et al., 2005) as reference to define forest land. We took Hansen2013 with a 15% canopy cover threshold as our base forest cover map. We approximated the missing values in tree density and height (due to mismatches in forest cover) by the mean of a 5x5 window that is centered on the corresponding pixel. We quantified the potential impact of mismatches in

165 forest definition by looking into two different thresholds: 0% and 30%.

We merged several regional age maps to generate a global forest age map. The base age map was derived from biomass through age-biomass curve similarly as conducted in tropical regions in ref.(Poulter, 2019)~~.~~. This age map does not cover the northern region beyond 35 N. We filled the missing northern region with a North American age map (Pan et al., 2011) and a second age map covering China_(Zhang et al., 2017)~~.~~. The North American age map represents the year around

170 2003 and as a coarse approximation, we added 7 to represent the year 2010. The China age map was derived for the period 2009-2013. We took it to approximate age values in China around 2010. Remaining missing pixels were further filled with the age map derived from MODIS disturbance observations. For the final step, we filled the remaining pixels with the GFAD V1.1 age map(Poulter, 2019). GFAD V1.1 has 15 age classes and 4 plant functional types (PFTs). ~~We~~GFAD V1.1 represents the 2000-2010 era. As a coarse approximation, we choose the middle value of each age class and estimated the age as the

175 average among different PFTs.

Detailed information of all ancillary variables is listed in Supplementary Table 1. To stay coherent, we re-gridded each map to a common 1 km x 1 km grid through the nearest neighbourhood method.

2.4 Building predictive models

We investigated the performance of the allometric scaling and three non-parametric models: ~~RF, ANN and MARS.~~

180 ~~Allometric upscaling relates root biomass to shoot biomass in the form of $R \propto S^{\beta}$.~~the random forest (RF), the artificial neural networks (ANN) and multiple adaptive regression splines (MARS). Allometric upscaling relates root biomass to shoot biomass in the form of $R \propto S^{\beta}$. We assume that one universal allometric equation governs (Enquist and Niklas, 2002) biomass partitioning across species without stratifying the data by species. RF is an ensemble machine learning method that builds a number of decision trees through training samples(Breiman, 2001). A decision tree is a flow-chart-like structure,

185 where each internal (non-leaf) node denotes a binary test on a predictor variable, each branch represents the outcome of a test, and each leaf (or terminal) node holds a predicted target variable. With a combination of learning trees (models), RF generally increases the overall prediction performance and reduces over-fitting. ANN computes through an interconnected group of nodes, inspired by a simplification of neurons in a brain. MARS is a non-parametric regression method that builds multiple linear regression models across a range of predictors.

190 Tree shoot biomass from the *in-situ* observation data spans a wider range than shoot biomass per plant derived from global maps (1×10^{-7} to 8800 vs. 7.9×10^{-5} to 933 kg/plant). To reduce potential mapping errors, we selected training samples with shoot biomass between 5×10^{-5} and 1000 kg/plant. The medians and means of shoot biomass, root biomass and $R{:}S$ from the selected training samples are similar to those from the entire database. Also, to reduce the potential impact of outliers, we analyzed samples with $R{:}S$ falling between the 1st and 99th percentiles, which consists of 9589 samples with $R{:}S$ ranging

195 from 0.05 to 2.47 and a mean of 0.47 and a median of 0.36. Sample filtering slightly deteriorated model performance and had minor impact on the final global root biomass prediction (145 from whole samples vs.142 Pg from filtered data). We chose root biomass as our target variable instead of $R{:}S$ because big and small trees contribute equally to $R{:}S$ while big trees are relatively more important in biomass quantification. In our observation database, we have more samples being small

woody plants (Supplementary Figure 6). ~~A model with an overall good performance will not guarantee a good prediction on woody plants with higher biomass.~~ We furthermore split the *in-situ* measured shoot biomass into three groups, namely measurements with shoot biomass smaller than 0.1, between 0.1 and 10, and larger than 10 kg/plant, and trained a specific model for each class. The rationale behind this splitting is: (1), to remove the bias of small plants from the distribution of *in-situ* measured woody shoot biomass (Supplementary Figure 6); (2), to account for the shift of root shoot allometry with tree size(Poorter et al., 2015) (Ledo et al., 2018; Zens and Webb, 2002); (3), to improve the performance of independent validation through numerous combinations of splitting trials; (4), and because tests through weighting samples or resampling samples (e.g., over-sampling using Synthetic Minority Over-sampling Technique) showed no better performance.

Model performances were assessed by 4-fold cross-validation using two criteria: the mean absolute error (MAE) and the R-squared value (R^2). MAE quantifies the overall error while R^2 estimates the proportion of variance in root biomass that is captured by the predictive model. We favoured the model with a smallest MAE, and a highest R^2. For models with comparable MAE and R^2, we favoured the model with the minimum number of predictors. For non-parametric models, starting from a model with all 47 predictors, we sequentially excluded predictors that did not improve model performance one after another. The order of removing predictors was random. After a combination of trials, the best model was from RF (Supplementary Table 8) and the final set of predictors included shoot biomass, height, soil nitrogen, pH, bulk density, clay content, sand content, base saturation, cation exchange capacity, vapor pressure, mean annual precipitation, mean annual temperature, aridity and water table depth. Note the maximum rooting depth had a minor impact on model performance and was not selected in the final model. The depth to which roots inhabit varies among species and environment. Our model predictions are therefore not specific to a certain soil depth.

2.5 Generation of the global root biomass map

Over an area of 1km x 1km, we assumed a tree with an average shoot biomass follows the RF model trained above. Building upon a large set of samples with each field measurement being an outcome of complex local interactions (including within-vegetation competition), we implicitly accounted for some sub-pixel variability (e.g., resource competition and responses to environmental conditions) on root biomass. ~~We combined the RF model with global maps of selected predictor variables to produce the root biomass map which has a unit of weight per tree. This map was multiplied by tree density at 1-km resolution to obtain the final root biomass map with a unit of weight per area (Supplementary Figure 1).~~ Our RF model was built upon individual woody trees. We combined the RF model with global maps of selected predictor variables to produce the map of root biomass for an average woody tree which has a unit of weight per tree. This map was multiplied by tree density (number of woody trees per area) (Crowther et al., 2015) at 1-km resolution to obtain the final root biomass map with a unit of weight per area (Supplementary Figure 1).

2.6 Uncertainty quantification

We estimated the overall uncertainty of the root biomass estimates through quantifying errors caused by predicting root biomass at the 1-km resolution (η_{pred}) and converting root biomass per tree to root biomass per unit area (η_{con}). We quantified the prediction uncertainty through an ensemble of predictions. We collected 8 additional global predictor datasets

(3 shoot biomass, 2 soil and 3 climate datasets) (Supplementary Table 2) and carried out 8x4 (4 folds) sets of additional predictions replacing the predictors by each of these additional data maps. We calculated the standard deviation among 36 predictions for each pixel (Supplementary Figure 4a). Converting root biomass from per tree to per area is through the tree density(Crowther et al., 2015). We assumed the coefficient of variation (CV, i.e., the ratio of the standard deviation to the mean) in tree density mapping caused the same relative uncertainty in our per unit area root biomass. CV in tree density mapping at the biome scale was derived from Ref(Crowther et al., 2015) through dividing uncertainties in quantifying total tree numbers by the total tree numbers. η_{con} in terms of standard deviation is therefore equal to the product of CV and the mean root biomass at each pixel (Supplementary Figure 4b). At last we propagated these two sources of uncertainty assuming these errors were random and independent. Note that we did not account for uncertainties in *in-situ* root biomass measurements used in training the RF model. The overall uncertainty (standard deviation) at the pixel level was calculated through,

$$\eta_{root} = \sqrt{\eta_{pred}^2 + \eta_{con}^2} \qquad (1)$$

At the biome and global scales, we obtained total root biomass for each of the 36 predictions and estimated the standard deviations of the total root biomass. η_{con} was estimated by multiplying CV by biome or global-level root biomass. We propagated these two sources of uncertainty through Equation 1. Note that the semivariogram of the random forest prediction errors do not show a clear autocorrelation pattern (Supplementary Figure 10).

2.7 Relative importance of predictor variables

The impact of predictors on predicting *R:S* was estimated through the Spearman's rank-order correlation at both the global and biome scales. We log-transformed the *R:S* and shoot biomass before standardizing these datasets. Partial dependence plots(Hastie et al., 2009) show the marginal effect that one predictor has on root biomass from a machine learning model, and serves as a supplement to the Spearman correlation.

3 Results

We estimated a global total root biomass of 142±25 (95% CI) Pg (see Methods for uncertainty estimation and Supplementary Figures 3, 4) for forests when forest is defined as all areas with tree cover larger than 15% from the Hansen et al. (2013) tree cover map. Note here we reported values in unit of Pg of dry biomass instead of Pg C. The corresponding global weighted mean *R:S* is 0.25 ± 0.10. The root biomass spatial distribution generally follows the pattern of shoot biomass, but there are significant local and regional deviations as shown by Figure 1. 51% of the global tree root biomass comes from tropical moist forest, 14% from boreal forest, 12% from temperate broadleaf forest and 10% from woody plants in tropical and subtropical grasslands, savanna and shrublands (Supplementary Table 3). Given our use of a tree cover threshold of 15% at 30m100m resolution, our estimate ignores the roots of isolated woody plants present in arid or cold regions (Staver et al., 2011), as well as heterogeneous (e.g. urban or agriculture) landscapes and is possibly an under-estimate. Total root biomass decreases from 151 to 134 Pg when the canopy cover threshold used to define forest land is

increased from 0% to 30%. The root biomass density per unit of forest area is highest in tropical moist forest, followed by temperate coniferous and Mediterranean forest (Figure 1, Supplementary Table 3). Cross validation showed a good match between predictions from our RF model and *in-situ* observations (Figures 2e, all data; Supplementary Figure 7, for each biome; Supplementary Figure 8, for three tree size classes; Supplementary Figure 9, for each continent), with an overall coefficient of determination R^2 of 0.85, a mean absolute error (MAE) of 2.18 kg and a median *R:S* similar to validation samples (0.35 from *in-situ* observation vs. 0.38 from prediction).0.38 from prediction). RF shows better performance than the other two machine learning algorithms and the allometric fitting, as shown in Supplementary Table 8. By continents, the performance of RF is worst in Africa (R^2 = 0.6; MAE = 44 kg) partly due to limited observations (Supplementary Figure 9). Root biomass of tropical, temperate and boreal forests together is 44-226% lower compared to earlier studies (Table 1, Supplementary Table 5, see Supplementary Information Comparison with published results).

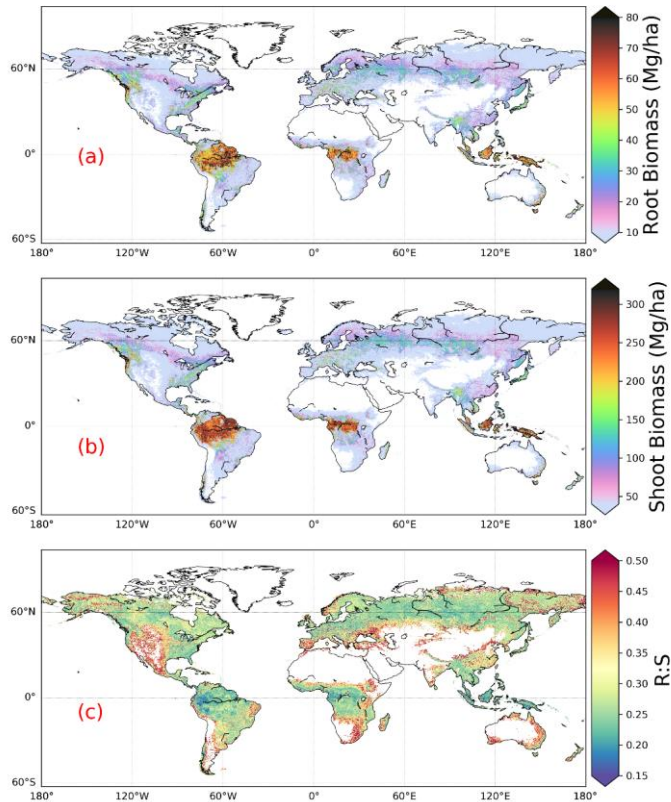



Figure 1. Global maps of forest root biomass generated through the random forest model (a), shoot biomass from GlobBiomass-AGB(Santoro, 2018b) (b) and *R:S* (c). Forest is defined as an area with canopy cover > 15% from the Hansen et al. (2013) tree cover map.

280

We then analysed the dominant factors explaining spatial variations of root biomass and *R:S* (see Methods). Broadly speaking, locations with small trees, low precipitation, strong aridity, deep water table depth, high acidity, low bulk density, low base saturation and low cation exchange capacity are more likely to have higher fractional root biomass (Figure 3). In line with the allometric theory, shoot biomass emerged as the most important predictor of *R:S* and root biomass, as

285 given by the Spearman correlation analysis shown in Figure 3, and partial importance plots (Supplementary Figures 11, 12,

13). Water related variables (precipitation, water table depth, aridity and vapor pressure) also emerged as important predictors in explaining *R:S* patterns (Figure 3)(Ledo et al., 2018), with trees and woody plants in dry regions generally having higher *R:S* (Supplementary Tables 3, 4), and with stronger dependence on precipitation especially when precipitation is low and on water table depth when the water table is deep. Temperature is slightly negatively correlated with *R:S* at the global scale, in line with Reich et al. (2014). However, the relationship between temperature and below-ground biomass is not consistent among biomes (Figure 3) and biomass size groups (Supplementary Figures 11, 12, 13).

The relationship between total soil nitrogen and root biomass is negative when soil nitrogen content is below 0.1-0.2 % (Supplementary Figure 11, 12, 13). Root biomass and *R:S* generally increases with soil alkalinity (Figure 3, Supplementary Figures 11, 12, 13). Low pH is toxic to biological activities and roots, especially as fine roots are sensitive to soil acidification, as revealed by a recent meta-analysis(Cheng Meng et al., 2019). Our results also indicate overall positive correlations between CEC, BS and R:S, but the processes that may account for these correlations are less clear from literature. Age has been shown to be important for *R:S*(Schepaschenko et al., 2018). How age regulates *R:S* remains elusive, with studies showing both positive(Waring and Powers, 2017) and slightly negative(Mokany et al., 2006) relationship between *R:S* and age. Including forest age (see Methods: Preparing predictor variables) as a predictor only marginally improved our model prediction (see Supplementary Information for details). It is likely that shoot biomass partially accounts for age information and the quality of the global forest age data might also affect the power of this variable in improving root biomass predictions.

Table 1. Comparison between studies quantifying root biomass in tropical, temperate and boreal forests.

	This study[S1]	This study[S2]	Jackson(Jackson et al., 1997)	Saugier(Saugier et al., 2001)	Robinson(Robinson, 2007)	Spawn (Spawn et al., 2020)	This study [S3]
Method	Machine learning	Machine learning	Biome average root biomass density	Biome average *R:S*, shoot biomass density	Biome average *R:S*, shoot biomass density	Relationships between root/total biomass and temperature	Allometric equations
Tropical (Tr, Pg)	92	76	114	147	246		
Temperate(Te, Pg)	26	25	51	59	98		
Boreal (Bo, Pg)	21	20	35	30	50		
Tr + Te + Bo (Pg)	139	121	200	236	394		
Globe (Pg)	142	142				188[#]	155-210
RD$_{S1}$[*]	0%		44%	70%	183%	32%	
RD$_{S2}$[&]		0%	65%	95%	226%	32%	

S1, Tropical moist forest (Biome 1), tropical dry forest (Biome 6), tropical/subtropical coniferous forest (Biome 11) and forest in tropical/subtropical grasslands/savannas and shrublands (Biome 3) are aggregated to represent tropical systems (Tr). Temperate broadleaf/mixed forest (Biome 4), temperate coniferous forest (Biome 5) and forest in temperate grasslands/savannas and shrublands (Biome 8) are merged together as temperate systems (Te). Boreal forest (Biome 2) and woody plants in tundra region (Biome 7) are aggregated as boreal forest (Bo). Biome classification is from The Nature

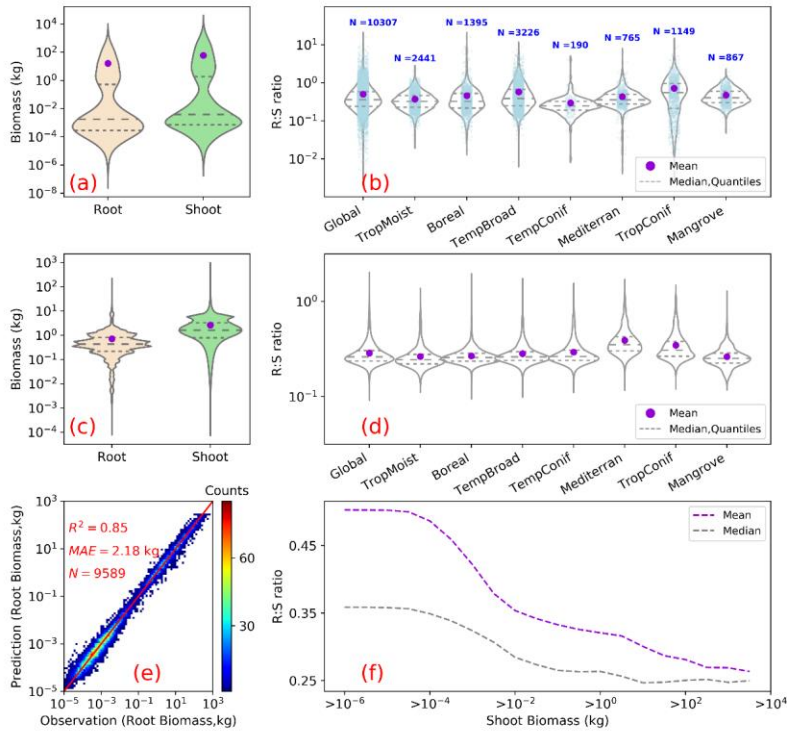
310 Conservancy(Olson and Dinerstein, 2002) and is shown in Supplementary Figure 2.

S2, Tropical systems (Tr): Biomes 1,6,11; Temperate systems (Te): Biomes 4,5; Boreal systems (Bo): Biome 2.

S3, Estimation based on allometric equations and the global above-ground biomass dataset from ref(Santoro, 2018b). See Supplementary Table 7 for details.

[#] This value is converted from 94 Pg C to 188 Pg biomass assuming a 50% carbon content of tree biomass.

315 [*] RD$_{S1}$, the relative difference of Tr + Te + Bo between this study (S1) and previous quantifications. RD$_{S1}$ = (previous study – this study)/this study x 100%. For example, in the column with the head Jackson, RD$_{S1}$ = (200-139)/139*100% = 44%.

[&] RD$_{S2}$, the same as RD$_{S1}$, but with the S2 definition of tropical, temperate and boreal systems.

320 Figure 2. Root biomass and root shoot ratio ($R{:}S$). (a) and (b) show as violin plots the distribution of root and shoot biomass

(in unit of kg/plant) and *R:S* ratios in the raw data used for upscaling. (c) and (d) are the distributions of model-predicted root biomass from this study, of above-ground biomass used for the prediction, and of modelled *R:S* ratios at the global and biome scales. (e) is a heat plot of observed vs. predicted root biomass in kg of root per individual woody plant (see Supplementary Figures 7, 8, 9 for cross-validation at biome, tree size class and continental scales). (f) shows the mean (purple) and median (grey) *R:S* as a function of shoot biomass from observations. A shift of the shoot biomass towards a larger size ((a), (c)) results in a smaller predicted mean *R:S* at the global scale ((b),(d)) (see Supplementary Table 4 for exact values) as the mean *R:S* is size dependent. R^2 is the coefficient of determination, *MAE* is the mean absolute error and *N* is the number of samples. TropMoist: tropical moist forest; Boreal: boreal forest/taiga; TempBroad: temperate broadleaf and mixed forest; TempConif: temperate coniferous forest; Mediterran: Mediterranean forests, woodlands and scrub; TropConif: tropical and subtropical coniferous forest; and Mangrove forest: mangrove forest. Note that the scales of y-axis are different between (a) and (c), (b) and (d). Model training and prediction were conducted on filtered data with *R:S* falling between the 1st and 99th percentiles and shoot biomass matching the range derived from GlobBiomass-AGB(Santoro, 2018b) to reduce impacts from outliers.

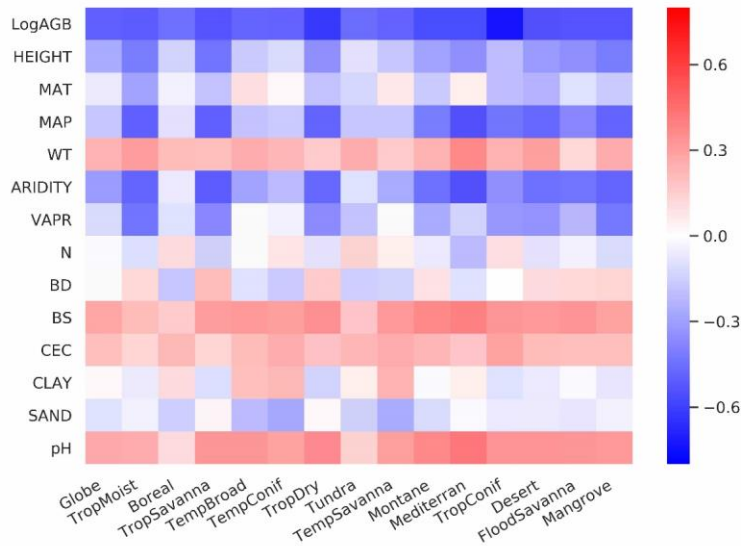



Figure 3. Spearman rank correlations between predictor variables and log-transferred *R:S*. Spearman coefficients are shown at both the global and biome scales for LogAGB: the logarithm of shoot biomass with base 10; HEIGHT: plant height; MAT: mean annual temperature; MAP: mean annual precipitation; WT: water table depth; ARIDITY: the aridity index; VAPR: water vapor pressure; N: soil nitrogen content; BD: soil bulk density; BS: soil base saturation; CEC, soil cation exchange capacity; CLAY: soil clay content; SAND: soil sand content; and pH: soil pH. From left to right, biomes are ordered by decreasing forest areas (Supplementary Figure 2).

4 Discussion

Our lower estimation of root biomass compared to earlier studies is attributable to differences in forest area (Supplementary Table 5), above-ground biomass density (Supplementary Table 5), root biomass measurement and upscaling methodology. For example, the forest area in temperate zones used in Jackson et al. (1997) was about one third higher than in this study. Using the root biomass density (Supplementary Table 5) and estimation method from Jackson et al. (1997), but using the updated forest area map from this study, we estimated total root biomass of tropical, temperate and boreal forests to be 147 Pg (or 184 Pg if sparse forests in tropical/subtropical/temperate grasslands/savannas and shrublands and tundra region are accounted, S1 biome definition in Table 1). This value is smaller than the 200 Pg from Jackson et al. (1997), but still larger than the 121 Pg (or 139 Pg) (Table 1) from our machine learning approach. Our lower values of root biomass compared to Saugier et al. (2001), Mokany et al. (2006) and Robinson (2007) are caused mainly by our lower above-ground biomass density and *R:S* (Supplementary Table 5). Shoot or above-ground biomass density (AGB) of tropical zones is 70% lower in our study than in Robinson (2007) who used sparse plot data collected more than a decade ago (Supplementary Table 5, case S2), and this lower AGB explains 27-46% of our lower root biomass (Supplementary Tables 5, 6). On the other hand, lower biome average *R:S* explains 41-48% of our underestimation compared to Robinson (2007). To elucidate this difference, we calculated weighted biome average *R:S* ratios through dividing total biome-level shoot biomass by root biomass (i.e., weighted mean *R:S*). These weighted mean *R:S,* ranging between 0.19 and 0.31 across biomes (Supplementary Table 3), are generally smaller than the *R:S* values reported in previous studies, which were based on average ratios obtained from sparser data (Supplementary Table 5). Note that the arithmetic means *R:S* (without weighting by biomass) from woody plants located in tropical, temperate and boreal zones (Supplementary Table 4) from our database are close to those from Robinson (2007) (Supplementary Table 5). And our predicted patterns of relatively large *R:S* in regions such as dry forests, savannas and boreal tundra woodlands (Figure 1c) are in line with data compiled from Mokany et al. (2006) and the 2019 IPCC refinement (IPCC2019, 2019). Spawn et al. (2020) estimated root biomass from shoot biomass and the correlation between root/total biomass and temperature. Among previous studies, the recent study from Spawn et al. (2020) shows the smallest difference from our study. After accounting for sparse forests, the 32% smaller estimation from our study is likely linked to differences in the definition of forests and upscaling methodology.

The common practice of estimating root biomass through an average *R:S* without considering the spatial variability of biomass and this ratio[4] is a source of systematic error, leading to overestimating the global root biomass for two reasons.

Firstly, upscaling ratios through arithmetic averages (possibly weighted by the number of trees or area, but not accounting for the fine-grained distribution of biomass) systematically overestimates the true mean $R{:}S$ because $R{:}S$ is a convex negative function of S given by $R{:}S \propto S^{\beta-1}$ with β taking typical values of about 0.9 (Mokany et al., 2006; West et al., 1997, 1999) (see also Supplementary Information: Arithmetic mean $R{:}S$ section). This explains why high-resolution S data

375 used to diagnose weighted mean $R{:}S$ ratios in our approach give generally smaller values than using arithmetic means across grid cells at the biome level (Weighted $R{:}S$ Ratio in Supplementary Table 3 vs. Mean (Gridded) in Supplementary Table 4). Multiplying this biome-level arithmetic mean $R{:}S$ by the average biome-level shoot biomass (Supplementary Table 3) yielded a global forest root biomass of 155 Pg, larger than 142 Pg. Secondly, available measurements tend to sample more small woody plants than big trees compared to real world distributions, because small plants are easier to excavate for

380 measuring roots (see Figure 2a, 2c) but smaller plants tend to have larger $R{:}S$ (Figure 2e, see also Refs(Enquist and Niklas, 2002) (Zens and Webb, 2002)). This sampling bias shifts the $R{:}S$ towards larger values. If we use the biome-level mean $R{:}S$ from our *in-situ* database (Mean (Obs) in Supplementary Table 4), multiplying the shoot biomass (Supplementary Table 3) yielded a global value of 233 Pg, larger than using the mean $R{:}S$ across grid cells through RF (155 Pg). Our RF approach uses *in-situ* data for training but in the upscaling, it accounts for realistic distributions of plant size (Supplementary Figure 5;

385 Supplementary Table 4). We further verified that our upscaled $R{:}S$ ratios are robust to sub-sampling the training data in observed distributions, so that the bias of training data towards small plants does not translate into a bias of upscaled results (see Method, Supplementary Figure 8).

The upscaling approach using allometric equations should also tend to overestimate (see Supplementary Information: Allometric upscaling section) the global root biomass due to the curvature of these allometric functions(Enquist

390 and Niklas, 2002; Zens and Webb, 2002). The global forest root biomass ranges between 154 – 210 Pg when root biomass was upscaled through different allometric equations collected from literature and fitted to our database (Supplementary Table 7), generally larger than from the RF mapping. The global root biomass is likely to be smaller than when applying the allometric equation to the spatial average of shoot biomass (Supplementary Figures 14,15,16,17). Thus, future *in-situ* characterization of the distribution of tree sizes across the world's forests (see Supplementary Information: Allometric

395 upscaling section) would greatly improve root biomass quantification. Note that how well our global estimate reflects the real root biomass is conditioned upon the accuracy of *in-situ* root measurement database used to train our RF model. Under-sampling is a common issue in many root studies due the fractal distribution of root systems in soils and the difficulty of implementing an efficient sampling strategy(Taylor et al., 2013), especially for large trees. We did not quantify the uncertainty of our estimates associated with *in-situ* root measurements due to lack of reliable information.

400 An accurate spatially explicit global map of root biomass helps to improve our understanding of the Earth system dynamics by facilitating fundamental studies on resource allocation, carbon storage, plant water uptake, nutrient acquisition and other aspects of biogeochemical cycles. For example, the close correlation (correlation coefficient: 0.8) between root biomass and rooting depth(Fan et al., 2017) at the global scale and the importance of roots for plant water uptake and transpiration reflect close interactions between vegetation and hydrological cycles. The quest for drivers that affect allocation

405 and consumption of photosynthetic production is a major focus of comparative plant ecology and evolution, as well as the basis of plant life history, ecological dynamics and global changes(McCarthy and Enquist, 2007). Turnover time and allocation are two key aspects that contribute to large uncertainties in current terrestrial biosphere model predictions(Bloom et al., 2016; Friend et al., 2014). Our root biomass map does not provide data on turnover or allocation, but an outcome on their aggregated effects. Future studies combining the root biomass map with upscaled root turnover data could shed light on

410 the allocation puzzle. The growth of the fast turnover part of roots, mostly fine roots, and leaves are highly linked. If we assume an annual turnover of leaves and fine roots, a preliminary estimation of average forest fine root biomass (from leaf biomass) reaches 6.7-7.7 Pg (see Supplementary Information: Preliminary estimation of fine root biomass). Despite being a small portion of total plant biomass and highly uncertain, fine roots are temporally variable and functionally critical in ecosystem dynamics. Future studies on global distribution and temporal dynamics of fine roots are valuable. Considering

415 specific biomes, tropical savannas would benefit from better root biomass estimation due to its large land area, and in tropical dry forests, field measurements of root and shoot biomass are needed to refine root biomass quantifications.

Data availability

Raw datasets and global maps generated in this study are deposited at the open access repository Figshare (https://figshare.com/articles/Supporting_data_and_code_for_A_global_map_of_root_biomass_across_the_world_s_forests/

420 12199637) (Huang et al., 2020). The source data and code underlying Figs 1, 2, 3 Supplementary Figs 2-17 are also provided at the Figshare.

Code availability

Calculations were conducted through Python 2.7.15 and ferret 6.72. The code is deposited at the open access repository Figshare

425 (https://figshare.com/articles/Supporting_data_and_code_for_A_global_map_of_root_biomass_across_the_world_s_forests/ 12199637) (Huang et al., 2020).

Author contributions

Y.H. and P.C. designed this study. Y. H., P.C., M.S., J.C and D.S. collected the data. D.M., P.C., M.S., J.C., Y.C. and Y.H. discussed analyzing methods. Y.H. conducted the analysis and drafted the manuscript. All authors discussed the results and

430 contributed to the manuscript.

Competing interests

The authors declare that they have no conflict of interest.

Acknowledgements

Y.H., D.S.G and P.C. received support from the European Research Council Synergy project SyG-2013-610028

435 IMBALANCE-P and P.C. and Y.H. from the ANR CLAND Convergence Institute. H.Y., D.S. and M. S. were funded through the ESA Climate Change Initiative BIOMASS project. Collecting Russian data were supported by The Russian Science Foundation (project no. 19-77-30015). R.Z.A. received support from the French government grant "Make Our Planet Great Again".

Reference

440 ~~Baccini, A., Walker, W., Carvalho, L., Farina, M., Sulla-Menashe, D., Houghton, R.A. (2017) Tropical forests are a net carbon source based on aboveground measurements of gain and loss. Science 358, 230-233.~~

Bloom, A.A., Exbrayat, J.F., van der Velde, I.R., Feng, L., Williams, M. (2016) The decadal state of the terrestrial carbon cycle: Global retrievals of terrestrial carbon allocation, pools, and residence times. Proceedings of the National Academy of Sciences of the United States of America 113, 1285-

445 1290.

Breiman, L. (2001) Random forests. Machine Learning 45, 5-32.

Cairns, M.A., Brown, S., Helmer, E.H., Baumgardner, G.A. (1997) Root biomass allocation in the world's upland forests. Oecologia 111, 1-11.

Cheng, D.L., Niklas, K.J. (2007) Above- and below-ground biomass relationships across 1534

450 forested communities. Annals of Botany 99, 95-102.

Cheng Meng, Dashuan Tian, Hui Zeng, Zhaolei Li, Yi, C., Niu, S. (2019) Global soil acidification impacts on belowground processes. Environmental Research Letters, Volume 14, Number 7.

Crowther, T.W., Glick, H.B., Covey, K.R., Bettigole, C., Maynard, D.S., Thomas, S.M., Smith, J.R., Hintler, G., Duguid, M.C., Amatulli, G., Tuanmu, M.N., Jetz, W., Salas, C., Stam, C., Piotto, D., Tavani, R.,

455 Green, S., Bruce, G., Williams, S.J., Wiser, S.K., Huber, M.O., Hengeveld, G.M., Nabuurs, G.J., Tikhonova, E., Borchardt, P., Li, C.F., Powrie, L.W., Fischer, M., Hemp, A., Homeier, J., Cho, P., Vibrans, A.C., Umunay, P.M., Piao, S.L., Rowe, C.W., Ashton, M.S., Crane, P.R., Bradford, M.A. (2015) Mapping tree density at a global scale. Nature 525, 201-+.

Enquist, B.J., Niklas, K.J. (2002) Global allocation rules for patterns of biomass partitioning in seed

460 plants. Science 295, 1517-1520.

Falster, D.S., Duursma, R.A., Ishihara, M.I., Barneche, D.R., Fitzjohn, R.G., Varhammar, A., Aiba, M., Ando, M., Anten, N., Aspinwall, M.J., Baltzer, J.L., Baraloto, C., Battaglia, M., Battles, J.J., Bond-Lamberty, B., van Breugel, M., Camac, J., Claveau, Y., Coll, L., Dannoura, M., Delagrange, S., Domec, J.C., Fatemi, F., Feng, W., Gargaglione, V., Goto, Y., Hagihara, A., Hall, J.S., Hamilton, S., Harja, D.,

465 Hiura, T., Holdaway, R., Hutley, L.S., Ichie, T., Jokela, E.J., Kantola, A., Kelly, J.W.G., Kenzo, T., King, D., Kloeppel, B.D., Kohyama, T., Komiyama, A., Laclau, J.P., Lusk, C.H., Maguire, D.A., le Maire, G., Makela, A., Markesteijn, L., Marshall, J., McCulloh, K., Miyata, I., Mokany, K., Mori, S., Myster, R.W., Nagano, M., Naidu, S.L., Nouvellon, Y., O'Grady, A.P., O'Hara, K.L., Ohtsuka, T., Osada, N., Osunkoya, O.O., Peri, P.L., Petritan, A.M., Poorter, L., Portsmuth, A., Potvin, C., Ransijn, J., Reid, D., Ribeiro, S.C.,

470 Roberts, S.D., Rodriguez, R., Saldana-Acosta, A., Santa-Regina, I., Sasa, K., Selaya, N.G., Sillett, S.C., Sterck, F., Takagi, K., Tange, T., Tanouchi, H., Tissue, D., Umehara, T., Utsugi, H., Vadeboncoeur, M.A., Valladares, F., Vanninen, P., Wang, J.R., Wenk, E., Williams, R., Ximenes, F.D., Yamaba, A., Yamada, T., Yamakura, T., Yanai, R.D., York, R.A. (2015) BAAD: a Biomass And Allometry Database for woody plants. Ecology 96, 1445-1445.

475 Fan, Y., Li, H., Miguez-Macho, G. (2013) Global Patterns of Groundwater Table Depth. Science 339, 940-943.

Fan, Y., Miguez-Macho, G., Jobbagy, E.G., Jackson, R.B., Otero-Casal, C. (2017) Hydrologic regulation of plant rooting depth. Proceedings of the National Academy of Sciences of the United States of America 114, 10572-10577.

480 Fick, S.E., Hijmans, R.J. (2017) WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. International Journal of Climatology 37, 4302-4315.

Friend, A.D., Lucht, W., Rademacher, T.T., Keribin, R., Betts, R., Cadule, P., Ciais, P., Clark, D.B., Dankers, R., Falloon, P.D., Ito, A., Kahana, R., Kleidon, A., Lomas, M.R., Nishina, K., Ostberg, S., Pavlick, R., Peylin, P., Schaphoff, S., Vuichard, N., Warszawski, L., Wiltshire, A., Woodward, F.I.
485 (2014) Carbon residence time dominates uncertainty in terrestrial vegetation responses to future climate and atmospheric CO2. Proceedings of the National Academy of Sciences of the United States of America 111, 3280-3285.

GlobalForestWatch, (2019) Aboveground live woody biomass density, https://data.globalforestwatch.org/datasets/8f93a6f94a414f9588ce4657a39c59ff_1.

490 Hagolle, O., Lobo, A., Maisongrande, P., Cabot, F., Duchemin, B., De Pereyra, A. (2005) Quality assessment and improvement of temporally composited products of remotely sensed imagery by combination of VEGETATION 1 and 2 images. Remote Sensing of Environment 94, 172-186.

Hansen, M.C., Potapov, P.V., Moore, R., Hancher, M., Turubanova, S.A., Tyukavina, A., Thau, D., Stehman, S.V., Goetz, S.J., Loveland, T.R., Kommareddy, A., Egorov, A., Chini, L., Justice, C.O.,
495 Townshend, J.R.G. (2013) High-Resolution Global Maps of 21st-Century Forest Cover Change. Science 342, 850-853.

Hastie, T., Tibshirani, R., Friedman, J. (2009) The Elements of Statistical Learning Data Mining, Inference, and Prediction, Second Edition, Section 10.13.2, Springer.

IPCC2019, (2019) 2019 Refinement to the 2006 IPCC Guidlines for National Greenhouse Gas
500 Inventories. vol. 4 (IPCC National Greenhouse Gas Inventories Programme, 2019).

Iversen, C.M., McCormack, M.L., Powell, A.S., Blackwood, C.B., Freschet, G.T., Kattge, J., Roumet, C., Stover, D.B., Soudzilovskaia, N.A., Valverde-Barrantes, O.J., van Bodegom, P.M., Violle, C. (2017) A global Fine-Root Ecology Database to address below-ground challenges in plant ecology. New Phytologist 215, 15-26.

505 Jackson, R.B., Canadell, J., Ehleringer, J.R., Mooney, H.A., Sala, O.E., Schulze, E.D. (1996) A global analysis of root distributions for terrestrial biomes. Oecologia 108, 389-411.

Jackson, R.B., Mooney, H.A., Schulze, E.D. (1997) A global budget for fine root biomass, surface area, and nutrient contents. Proceedings of the National Academy of Sciences of the United States of America 94, 7362-7366.

510 Jiang, Y.T., Wang, L.M. (2017) Pattern and control of biomass allocation across global forest ecosystems. Ecology and Evolution 7, 5493-5501.

Ledo, A., Paul, K.I., Burslem, D., Ewel, J.J., Barton, C., Battaglia, M., Brooksbank, K., Carter, J., Eid, T.H., England, J.R., Fitzgerald, A., Jonson, J., Mencuccini, M., Montagu, K.D., Montero, G., Mugasha, W.A., Pinkard, E., Roxburgh, S., Ryan, C.M., Ruiz-Peinado, R., Sochacki, S., Specht, A., Wildy, D.,
515 Wirth, C., Zerihun, A., Chave, J. (2018) Tree size and climatic water deficit control root to shoot ratio in individual trees globally. New Phytologist 217, 8-11.

Luo, Y.J., Wang, X.K., Zhang, X.Q., Booth, T.H., Lu, F. (2012) Root:shoot ratios across China's forests: Forest type and climatic effects. Forest Ecology and Management 269, 19-25.

McCarthy, M.C., Enquist, B.J. (2007) Consistency between an allometric approach and optimal
520 partitioning theory in global patterns of plant biomass allocation. Functional Ecology 21, 713-720.

Mokany, K., Raison, R.J., Prokushkin, A.S. (2006) Critical analysis of root: shoot ratios in terrestrial biomes. Global Change Biology 12, 84-96.

Moser, G., Leuschner, C., Hertel, D., Graefe, S., Soethe, N., Iost, S. (2011) Elevation effects on the carbon budget of tropical mountain forests (S Ecuador): the role of the belowground compartment. Global Change Biology 17, 2211-2226.

Niklas, K.J. (2005) Modelling below- and above-ground biomass for non-woody and woody plants. Annals of Botany 95, 315-321.

Olson, D.M., Dinerstein, E. (2002) The Global 200: Priority ecoregions for global conservation. (PDF file) Annals of the Missouri Botanical Garden 89:125-126. -The Nature Conservancy, USDA Forest Service and U.S. Geological Survey, based on Bailey, Robert G. 1995. Description of the ecoregions of the United States (2nd ed.). Misc. Pub. No. 1391, Map scale 1:7,500,000. USDA Forest Service. 108pp. -The Nature Conservancy (2003), based on Wiken, E.B. (compiler). 1986. Terrestrial ecozones of Canada. Ecological Land Classification Series No. 19. Environment Canada, Hull, Que. 26 pp. + map.

Pan, Y., Chen, J.M., Birdsey, R., McCullough, K., He, L., Deng, F. (2011) Age structure and disturbance legacy of North American forests. Biogeosciences 8, 715-732.

Poorter, H., Jagodzinski, A.M., Ruiz-Peinado, R., Kuyah, S., Luo, Y.J., Oleksyn, J., Usoltsev, V.A., Buckley, T.N., Reich, P.B., Sack, L. (2015) How does biomass distribution change with size and differ among species? An analysis for 1200 plant species from five continents. New Phytologist 208, 736-749.

Poulter, B.A., Luiz; Andela, Niels; Bellassen, Valentin; Ciais, Philippe; Kato, Tomomichi; Lin, Xin; Nachin, Baatarbileg; Luyssaert, Sebastiaan; Pederson, Niel; Peylin, Philippe; Piao, Shilong; Pugh, Tom; Saatchi, Sassan; Schepaschenko, Dmitry; Schelhaas, Martjan; Shivdenko, Anatoly (2019) The global forest age dataset and its uncertainties (GFADv1.1). NASA National Aeronautics and Space Administration, PANGAEA, https://doi.org/10.1594/PANGAEA.897392.

Reich, P.B., Luo, Y.J., Bradford, J.B., Poorter, H., Perry, C.H., Oleksyn, J. (2014) Temperature drives global patterns in forest biomass distribution in leaves, stems, and roots. Proceedings of the National Academy of Sciences of the United States of America 111, 13721-13726.

Robinson, D. (2004) Scaling the depths: below-ground allocation in plants, forests and biomes. Functional Ecology 18, 290-295.

Robinson, D. (2007) Implications of a large global root biomass for carbon sink estimates and for soil carbon dynamics. Proceedings of the Royal Society B-Biological Sciences 274, 2753-2759.

Santoro, M., Beaudoin, A., Beer, C., Cartus, O., Fransson, J.B.S., Hall, R.J., Pathe, C., Schmullius, C., Schepaschenko, D., Shvidenko, A., Thurner, M., Wegmuller, U. (2015) Forest growing stock volume of the northern hemisphere: Spatially explicit estimates for 2010 derived from Envisat ASAR. Remote Sensing of Environment 168, 316-334.

Santoro, M., Cartus, O., Mermoz, S., Bouvet, A., Le Toan, T., Carvalhais, N., Rozendaal, D., Herold, M., Avitabile, V., Quegan, S., Carreiras, J., Rauste, Y., Balzter, H., Schmullius, C. and Seifert, F. M.. (2018a) A detailed portrait of the forest aboveground biomass pool for the year 2010 obtained from multiple remote sensing observations. Geophysical Research Abstracts, 20, EGU2018-18932, EGU General Assembly 2018.

Santoro, M.e.a. (2018b) GlobBiomass - global datasets of forest biomass. PANGAEA, https://doi.org/10.1594/PANGAEA.894711.

Saugier, B., Roy, J., Mooney, H.A. (2001) Estimations of global terrestrial productivity: converging toward a single number? In: Terrestrial Global Productivity (eds Roy J, Saugier B, Mooney HA), pp. 543–556. Academic Press, San Diego.

Schepaschenko, D., Moltchanova, E., Shvidenko, A., Blyshchyk, V., Dmitriev, E., Martynenko, O., See, L., Kraxner, F. (2018) Improved Estimates of Biomass Expansion Factors for Russian Forests. Forests 9.

Schepaschenko, D., Shvidenko, A., Usoltsev, V., Lakyda, P., Luo, Y.J., Vasylyshyn, R., Lakyda, I., Myklush, Y., See, L., McCallum, I., Fritz, S., Kraxner, F., Obersteiner, M. (2017) A dataset of forest biomass structure for Eurasia. Scientific Data 4.

Shangguan, W., Dai, Y.J., Duan, Q.Y., Liu, B.Y., Yuan, H. (2014) A global soil data set for earth system modeling. Journal of Advances in Modeling Earth Systems 6, 249-263.

Simard, M., Pinto, N., Fisher, J.B., Baccini, A. (2011) Mapping forest canopy height globally with spaceborne lidar. Journal of Geophysical Research-Biogeosciences 116.

Spawn, S.A., Sullivan, C.C., Lark, T.J., Gibbs, H.K. (2020) Harmonized global maps of above and belowground biomass carbon density in the year 2010. Scientific Data 7.

Staver, A.C., Archibald, S., Levin, S.A. (2011) The Global Extent and Determinants of Savanna and Forest as Alternative Biome States. Science 334, 230-232.

Taylor, B.N., Beidler, K.V., Cooper, E.R., Strand, A.E., Pritchard, S.G. (2013) Sampling volume in root studies: the pitfalls of under-sampling exposed using accumulation curves. Ecology Letters 16, 862-869.

Trabucco, A., Zomer, R. (2019) Global Aridity Index and Potential Evapotranspiration (ET0) Climate Database v2. figshare. Dataset.

Tuanmu, M.N., Jetz, W. (2014) A global 1-km consensus land-cover product for biodiversity and ecosystem modelling. Global Ecology and Biogeography 23, 1031-1045.

Wang, Y.Y., Xie, Z.H., Jia, B.H. (2016) Incorporation of a dynamic root distribution into CLM4.5: Evaluation of carbon and water fluxes over the Amazon. Advances in Atmospheric Sciences 33, 1047-1060.

Waring, B.G., Powers, J.S. (2017) Overlooking what is underground: Root:shoot ratios and coarse root allometric equations for tropical forests. Forest Ecology and Management 385, 10-15.

Warren, J.M., Hanson, P.J., Iversen, C.M., Kumar, J., Walker, A.P., Wullschleger, S.D. (2015) Root structural and functional dynamics in terrestrial biosphere models - evaluation and recommendations. New Phytologist 205, 59-78.

West, G.B., Brown, J.H., Enquist, B.J. (1997) A general model for the origin of allometric scaling laws in biology. Science 276, 122-126.

West, G.B., Brown, J.H., Enquist, B.J. (1999) A general model for the structure and allometry of plant vascular systems. Nature 400, 664-667.

Yunjian Luo, Xiaoke Wang, Zhiyun Ouyang, Fei Lu, Liguo Feng, Tao, J. (2018) ChinAllomeTree 1.0.

Zens, M.S., Webb, C.O. (2002) Sizing up the shape of life. Science 295, 1475-1476.

Zhang, Y., Yao, Y.T., Wang, X.H., Liu, Y.W., Piao, S.L. (2017) Mapping spatial distribution of forest age in China. Earth and Space Science 4, 108-116.

605