



1 **Biogeochemical climatology for the Southern Benguela Upwelling System,**

2 **constructed from *in situ* monitoring data**

3

4 Stephanie de Villiers

5

6 Centre for Coastal Paleoscience, Nelson Mandela University, Port Elizabeth, South Africa

7

8 *Correspondence to:* Stephanie de Villiers (steph.devilliers@gmail.com)

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24



1 **Abstract.** An annual and a seasonal biogeochemical climatology had been constructed for the
2 Southern Benguela Upwelling System, from *in situ* data collected along a 12 station
3 monitoring line, sampled at monthly intervals from 2001 to 2012. The monitoring line
4 reaches a maximum offshore distance of almost 190 km, with monitoring station depths
5 ranging from 27 to 1 465 m. In addition to temperature, salinity and oxygen CTD profile
6 data, archived monitoring data for the macro-nutrients (phosphate, nitrate + nitrite, silicate)
7 and chlorophyll-a was evaluated. The climatologies exhibit clear spatial and seasonal
8 variability patterns for all parameters, that yield important insight into the SBUS upwelling
9 cycle. These data sets comprise valuable additions to our knowledge base, and will aid both
10 future modelling efforts and studies of biogeochemical processes in upwelling systems. Data
11 for the constructed climatologies has been made available via the PANGAEA Data Archiving
12 and Publication database at <http://doi.pangaea.de/10.1594/PANGAEA.882218>.

13

14 **Keywords:** Upwelling; Benguela; climatology; biogeochemistry; oxygen; nutrients;
15 chlorophyll-a.

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31



1 Introduction

The world's four Eastern Boundary Upwelling Systems (EBUSs), the California, Humboldt, Canary/Iberian and the Benguela, are regions of intense biological production and ocean-atmosphere exchange of CO₂. Together they account for more than 10% of new oceanic primary production and provide more than 20% of the world's commercial fish catches (Chavez and Toggweiler, 1995; Fennel, 1999; Pauly and Christensen, 1995). In addition, they serve as important sources of nutrients to adjacent oligotrophic subtropical gyres. The strength of the biological pump in these large coastal upwelling systems depends on the availability of nutrients in the photic zone. Nutrient supply and export, in turn, are controlled by complex and climate-sensitive physical forcing factors, such as thermal stratification, coastal wind fields and upwelling intensity (Bakun, 1990; McGowan et al., 1998; Wang et al., 2005; Barross et al., 2014; Bakun et al., 2015; Rykaczewski et al., 2015). Increased stratification (e.g. in response to warmer surface ocean temperatures) will result in decreased nutrient transport into the photic zone, whereas enhanced upwelling intensity (e.g. in response to increased atmospheric temperature gradients) will increase nutrient transport into the photic zone. The latter may also potentially increase offshore transport of nutrients.

In order to understand variability in the productivity of EBUSs, and to predict the potential impact of climate and global change, the relationship between nutrient availability and coastal upwelling processes need to be much better understood than it is at present (Cury and Roy, 1989; Bakun et al., 2010; 2015; Botsford et al., 2006). Coupled physical-biogeochemical models have played a vital role in advancing our understanding of these systems (Skogen, 1999; Qian, 2012; Gutknecht et al., 2013). Such modelling efforts benefit greatly from the availability of climatological data. Data for parameters such as surface ocean temperature and chlorophyll-a levels are relatively easily measured and monitored with remote sensing techniques (Fiúza et al., 1982; Chen et al., 2012; Tim et al., 2015). Construction of comparative climatologies for key biogeochemical parameters such as dissolved oxygen and nutrients, however, require systematic *in situ* sampling. The scarcity of data sets that are appropriate for the construction of biogeochemical climatologies, is problematic. This data gap limits our understanding of upwelling system dynamics (Palacio et al., 2004; Garcia-Reyes et al 2015).



1 The Benguela Upwelling System (BUS) is located along the south-west coast of
2 Africa (Figure 1). It consists of two parts that differ in their mean seasonality, atmospheric
3 drivers and large-scale climate modes (Agenbag and Shannon, 1988; Tim et al., 2015), the
4 Northern Benguela Upwelling Systems (NBUS) and the SBUS. At least eight discrete
5 upwelling cells have been identified within the BUS (Figure 1; Lutjeharms and Meeuwis,
6 1987). The Lüderitz cell, in the NBUS, is the major upwelling cell and displays the highest
7 frequency of occurrence and strength (Lutjeharm and Meeuwis, 1987; Gutknecht et al.,
8 2013). The NBUS and the Lüderitz cell have been the subject of numerous, but irregular,
9 biogeochemical research cruises and studies (Dittmar and Birkicht, 2001; Emeis et al., 2004;
10 Kuypers et al., 2005; van der Plas et al., 2007; Mohrholz et al., 2008; Sohm et al., 2011;
11 Noble et al., 2012; Nagel et al., 2013; Flohr et al., 2014;). The SBUS had been monitored for
12 longer and in a more systematic manner than the NBUS, including along the St. Helena Bay
13 Monitoring Line (SHBML) across the Columbine upwelling cell (Figure 1). The monitoring
14 data for the SBUS, however, had been available to a limited extent (Hutchings et al., 2009;
15 Lamont et al., 2015) and most of the bottle data had not been quality controlled or compiled
16 into a distinct long-term monitoring data set. The latter issue had recently been addressed.
17 This biogeochemical monitoring data have now been used to construct both an annual and a
18 seasonal climatology for the SBUS, accompanied by analogous climatologies for
19 temperature, salinity and oxygen, from CTD data.

20 **2 Data processing**

21 **2.1 Sampling location, frequency and data availability**

22 The location, bottom depth and sampling depths for the 12 monitoring stations along the
23 SHBML are given in Table 1 and illustrated in Figure 1. The sampling frequency was
24 approximately monthly, with the actual cruise occurrences summarized in Table 2. The
25 months of March and September were sampled every year from 2001 to 2012. The other
26 months were sampled at least 9 times during the 12 year monitoring period, with the
27 exception of November, which was sampled only 4 times. The bottle sampling depths used
28 to construct a climatology for dissolved phosphate (PO_4^{3-} or P), nitrate + nitrite ($\text{NO}_3^- + \text{NO}_2^-$,
29 or TN), silicate (SiO_2 or Si) and chlorophyll-a (Chl-a) are listed in Table 1. These depths
30 were sampled routinely; depths that were sparsely sampled were not included in the
31 construction of the climatology, to avoid possible bias. CTD data (T, S and O_2) was
32 condensed to 1 m sampling intervals, for the surface-to-bottom depth ranges indicated in



1 Table 1. The following data was not used in the construction of the climatologies: (i) CTD
2 data from 2009 to 2012, because of concerns about infrequent sensor calibrations since 2009,
3 (ii) Bottle data for 2001 to 2003, because of concerns about nutrient data quality prior to
4 2004. The total number of discrete data points used to construct a climatology for each of the
5 parameters are given in Table 3.

6

7 CTD data and bottle samples were collected according to published protocols and analytical
8 methods (Lamont et al., 2015; Ismail et al., 2015). CTD measurements were carried out
9 using three different vessels and multiple Sea-Bird Electronics SBE 911 systems according to
10 international standards, including Winkler titrations to calibrate dissolved oxygen profiles
11 (Lamont et al., 2015). Bottle samples for nutrient analysis were collected using the same
12 protocols, and analysed with the same equipment and analytical methods (Astoria Analyzer
13 Series 300; Ismail et al., 2015), over the monitoring period. It is difficult to definitively
14 establish long-term data uncertainty and accuracy over such an extended period of time, for
15 any of the parameters, not least of all because of intrinsic spatial and temporal variability.
16 The most practical method with which to evaluate data uncertainty, for the purpose of
17 constructing this climatology, is crossover analysis of measurements made at the furthest
18 offshore station (Station 12), at depth (1 350 m), where temporal and spatial variability is
19 expected to be small. Since temporal variability cannot be ruled out, this provides an upper
20 estimate of the uncertainty associated with analytical methodologies. The signal-to-noise
21 ratios (calculated as the average/standard deviation, using all measurements at this depth,
22 over the monitoring period) are as follows: ~ 65 for T, ~ 1634 for S, ~ 24 for O_2 , ~ 6.5 for P,
23 ~ 15.6 for TN and 17.3 for Si. These values suggest that long-term analytical data
24 uncertainty is appropriate for the construction of a climatology.

25

26 **2.2 Data reduction and construction of seasonal and annual climatologies**

27 The monthly monitoring data was first reduced to a seasonal climatology for each parameter
28 (X), by calculating average monthly values for each parameter at each of the 12 stations, at
29 each of the depths provided in Table 1 (results illustrated in Figure 2 and 3). The seasonal
30 climatologies were then used to construct annual climatologies for each parameter (results
31 illustrated in Figure 4, 5 and 6), as described below.

32



1 At each of the 12 monitoring stations, every parameter is represented by a set of calculated
2 average monthly values, $X_M[S_i, D_Z]$, at each of the discrete sampling depths listed in Table 1:

3
4 $X =$ T = Temperature ($^{\circ}\text{C}$), S = Salinity (psu), O_2 = Dissolved oxygen ($\text{ml O}_2 \text{ dm}^{-3}$), P =
5 Dissolved phosphate ($\mu\text{mol P dm}^{-3}$), TN = Dissolved nitrate + nitrite ($\mu\text{mol N dm}^{-3}$),
6 Si = Dissolved silicate ($\mu\text{mol Si dm}^{-3}$), or Chl = Chlorophyll-a ($\mu\text{g dm}^{-3}$)

7
8 $S_i =$ Station number (S_1 to S_{12})

9
10 $Y_j =$ Year (Y_{first} to Y_{last} ; 2001 to 2008 for CTD data, 2004 to 2013 for bottle data)

11
12 $M =$ Month (1 to 12, or January to December)

13
14 $D_Z =$ Depth (5 m etc. as listed in Table 1, 1 m increments for CTD data)

15
16 For Station S_i at discrete sampling depth D_i , the seasonal climatology for parameter X
17 consists of the following 12 monthly average values (for January to December):

18
19 $X_{\text{JAN}}[S_i, D_Z]$ $X_{\text{FEB}}[S_i, D_Z]$ $X_{\text{MAR}}[S_i, D_Z]$ $X_{\text{APR}}[S_i, D_Z]$ $X_{\text{MAY}}[S_i, D_Z]$ $X_{\text{JUN}}[S_i, D_Z]$
20 $X_{\text{JUL}}[S_i, D_Z]$ $X_{\text{AUG}}[S_i, D_Z]$ $X_{\text{SEP}}[S_i, D_Z]$ $X_{\text{OCT}}[S_i, D_Z]$ $X_{\text{NOV}}[S_i, D_Z]$ $X_{\text{DEC}}[S_i, D_Z]$

21
22 with $X_M[S_i, D_Z] = \text{AVERAGE}\{X_M[S_i, Y_{\text{first}}, D_Z], \dots, X_M[S_i, Y_{\text{last}}, D_Z]\}$

23
24 The seasonal climatology is then reduced to an annual climatology for each parameter
25 (X_{ANN}), by calculating the average of the twelve monthly $X_M[S_i, D_Z]$ values (at discrete
26 sampling depths, for each of the 12 stations):

27
28 $X_{\text{ANN}}[S_i, D_Z] = \text{AVERAGE}\{X_{\text{JAN}}[S_i, D_Z]; X_{\text{FEB}}[S_i, D_Z], \dots, X_{\text{DEC}}[S_i, D_Z]\}$

29
30 The standard deviation ($X_{\text{sd}}[S_i, D_Z]$) associated with the calculated annual averages represents
31 the intra-annual variability in each parameter, i.e. the magnitude and location of variability:

32
33 $X_{\text{sd}}[S_i, D_Z] = \text{STANDARD DEVIATION}\{X_{\text{JAN}}[S_i, D_Z]; X_{\text{FEB}}[S_i, D_Z], \dots, X_{\text{DEC}}[S_i, D_Z]\}$

34
35 For each parameter X the annual climatology, as a function of depth along the monitoring
36 transect, is represented by a matrix of values:

37 $X_{\text{ANN}}[S_1, 5]$ $X_{\text{ANN}}[S_1, 10]$ $X_{\text{ANN}}[S_1, 23]$
38 $X_{\text{ANN}}[S_2, 5]$ $X_{\text{ANN}}[S_2, 10]$ $X_{\text{ANN}}[S_2, 21]$ $X_{\text{ANN}}[S_2, 30]$
39 $X_{\text{ANN}}[S_3, 5]$ $X_{\text{ANN}}[S_3, 10]$ $X_{\text{ANN}}[S_3, 20]$ $X_{\text{ANN}}[S_3, 30]$ $X_{\text{ANN}}[S_3, 55]$ $X_{\text{ANN}}[S_3, 70]$
40 $X_{\text{ANN}}[S_4, 5]$ $X_{\text{ANN}}[S_4, 10]$ $X_{\text{ANN}}[S_4, 20]$ $X_{\text{ANN}}[S_4, 30]$ $X_{\text{ANN}}[S_4, 55]$ $X_{\text{ANN}}[S_4, 79]$



1 etc.

2

3 The calculated seasonal climatologies for each of the parameters are available as a text file on
4 the PANGAEA database (<http://doi.pangaea.de/10.1594/PANGAEA.882218>).

5

6 **3 Description of the climatologies**

7 **3.1 Annual climatologies**

8 The annual average and intra-annual variability patterns (i.e. standard deviation) for T, S, O₂
9 and the dissolved nutrients are illustrated with cross shelf vertical transects, for the upper 600
10 m, in Figure 4 and 5. The data for chlorophyll-a is plotted to a 40 m depth only (Figure 6). At
11 the surface (5 m depth), temperature ranges from 13.1 to 18.2°C along the 190 km long
12 transect from Station 1 (inshore) to Station 12 (offshore), and salinity from 34.77 to 35.40
13 (Figure 4). Along the bottom, temperature ranges from 11.3 to 3.0°C, and salinity from 34.77
14 to 34.65, in an offshore direction. Surface water nutrient concentrations vary from high
15 inshore to much lower levels offshore: 1.55 to 0.36 $\mu\text{mol dm}^{-3}$ for P, 10.6 to 1.4 $\mu\text{mol dm}^{-3}$
16 for TN and 16.2 to 2.6 $\mu\text{mol dm}^{-3}$ for Si (Figure 4). Surface water chlorophyll-a ranges from
17 values $> 8 \mu\text{g dm}^{-3}$ inshore to $< 1 \mu\text{g dm}^{-3}$ offshore (Figure 6). High chlorophyll-a values (> 3
18 $\mu\text{g dm}^{-3}$) are also restricted to the upper ~ 25 m depth interval at the inshore stations. The
19 highest nutrient values, and lowest oxygen levels, are observed in the bottom waters
20 underlying Stations 2 to 5 (Figure 4). The calculated annual climatology for Stations 1 to 5
21 yield bottom water ranges of 2.0 to 2.5 $\mu\text{mol dm}^{-3}$ for P, 20 to 25 $\mu\text{mol dm}^{-3}$ for TN, and 28 to
22 33 $\mu\text{mol dm}^{-3}$ for Si. Calculated values for the annual dissolved oxygen climatology in these
23 bottom waters fall within the 1 to 2 ml dm^{-3} range.

24

25 Water is known to upwell from a depth of 200 to 300 m in the BUS (Nelson and
26 Hutchings, 1983). The 10°C isotherm shallows from a depth of 293 m at Station 12, to 46 m
27 at Station 4 (Figure 4). If this isotherm is used to trace the flow path of upwelling water, it
28 suggests the following “pre-formed” values (for 10°C water): salinity ~ 34.8 , O₂ $\sim 4.1 \text{ ml}$
29 dm^{-3} , P $\sim 1.3 \mu\text{mol dm}^{-3}$, TN $\sim 16 \mu\text{mol dm}^{-3}$ and Si $\sim 12 \mu\text{mol dm}^{-3}$. At Station 1, the
30 shallowest station, the average nutrient content is 1.68 $\mu\text{mol dm}^{-3}$ for P, 13.1 $\mu\text{mol dm}^{-3}$ for
31 TN and 19.0 $\mu\text{mol dm}^{-3}$ for Si. This is much higher than the values for warm offshore surface
32 water (P = 0.40 $\mu\text{mol dm}^{-3}$, TN = 1.80 $\mu\text{mol dm}^{-3}$ and Si = 2.88 $\mu\text{mol dm}^{-3}$ for the upper 30 m



1 at Station 12). The average nutrient content of water at Station 1 is also higher, for P and Si,
2 than the “pre-formed” values at a depth of 293 m at Station 12 detailed above. This indicates
3 enrichment of shallow coastal water in P and Si relative to upwelled water, from either land-
4 based sources or from entrainment of the nutrient-enriched bottom water that is present at
5 Stations 1 to 6.

6

7 There are considerable spatial differences in intra-annual variability for all the
8 parameters, expressed as the standard deviation associated with calculated annual average
9 values (Figure 5 and 6). However, physical and biogeochemical parameters exhibit distinctly
10 different patterns. Intra-annual variability in temperature and salinity is highest within the
11 upper 100 m of the water column at the offshore stations, while oxygen and the nutrients are
12 more variable at the inshore stations and at sub-surface to bottom water depths. Chlorophyll-
13 a is most variable at shallow depths close to the coast (Figure 6). A detailed evaluation of
14 these zones of high variability, and discussion of the factors responsible, is not the focus of
15 this data discussion. Suffice it to say that areas of high intra-annual variability in temperature
16 and salinity are most likely associated with seasonal changes in the depth of the thermocline
17 and intrusion of mesoscale features, such as warm, salty Agulhas rings. High variability in
18 the biogeochemical parameters are most likely the result of the seasonal cycle of organic
19 matter production and regeneration.

20

21 3.2 Seasonal climatologies

22 The January to December seasonal climatologies exhibit temporal and spatial variability, for
23 all of the parameters, that are typical of coastal upwelling systems (Figure 2, 3 and 8). The
24 seasonal upwelling cycle is most pronounced at Stations 2, 3 and 4 (Figure 7). The 10°C
25 isotherm in the temperature climatology again serves as a useful indicator of the evolution of
26 the seasonal upwelling cycle. At Stations 4 to 12 the 10°C isotherm is present throughout the
27 water column all year round (Figure 2). At Station 4, the 10°C isotherm reaches its
28 maximum depth (~ 60 m) mid-winter (July), starts to shoal in August and reaches its
29 shallowest depth (~ 30 m) mid-summer (January) (Figure 7). At Station 3, the 10°C isotherm
30 makes its appearance in the bottom waters in August, in September at Station 2, and in
31 October at Station 1. This suggests the timing and evolution of the seasonal upwelling cycle.
32 The deeper 9°C isotherm starts to shoal towards shallower depths even earlier, during the
33 months of May and June, according to the seasonal climatology at Station 4 (Figure 7).



1 Another pronounced seasonal change evident in the physical water mass parameters, is the
2 movement of an upper ocean warm and salty ($S > 34.9$) water mass, that migrates closer to
3 shore during the winter months and that is located further offshore in summer (Figure 2).

4

5 The mixing of warm, oxygen-rich and nutrient-depleted surface water to deeper depth
6 during winter is a prominent feature in the seasonal climatologies at Stations 2 to 4 (Figure
7 7). This is followed by the upwelling of nutrient-rich, oxygen-depleted, bottom water, from
8 September onwards. Surface water chlorophyll-a values start to increase with the spring
9 upwelling cycle and is reduced during the winter months (Figure 8). The depth of the
10 chlorophyll-a rich surface layer reaches its deepest depth in early summer, November to
11 December. This period of peak summer productivity evident in the chlorophyll-a climatology
12 is followed by a further reduction of oxygen, and enrichment of nutrients, in the bottom
13 waters at the inshore stations in later summer/early autumn (Figure 7), resulting from the
14 respiration of organic matter.

15

16 **4 Conclusions**

17 Annual and seasonal climatologies constructed from long-term monitoring data exhibit well-
18 resolved spatial and temporal changes. This demonstrates the importance of long-term
19 monitoring efforts, and illustrates what is achievable with systematic sampling and
20 measurement strategies, even for biogeochemical parameters such as dissolved nutrients.
21 This data set can be used to optimize future and ongoing monitoring efforts. It contains a
22 wealth of information for the study of biogeochemical cycles and processes in this and
23 analogous upwelling systems, and it should prove invaluable to coupled physical-
24 biogeochemical modelling efforts.

25

26

27

28

29

30

31

32

33



1 **Data availability**

2 Data for the constructed climatologies have been made available via the PANGAEA Data Archiving and
3 Publication database at <http://doi.pangaea.de/10.1594/PANGAEA.882218>.

5 **Author contribution**

6 The author identified the value of the unused archived data sets used in this study, retrieved the data, performed
7 all data rescue tasks such as quality control and data reduction, and conceived of and wrote this manuscript.

9 **Acknowledgements**

10 Numerous present and past employees of the Department of Environmental Affairs and Marine and Coastal
11 Management were involved in sample collection, analysis and data archiving; most notable have been the long-
12 term efforts of Christien Illert, Gavin Tutt and Marcel van der Bergh.

13 **References**

- 14 Agenbag JJ, Shannon LV. A suggested physical explanation for the existence of a biological boundary at
15 24°30'S in the Benguela system. *S.A.J. Mar. Sci.* 6: 119-132 (1988).
16
17 Bakun A. Global climate change and intensification of coastal ocean upwelling. *Science* 247: 198-201 (1990).
18
19 Bakun A, Black BA, Bograd SJ, García-Reyes M, Miller AJ, Rykaczewski RR et al. Anticipated effects of
20 climate change on coastal upwelling ecosystems. *Curr. Clim. Change Rep.* 1: 85-93 (2015).
21
22 Bakun A, Field DB, Redondo-Rodriquez A and Weeks SJ. Greenhouse gas, upwelling-favorable winds, and the
23 future of coastal ocean upwelling ecosystems. *Glob. Change Biol.* 16: 1213-1228 (2010).
24
25 Barros VR, Field DB, Dokken DJ, Mastrandrea MD, Mach KJ, Bilir TE et al. (eds). *Climate Change 2014:*
26 *Impacts, Adaptation, and Vulnerability. Part B: Regional Aspects. Contribution of Working Group II to the*
27 *Fifth Assessment Report of the Intergovernmental Panel on Climate Change.* Cambridge, New York, NY:
28 Cambridge University Press. 688pp. (2014).
29
30 Botsford LW, Lawrence CA, Dreyer EP, Hastings A and Largier J. Effects of variable winds on biological
31 productivity on continental shelves in coastal upwelling systems. *Deep-Sea Res. Pt. II* 53: 3116-3140 (2006).
32
33 Chavez FP, Toggweiler JR. Physical estimates of global new production: The upwelling contribution. In
34 *Dahlem Workshop on Upwelling in the Ocean: Modern Processes and Ancient Records*, Chichester, UK, John
35 Wiley Sons, 313-320 (1995).
36
37 Chen Z, Yan X-H, Jo Y-H, Jiang L, Jiang Y. A study of Benguela upwelling system using different upwelling
38 indices derived from remotely sensed data. *Cont. Shelf Res.* 45: 27-33 (2012).
39



- 1 Cury P, Roy C. Optimal environmental window and pelagic fish recruitment success in upwelling areas. *Can.*
- 2 *J. Fish. Aquat. Sci.* 46: 670-680 (1989).
- 3
- 4 Dittmar T, Birkicht M. Regeneration of nutrients in the northern Benguela Upwelling and the Angola-Benguela
- 5 Front Areas. *S. Afr. J. Sci.* 97: 239-246 (2001).
- 6
- 7 Emeiss K-C, Brüchert V, Currie B, Endler R, Ferdelman T, Kiessling A, Leipe T, Noli-Pearl K, Struck U,
- 8 Vogt T. Shallow gas in shelf sediments of the Namibian coastal upwelling ecosystem. *Cont. Shelf Res.* 24:
- 9 627-642 (2004).
- 10
- 11 Fennel W. Theory of the Benguela upwelling system. *J. Phys. Ocean.* 29: 177-190 (1999).
- 12
- 13 Fiúza AF de G, de Macedo ME, Guerreiro MR. Climatological space and time variation of the Portuguese
- 14 coastal upwelling. *Oceanologica Acta* 5(1): 31-40 (1982).
- 15
- 16 Flohr A, van der Plas AK, Emeis K-C, Mohrholz V, Rixen T. Spatio-temporal patterns of C:N:P ratios in the
- 17 northern Benguela upwelling system. *Biogeosciences* 11: 885-897 (2014).
- 18
- 19 Garcia-Reyes M, Sydeman WJ, Schoeman DS, Rykaczewski R, Black BA, Smit AJ, Bograd SJ. Under
- 20 pressure: climate change, upwelling, and Eastern Boundary upwelling ecosystems. *Front. Mar. Sci.* 2: 109.
- 21 Doi: 10.3389/fmars.2015.00109 (2015).
- 22
- 23 Gutknecht E, Dadou I, Le Vu B, Cambon G, Sudre J, Garçon V, Machu E, Rixen T, Kock A, Flohr A, Paulmier
- 24 A, Lavik G. Coupled physical/biogeochemical modelling including O₂-dependent processes in the Eastern
- 25 Boundary Upwelling System: application in the Benguela. *Biogeosciences* 10: 3559-3591 (2013).
- 26
- 27
- 28 Hutchings L, Van der Lingen CD, Shannon LJ, Crawford RJ, Verheye HMS, Bartholomae CH, van der Plas
- 29 AK, et al. The Benguela Current: an ecosystem of four components. *Prog. Oceanogr.* 83: 15-32 (2009).
- 30
- 31 Ismail HE, Agenbag JJ, de Villiers S, Ximba BJ. Relation between upwelling intensity and the variability of
- 32 physical and chemical parameters in the Southern Benguel Upwelling System. *Int. J. Ocean.*
- 33 *Dx.doi.org/10.1155/2015/510713* (2015).
- 34
- 35 Kuypers MMM, Lavik G, Woebken D, Schmid M, Fuchs BM, Amann R, Jørgensen BB, Jetten MSM. Massive
- 36 nitrogen loss from the Benguela upwelling system through anaerobic ammonium oxidation. *PNAS* 102: 6478-
- 37 6483 (2005).
- 38



- 1 Lamont T, Hutchings L, van den Berg MA, Goschen WS, Barlow RG. Hydrographic variability in the St Helena
- 2 Bay region of the southern Benguela ecosystem. *J. Geophys. Res: Oceans* 120: 2920-2944,
- 3 doi:10.1002/2014JC010619 (2015).
- 4
- 5 Lutjeharms JRE and Meeuwis JM. The extent and variability of South-East Atlantic upwelling. *S.Afr. J. Mar.*
- 6 *Sci.* 5: 51-62, doi:10.2989/025776187784522621 (1987).
- 7
- 8 Mohrholz V, Bartholomae CH, van der Plas AK, HU Lass HU. The seasonal variability of the northern
- 9 Benguela undercurrent and its relation to the oxygen budget on the shelf. *Cont. Shelf Res.* 28: 424-441 (2008).
- 10
- 11 Nagel B, Emeis K-C, Flohr A, Rixen T, Schlarbaum T, Mohrholz V, van der Plas A. N-cycling and balancing
- 12 of the N-deficit generated in the oxygen minimum zone over the Namibian shelf – an isotope-based approach.
- 13 *J. Geophys. Res: Biogeosci.* 118: 361-371 (2013).
- 14
- 15 Nelson G, Hutchings L. The Benguela Upwelling Area. *Prog. Oceanog.* 12: 333-356 (1983).
- 16
- 17 Noble AE, Lamborg CH, Ohnemus DC, Lam PJ, Goepfert TJ, Measures CI, Frame CH, Casciotti KL, DiTullio
- 18 GR, Jennings J, Saito MA. Basin-scale inputs of cobalt, iron, and manganese from the Benguela-Angola front
- 19 to the South Atlantic Ocean. *Limnol. Oceanogr.* 57: 989-1010 (2012).
- 20
- 21 Qian S. Construction of a high resolution biogeochemical climatology for Eastern Boundary Upwelling
- 22 System: Case Study in Canary Upwelling System. Masters Thesis, ETH Zurich (2012).
- 23
- 24 Palacios DM, Bograd SJ, Mendelssohn R, Schwing FB. Long term and seasonal trends in stratification in the
- 25 California Current, 1950-1993. *J. Geophys. Res.* 109: C10016, doi:10.1029/2004JC002380 (2004).
- 26
- 27 Rykaczewski RR, Dunne JP. Enhanced nutrient supply to the California Current Ecosystem with global
- 28 warming and increased stratification in an earth system model. *Geophys. Res. Lett.* 37: L21606.
- 29 doi:10.1029/2010GL045019 (2010).
- 30
- 31 Sohm JA, Hilton JA, Noble AE, Zehr JP, Saito MA, Webb EA. Nitrogen fixation in the South Atlantic Gyre
- 32 and the Benguela Upwelling System. *Geophys. Res. Lett.* 38: 1-6, doi:10.1029/2011GL048315 (2011).
- 33
- 34 Tim N, Zorita E, Hünicke B. Decadal variability and trends of the Benguela upwelling system as simulated in a
- 35 high-resolution ocean simulation. *Ocean Sci.* 11: 483-502 (2015).
- 36
- 37 Van der Plas AK, Monteiro PMS, Pascall A. Cross-shelf biogeochemical characteristics of sediments in the
- 38 central Benguela and their relationship to overlying water column hypoxia. *Afr. J. Marine Sci.* 29: 37-47
- 39 (2007).
- 40



- 1 Wang D, Gouhier TC, Menge BA, Ganguly AR. Intensification and spatial homogenization of coastal
- 2 upwelling under climate change. Nature 518: 390-394 (2015).

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39



1 Figure 1: The Benguela Upwelling System (shaded green area), off the west coast of southern Africa. The
2 upwelling cells identified by Lutjeharms and Meeuwis (1987) are indicated by the stippled yellow lines. The St
3 Helena Bay Monitoring Line (SHBML) is indicated by the line of green circles, representing the 12 monitoring
4 stations; the shelf bathymetry along this line and the sample bottle depths used to construct climatologies for
5 nutrients and chlorophyll-a, are shown in the inset.

6

7 Figure 2: Monthly climatologies for temperature, salinity and oxygen.

8

9 Figure 3: Monthly climatologies for P, N and Si.

10

11 Figure 4: Cross-shelf transects for the annual climatology of the parameters temperature, salinity, dissolved
12 oxygen and the three dissolved nutrients, constructed as outlined in the text. Data is plotted for the upper 600 m
13 only, to accentuate the upper ocean gradients.

14

15 Figure 5: Cross-shelf transects for intra-annual variability, or the standard deviation associated with the annual
16 averages for temperature, salinity, dissolved oxygen and the three dissolved nutrients.

17

18 Figure 6: Annual climatology and intra-annual variability, expressed as the standard deviation, for chlorophyll-
19 a along the SHBML, shown for the upper 40 m of the water column only.

20

21 Figure 7: Seasonal changes, as a function of depth, in temperature, dissolved oxygen, phosphate and nitrate +
22 nitrite, for Stations 2, 3 and 4, constructed as discussed in the text.

23

24 Figure 8: Average chlorophyll-a profiles along the SHBML, illustrated for each month and the upper 50 m of
25 the water column

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40



1

2 **Table 1:** Monitoring station locations, bottom depth, standard bottle sampling depths and CTD depth ranges
 3 used to construct the climatology

4

Station	LAT (°S)	LONG (°E)	Bottom (m)	Offshore distance (km)	Standard bottle depths (m)	CTD depth range (m)
1	32.300	18.311	27	3	5 - 10 - 23	5 - 23
2	32.310	18.273	32	7	5 - 10 - 21 - 30	5 - 30
3	32.332	18.178	77	16	5 - 10 - 20 - 30 - 50 - 70	5 - 70
4	32.374	17.991	107	35	5 - 10 - 20 - 30 - 50 - 79	5 - 100
5	32.416	17.809	152	53	5 - 10 - 20 - 30 - 50 - 73 - 100	5 - 145
6	32.464	17.611	192	73	5 - 10 - 20 - 30 - 50 - 75 - 100 - 150 - 180	5 - 180
7	32.502	17.422	244	92	5 - 10 - 20 - 30 - 50 - 100 - 235	5 - 235
8	32.573	17.194	285	115	5 - 10 - 20 - 30 - 50 - 100 - 200 - 275	5 - 275
9	32.618	16.991	310	134	5 - 10 - 20 - 30 - 50 - 100 - 200 - 305	5 - 305
10	32.661	16.804	392	153	5 - 10 - 20 - 30 - 100 - 370	5 - 370
11	32.706	16.622	560	172	5 - 10 - 20 - 30 - 100 - 540	5 - 540
12	32.745	16.434	1 465	190	5 - 10 - 20 - 30 - 100 - 1 350	5 - 1 350

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26



1

2 **Table 2** SHBML monitoring cruises occurrences, from 2001 to 2012

3

YEAR	JAN	FEB	MAR	APR	MAY	JUN	JUL	AUG	SEP	OCT	NOV	DEC
2001	yes	yes	yes	yes	yes	yes	yes	yes	yes	yes	no	yes
2002	yes	no	yes	yes	yes	yes	no	no	yes	yes	yes	yes
2003	yes	yes	yes	no	yes	yes	yes	yes	yes	yes	no	yes
2004	yes	yes	yes	yes	yes	yes	yes	yes	yes	no	yes	yes
2005	yes	yes	yes	yes	yes	yes	yes	yes	yes	yes	no	yes
2006	yes	yes	yes	yes	yes	yes	no	yes	yes	yes	no	yes
2007	yes	yes	yes	yes	yes	no	yes	yes	yes	yes	no	yes
2008	yes	yes	yes	no	yes	yes	no	yes	yes	yes	yes	no
2009	yes	yes	yes	yes	yes	yes	yes	yes	yes	yes	no	yes
2010	yes	yes	yes	yes	no	yes	yes	yes	yes	yes	no	no
2011	no	yes	yes	no	yes	yes	yes	yes	yes	no	no	yes
2012	yes	no	yes	yes	yes	yes	yes	yes	yes	yes	yes	no

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27



Table 3 Number of discrete measurements used to construct a climatology for each of the parameters (as discussed in the text, data from 2004 to 2012 for nutrients and chl-a; and CTD data from 2001 to 2008).

Station	PO_4^{3-}	$\text{NO}_3^- + \text{NO}_2^-$	SiO_2	Chl-a	CTD (T, S and O_2)
1	231	231	231	216	1 536
2	293	293	293	261	2 016
3	406	409	409	287	6 336
4	413	414	414	279	9 216
5	460	460	460	276	13 536
6	483	483	483	284	16 896
7	509	509	509	282	22 176
8	519	519	519	279	26 016
9	516	513	517	276	28 896
10	546	537	550	274	35 136
11	540	535	540	231	51 456
12	543	539	543	237	129 216
TOTAL	5 459	5 442	5 468	3 182	342 432

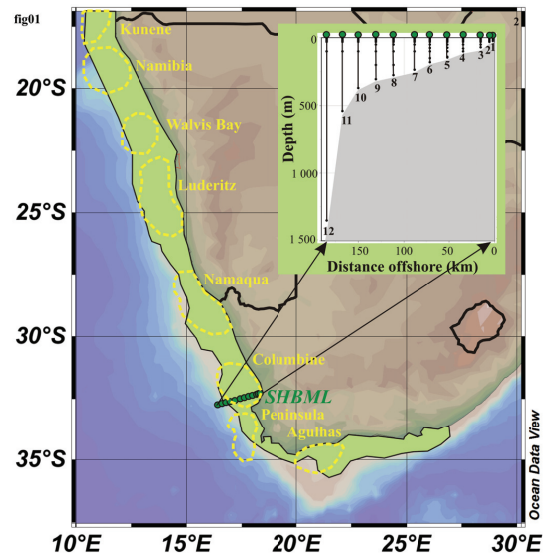




fig02

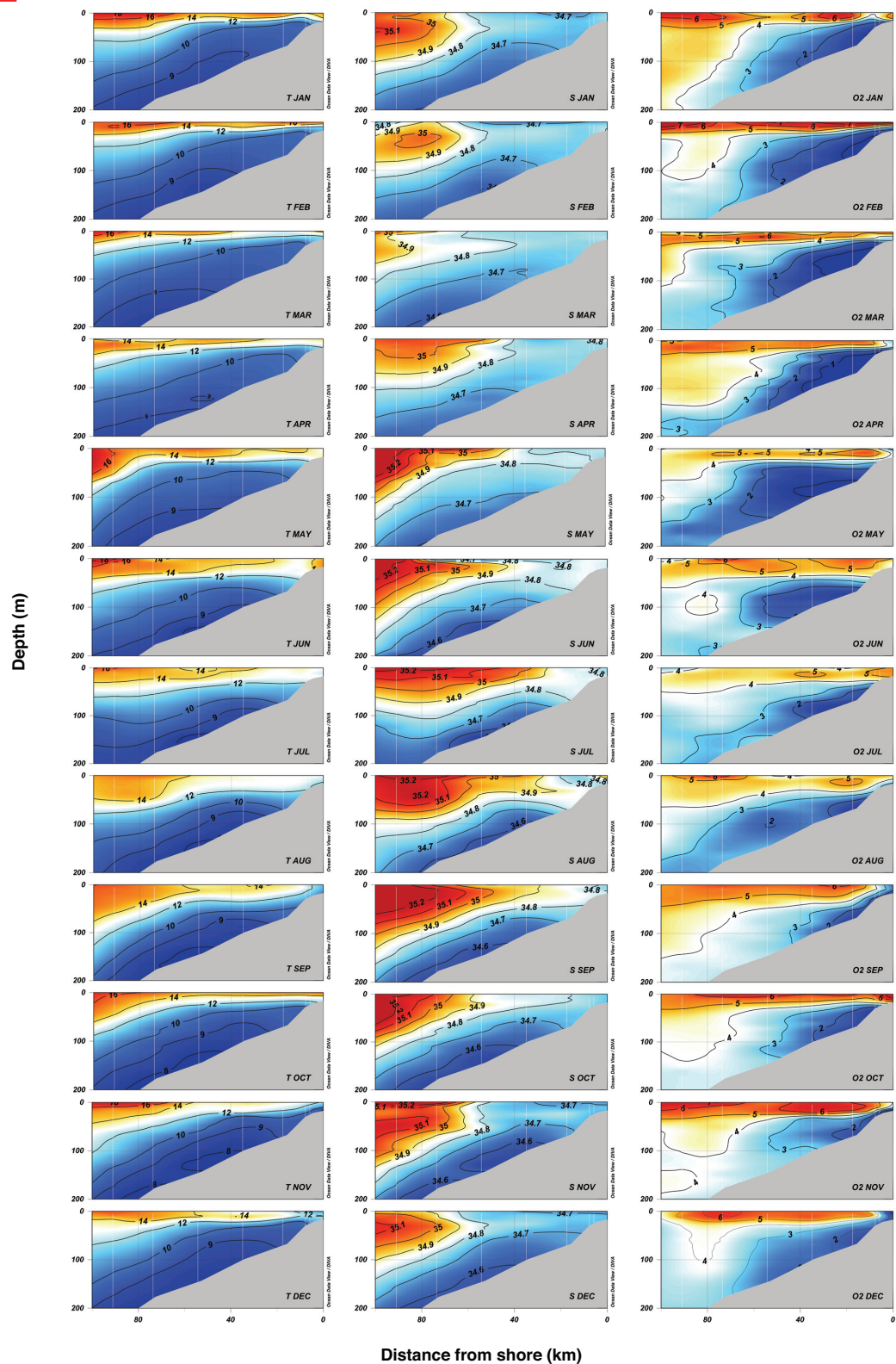




fig03

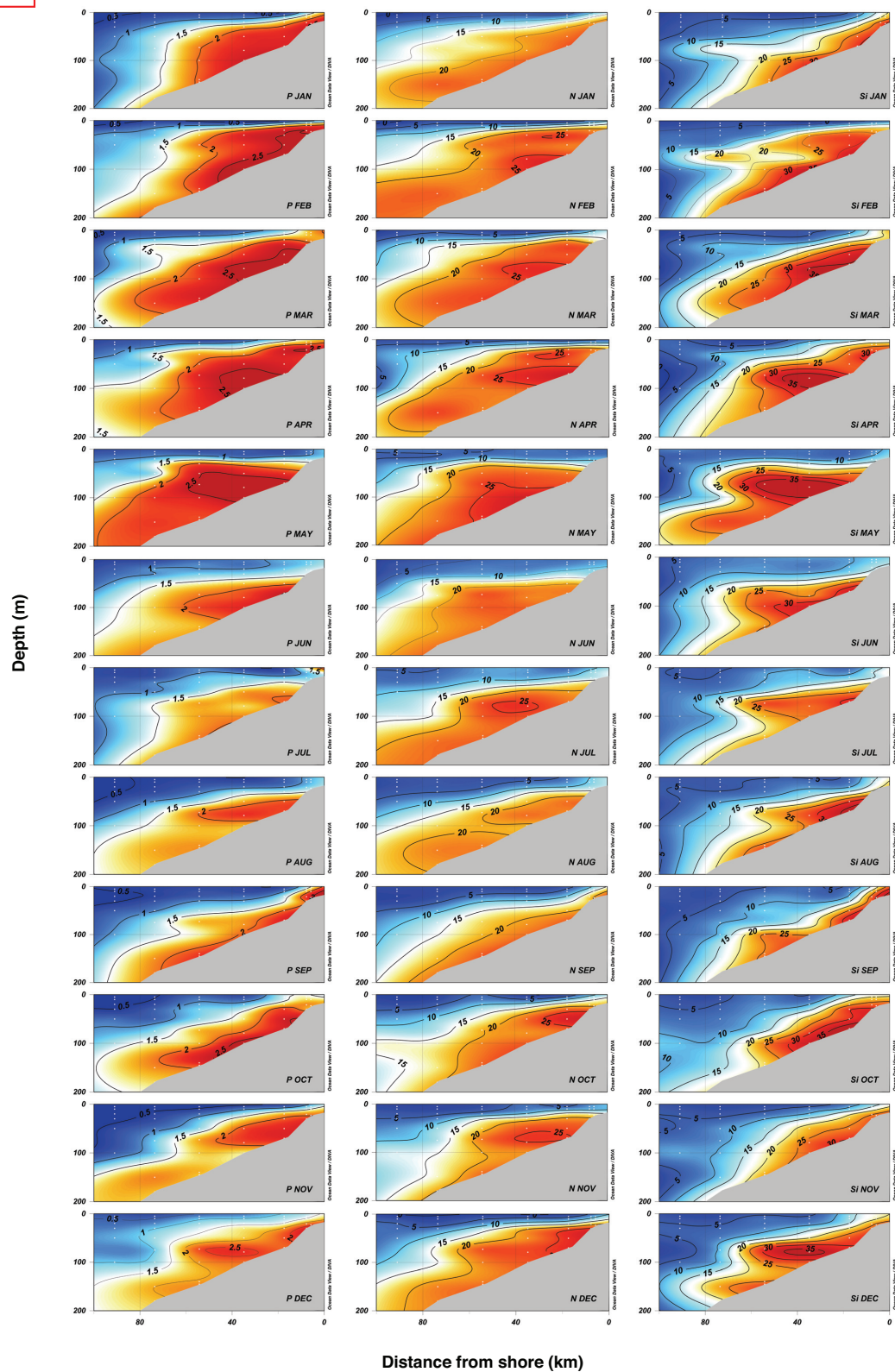




fig04

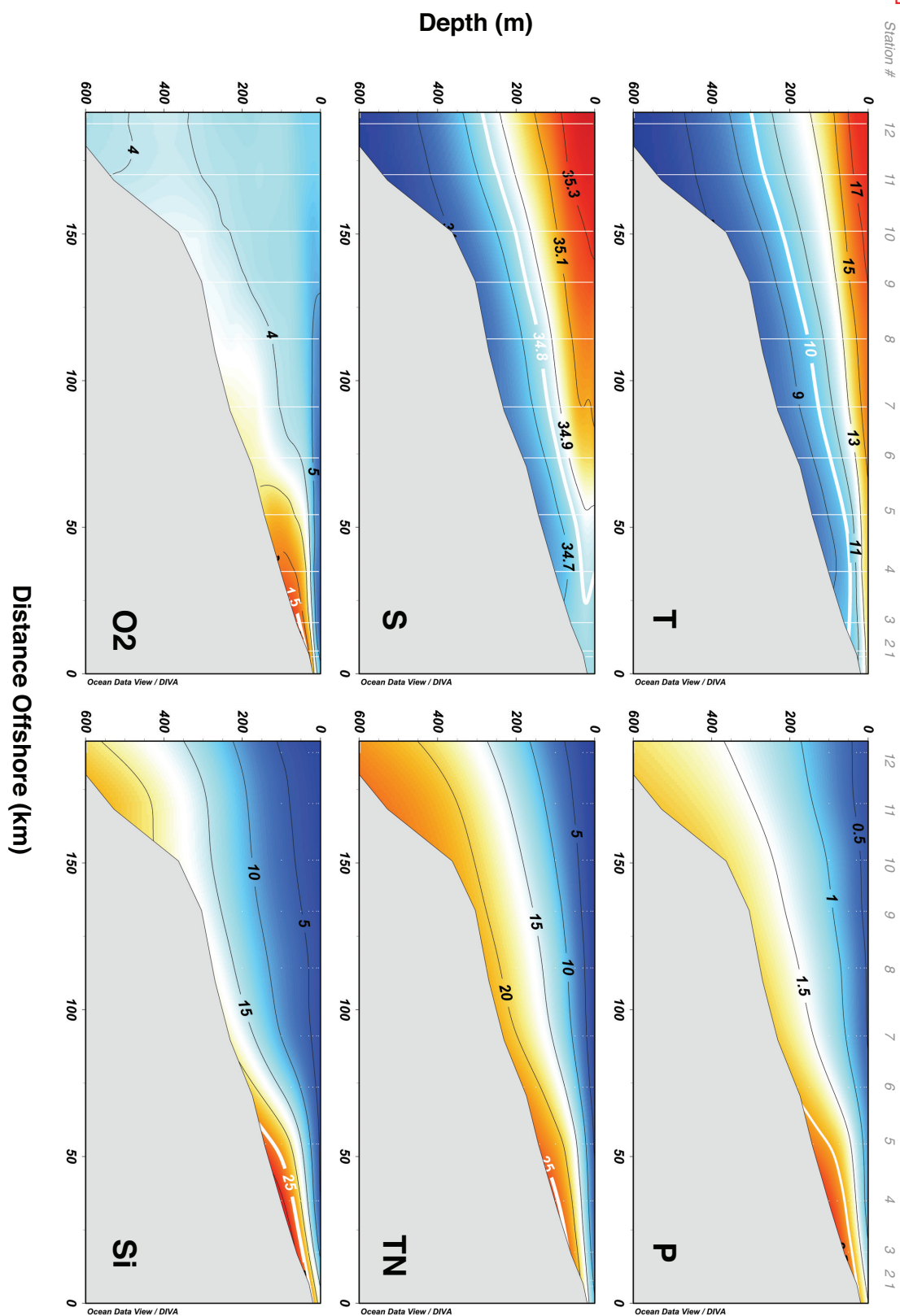




fig05

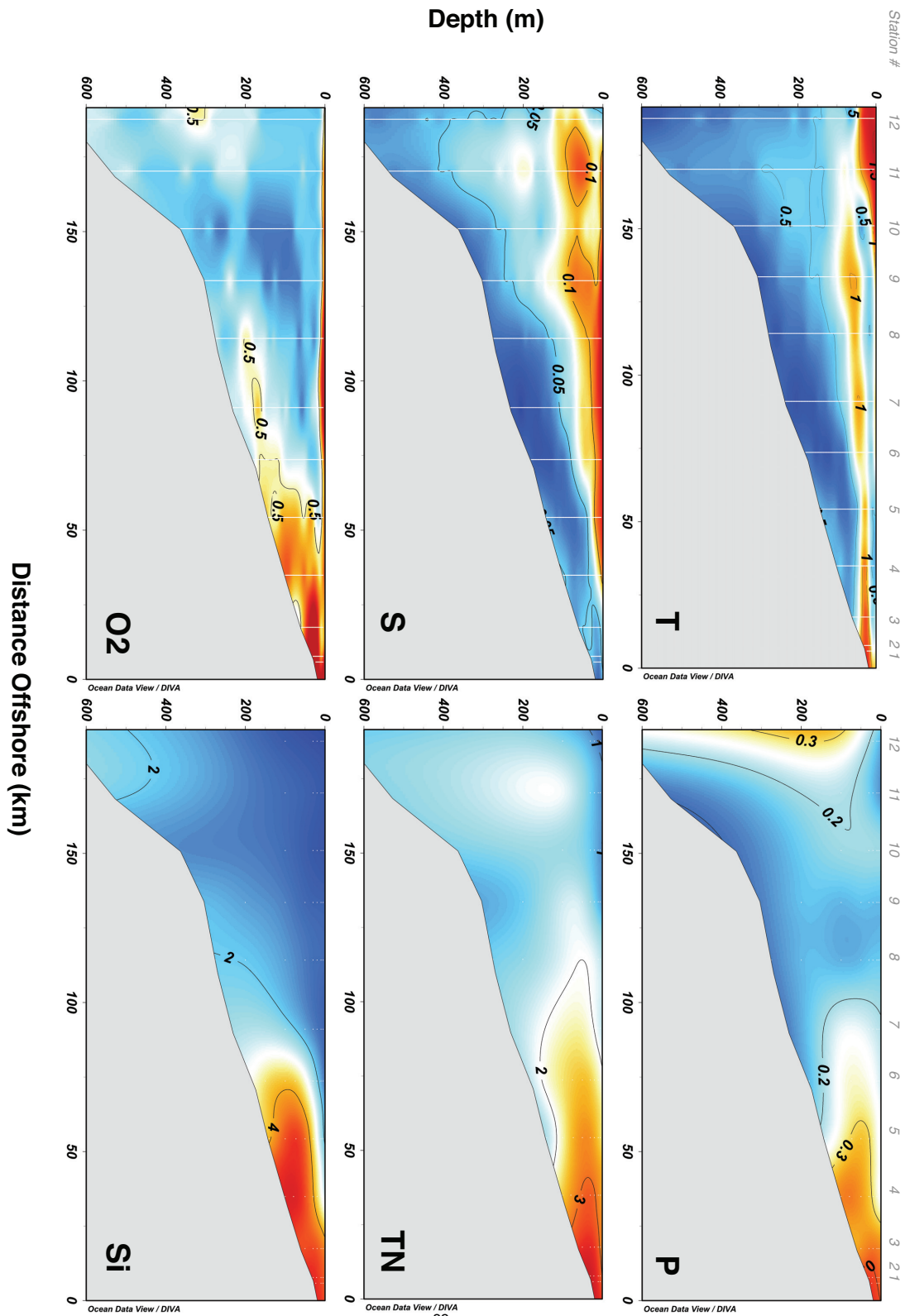




fig06

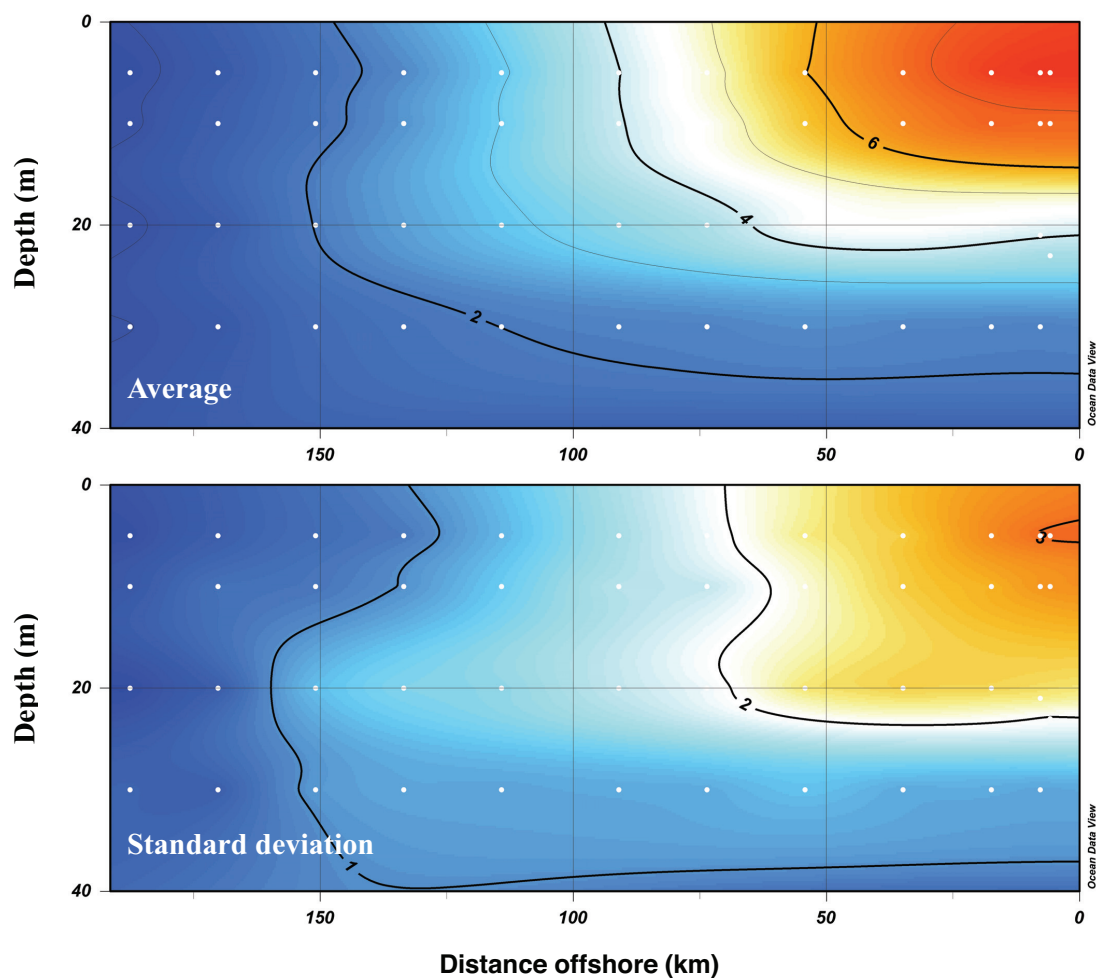




fig07

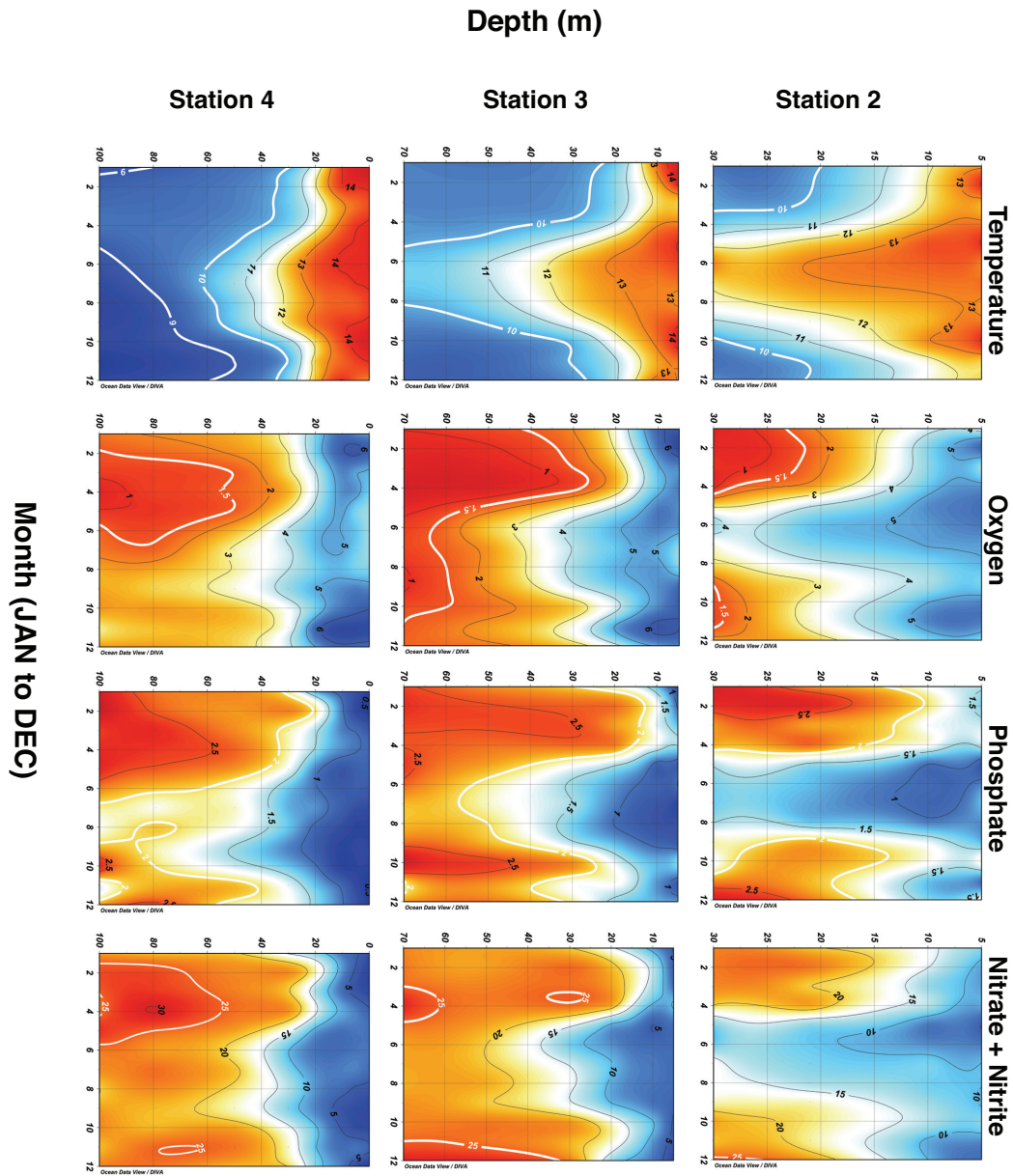




fig08

