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4	Expanding understanding of optical variability in Lake Superior with a four-year dataset
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7	Colleen B. Mouw ^{1*} , Audrey B. Ciochetto ¹ , Brice Grunert ² , Angela Yu ²
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10	,
11	¹ University of Rhode Island, Graduate School of Oceanography
12	215 South Ferry Road
13	Narragansett, RI 02882
14	USA
15	
16	² Michigan Technological University
17	1400 Townsend Drive
18	Houghton, MI 49931
19	USA
20	
21	
22	*Corresponding author, cmouw@uri.edu, +1 401-874-6506
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Abstract

26 Lake Superior is one of the largest freshwater lakes on our planet, but few optical observations

have been made to allow for development and validation of visible spectral satellite remote

sensing products. The dataset described here focuses on coincidently observing inherent and

apparent optical properties along with biogeochemical parameters. Specifically, we observe

30 remote sensing reflectance, absorption, scattering, backscattering, attenuation, chlorophyll

concentration, and suspended particulate matter over the ice-free months of 2013-2016. The

dataset substantially increases the optical knowledge of the lake. In addition to visible spectral

satellite algorithm development, the dataset is valuable for characterizing the variable light field,

particle, phytoplankton, and colored dissolved organic matter distributions, and helpful in food

35 web and carbon cycle investigations, among others. The compiled data can be freely accessed

at: https://seabass.gsfc.nasa.gov/archive/URI/Mouw/LakeSuperior/.

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1. Introduction

Lake Superior is the largest of the Laurentian Great Lakes, the largest lake on Earth by area and third largest lake on Earth by volume. Direct observations of Lake Superior are rare for nearly half of the year due to the long winter climatic conditions of the region. *In situ* observations are scarce November through April, and in many years October and May are also precluded as seasonal storms and ice coverage hinder ship-based observations for long periods of time. As such, remote sensing is an important observational platform for observing the lake.

Remote sensing of inland and coastal waters requires continued investment in *in situ* sampling to fully capture the dynamic range of parameter values and shifting relationships due to response to climate change (Dierssen 2010), anthropogenic stress, and invasive species (Mouw et al., 2015). Remote sensing efforts on the lake initially focused on estimating chlorophyll concentration with band-ratio algorithms (Budd and Warrington, 2004; Li et al., 2004) and met with limited success. More recently, Mouw et al. (2013) evaluated a variety of semi-analytical algorithms to identify the best performing for refinement. They found the retrieval of chlorophyll concentration was not possible via inversion since errors in derived colored dissolved organic matter (CDOM) absorption were greater than the total contribution of phytoplankton to the overall absorption budget. However, they did demonstrate success in retrieving absorption due to CDOM. In light of the challenge to retrieve chlorophyll concentration, Trochta et al. (2015) classified remote sensing reflectance spectra into optical water types and connected the spatial and temporal variability of the water types to physical processes of the lake.

In situ observations are an essential component of remote sensing satellite algorithm development and validation. At the time that Mouw et al. (2013) completed their evaluation of

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algorithms, there were only 8 stations with coincident observations of radiometry, absorption, and scattering for Lake Superior (Peng et al., 2009; Effler et al., 2010; O'Donnell et al., 2013). To enable continued development of remote sensing algorithms for Lake Superior, a greater investment in optical observations were needed. Here we describe a dataset of coincident inherent and apparent optical properties collected throughout Lake Superior over the ice-free months of 2013 to 2016. Inherent optical properties (IOPs, i.e. absorption and scattering/backscattering) depend on in-water dissolved and particulate constituents, while apparent optical properties (AOPs, i.e. reflectance and attenuation) are dependent on both inwater constituents and the ambient light field. Collectively, these properties include remote sensing reflectance, absorption, scattering, backscattering, attenuation, pigment concentrations, and total suspended matter. Beyond satellite algorithm development, the dataset is valuable for characterizing the variable light field, particle, phytoplankton, and colored dissolved organic matter distributions, and helpful in food web and carbon cycle investigations, among others.

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2. Data and Methodology

76 2.1 Study Site

Containing approximately 10% of the Earth's surface freshwater, Lake Superior (Figure 1) is the largest freshwater body on the planet by area $(8.21 \times 10^4 \text{ km}^2)$ and third largest by 78 79 volume $(1.21 \times 10^4 \text{ km}^3)$. The cold temperatures and low nutrient concentrations of Lake 80 Superior (Sterner, 2011), compared to the other Laurentian Great Lakes, result in oligotrophic conditions (e.g. Matheson and Munawar, 1978; Munawar and Munawar, 1973; Weller, 1978; Barbiero and Tuchman, 2001) with low primary production (Sterner, 2010) and low species richness (Hubbs and Lagler, 2004). Superior is the fastest warming of all the Great Lakes

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(Mason et al., 2016), with summer surface temperatures rising faster than the temperature of the surrounding atmosphere (Austin and Allen, 2011; Austin and Colman, 2007; Lenters, 2004).

This trajectory puts Lake Superior close to the upper range of global lake warming trends (O'Reilly et al., 2015). These warming trends impact the lake's energy balance, hydrology, ice cover, and mixing dynamics, which in turn impacts biotic and biogeochemical responses. The previous limited optical observations of Lake Superior reveal that light absorption is dominated

by colored dissolved organic matter (CDOM; ≥75% of the total absorption at 440 nm) (Effler et

al., 2010) and the majority of the backscattering in the lake is attributed to organic rather than

92 minerogenic sources (Peng et al., 2009).

2.2 In Situ Optical Observations

Optical and biogeochemical data are collected in Lake Superior during the ice-free months (May – October) of 2013 through 2016 (Figure 1). The dataset consists of a full suite of coincident IOPs and AOPs. Observed IOPs include absorption due to water itself $(a_w(\lambda), m^{-1})$, phytoplankton $(a_{ph}(\lambda), m^{-1})$, non-algal particles $(a_{NAP}(\lambda), m^{-1})$, colored dissolved organic matter $(a_{CDOM}(\lambda), m^{-1})$, and backscattering due to water itself $(b_{bw}(\lambda), m^{-1})$ and particles $(b_{bp}(\lambda), m^{-1})$. Observed AOPs include downwelling and surface irradiance $(E_d(\lambda))$ and $E_s(\lambda)$, respectively, μW cm⁻²) along with upwelling radiance $(L_u(\lambda), \mu W)$ cm⁻² sr⁻¹), which are used to retrieve remote sensing reflectance $(R_{rs}(\lambda), sr^{-1})$ and downwelling and upwelling diffuse attenuation coefficients $(K_d(\lambda))$ and $K_u(\lambda)$, m⁻¹, respectively. A summary of parameters and units can be found in Table 1.

2.2.1 Apparent Optical Properties

Radiometric measurements are made with three HyperOCR spectral radiometers

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(Satlantic Inc.) that measure between 350–800 nm with approximately 3 nm resolution (137 total wavelengths). In-water $E_d(\lambda)$ and $L_u(\lambda)$ HyperOCR sensors are attached to a free-falling Profiler II frame (Satlantic Inc.), while the $E_s(\lambda)$ sensor is mounted on top of the ship to allow for correction of the other measurements due to changing sky conditions. At each station, the system is deployed for three cast types: surface, multi- and full profile. To characterize the air/water interface, a floatation collar on the profiler frame enables continuous measurement of $L_u(\lambda)$ approximately 20 cm below the water surface for 5 minutes (surface profile). The flotation collar is removed and the profiler then deployed in free-fall mode, measuring 5 consecutive profiles from the surface to 10 m to characterize the near-surface light field (multi-profile). Finally, the profiler is allowed to free-fall to the 1% light level or to within 10 m of the bottom, whichever is shallower (full profile).

All methods and analysis follow the NASA ocean optics protocols for satellite ocean color sensor validation (Mueller et al., 2003a). Briefly, $E_d(\lambda)$, $L_u(\lambda)$ and $E_s(\lambda)$ are dark-corrected and quality controlled to exclude data collected at high instrument tilt (>4°). $E_s(\lambda)$ is smoothed over a 15 s interval and $E_d(t,\lambda)$ and $L_u(t,\lambda)$ normalized to the ratio of $E_s(t_0,\lambda)/E_s(t,\lambda)$ to account for variability in surface irradiance throughout each profile measurement. Data from surface, multi- and full profiles are each binned to 1 m.

Using binned multi-profile data, normalized $E_d(z,\lambda)$ and $L_u(z,\lambda)$ are natural log transformed and fit as a function of depth to retrieve attenuation coefficients $K_d(\lambda)$ and $K_u(\lambda)$, respectively (Smith and Baker, 1984; 1986):

$$ln[X(z,\lambda)] = ln[X(z_m,\lambda)] - (z - z_m)K(z_m) \tag{1}$$

where X is either E_d or L_u, z is depth, z_m is a central reference depth and K is either K_d or K_u.

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128 This method assumes K is uniform across the depth interval used in the regression. For the

majority of the Lake Superior dataset, the upper 8 m of the water column is typically utilized for

the retrieval of K, as recommended by Smith and Baker (1984; 1986). However, some locations

near river outflows show shallow layers with distinct optical changes, reducing the uniform

depth interval to about 5 m.

Using K-regression results, $E_d(\lambda)$ and $L_u(\lambda)$ are propagated to just below the surface

134 $(E_d(0^-,\lambda))$ and $L_u(0^-,\lambda)$. Water leaving radiance $(L_w(\lambda))$ is calculated from $L_u(0^-,\lambda)$ according

135 to:

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$$L_{w}(\lambda) = 0.543 \cdot L_{v}(0^{-}, \lambda) \tag{2}$$

where 0.543 accounts for the net effects of surface reflectance and the refractive index of

seawater as light travels across the sea/air interface (Lee et al., 2002; Mobley, 1994). $R_{rs}(\lambda)$ is

then retrieved as:

$$R_{rs}(\lambda) = \frac{L_w(\lambda)}{E_s(\lambda)} \tag{3}$$

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2.2.2 Inherent optical properties

IOPs are collected via a vertically profiled bio-optical package that measures absorption, attenuation (WET Labs AC-S) and backscattering (WET Labs ECO-BB9) along with concurrent temperature, salinity (SeaBird CTD 37SI) and fluorometeric chlorophyll *a* (WET Labs ECO-FL3). All methods and analysis follow the NASA ocean optics protocols for satellite ocean color sensor validation (Mueller et al., 2003b). The ECO-FL3 is calibrated on a yearly basis, regressing *in situ* chlorophyll *a* fluorescence with extracted chlorophyll *a* concentration ([Chl], mg m⁻³) derived from discrete field samples.

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Total absorption and attenuation $(a_t(\lambda))$ and $c_t(\lambda)$, m^{-1} , respectively) are resolved at 81 wavelengths between 400-750 nm. CTD data are used to correct $a_t(\lambda)$ and $c_t(\lambda)$ for temperature and salinity effects using the coefficients of Sullivan et al. (2006). The AC-S is calibrated before and after each cruise with ultrapure water (ion, particle, and bubble free). This pure-water absorption and attenuation $(a_w(\lambda))$ and $c_w(\lambda)$ is subtracted to retrieve the contribution of non-water components $(a_{nw}(\lambda))$ and $c_{nw}(\lambda)$. Data are corrected for scattering effects using the proportional method of Zaneveld et al. (1994). The bio-optical package is deployed for two successive casts: total and filtered. For the filtered cast, the AC-S intake is fit with a 0.2 μ m filter (PALL, Maxi Capsule Filter 12112) to allow for the measurement of dissolved components only $(a_{CDOM}(\lambda))$, m^{-1} . During deployment, the package is lowered at 0.2 m s⁻¹ or as slow as possible. All data are binned to 1-m during analysis. Particulate absorption $(a_p(\lambda))$, m^{-1} is retrieved by the difference between total and filtered casts $(=a_{nw}(\lambda)-a_{CDOM}(\lambda))$. Pure-water absorption and attenuation as measured by Pope and Fry (1997) is then added to $a_{nw}(\lambda)$ and $c_{nw}(\lambda)$ to return to total values.

The ECO-BB9 measures the volume scattering coefficient (β_t , m⁻¹ sr⁻¹) for an acceptance angle of 124° at 9 wavelengths: 412, 440, 488, 510, 532, 595, 650, 676 and 715 nm. Instrument calibration was performed at WET Labs in Narragansett, RI using 0.1 μ m NIST-traceable beads (Thermo-Scientific, 3100A, bead lot number 43585) at 9°C following the procedure of Sullivan et al. (2013). Processed AC-S measurements are interpolated to BB9 wavelengths and backscattering data corrected for absorption:

$$\beta = \beta_t \, e^{(l-a_t)} \tag{4}$$

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where l is the pathlength of the BB9 (=0.0391 m). The volume scattering function (β_w , m⁻¹ sr⁻¹)

and total scattering (b_w, m⁻¹) of pure-water is retrieved from temperature and salinity

measurements using the model of Zhang and Hu (2009). Backscattering of pure-water (b_{bw}, m⁻¹)

is considered half of total scattering (= $b_w/2$). Volume scattering of the particulate fraction (β_p ,

175 m⁻¹ sr⁻¹) can then be retrieved by difference (= $\beta - \beta_w$). Particulate backscattering is calculated

by taking acceptance angle into account (Sullivan et al., 2013):

$$b_{bp} = 2\pi \chi \beta_p \tag{5}$$

where $\chi=1.076$ for a 124° measurement angle. Total backscattering (b_b, m⁻¹) can then be

retrieved as the sum of particulate and water components (= $b_{bp} + b_{bw}$).

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2.2 Laboratory Analysis of Discrete Water Samples

Whole water samples are taken from multiple depths representative of the surface (5 m)

183 and at a mid-depth of the euphotic zone often coincident with the deep chlorophyll maximum if

one is present (11-50 m). During well-mixed times of year, when a deep chlorophyll layer is not

observed, samples are only taken in the surface layer. Collected water is stored in the dark on ice

and is processed within 8 hours. Discrete water samples are used for determining [Chl], $a_p(\lambda)$,

 $a_{NAP}(\lambda)$, $a_{ph}(\lambda)$, $a_{CDOM}(\lambda)$ and the organic and inorganic components of suspended particulate

matter (SPM₀ and SPM₁, mg L⁻¹, respectively).

Samples for [Chl] and $a_n(\lambda)$ are filtered in triplicate onto Whatman glass fiber filters

190 (GF/F, 0.7μm nominal pore size), stored in liquid nitrogen while at sea and moved to a -80°C

freezer in the lab until analysis. Samples for $a_{CDOM}(\lambda)$ are filtered through an acid-washed 0.2

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contaminants.





μm membrane filter (Whatmann Nucleopore, 111106) and the filtrate stored in clear borosilicate bottles (Qorpak, GLC-01151) in a refrigerator until analysis. Directly prior to each cruise, borosilicate bottles are acid-washed and muffled at 450°C for 4 hours to remove any potential

Spectral CDOM, particulate, non-algal and phytoplankton absorption are measured spectrophotometrically (Perkin-Elmer Lambda 35 UV/VIS dual-beam) for wavelengths between 300-800 nm. Absorption of CDOM filtrate is measured in a 10-cm cuvette following NASA's Ocean Optics Protocols (Mueller et al., 2003b) using a slit-width of 2 nm and a scan rate of 240 nm min⁻¹. For particulate and non-algal absorption, we follow the transmission-reflectance (T-R) method (Tassan and Ferrari ,2002; Lohrenz, 2000; Lohrenz et al., 2003) that utilizes an integrating sphere to correct measurements for the contribution of scattering. Daily blanks are prepared by filtering 25 mL of ultrapure water (ion and particle free) onto a GF/F and used to normalize T-R data during analysis. Filters are placed on a quartz slide at the entrance (transmittance) and exit (reflectance) of the sphere and scanned at a speed of 120 nm min⁻¹ with a slit width of 2 nm both before (a_p) and after (a_{NAP}) exposure to sodium hypochlorite. The difference between these measurements is the contribution due to phytoplankton ($a_{ph} = a_p - a_{NAP}$). Absorption is calculated following Lohrenz et al. (2000).

Extracted [Chl] and phaeopigments are determined following the NASA Ocean Optics Protocols (Trees et al., 2003) for both instrument calibration and sample analysis. Working in the dark, filtered samples are placed in pre-chilled 90% acetone and sonicated for 20 to 30s to break cells open. Sonicated samples are stored in the freezer (-20°C) and allowed to extract for 24 hours prior to analysis. Samples are brought to room temperature and centrifuged at 3000 RPM for 20 minutes to remove filter particulates. Fluorescence is measured before and after the

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215 addition of 50μL of 10% HCl on a Turner Designs 10-AU fluorometer (Optical Kit 10-037R).

Blanks of 90% acetone are measured daily. The fluorometer is calibrated yearly with a dilution

series of pure chlorophyll a (Sigma-Aldrich C6144) extracted in 90% acetone. Field samples are

analyzed within 6 months of fluorometer calibration. [Chl] is retrieved from blank-corrected

219 fluorescence as:

$$[Chl] = \frac{(F_b - F_a)}{F_R} \cdot \frac{\tau}{\tau - 1} \cdot \frac{V_S}{V_F}$$
 (6)

and phaeopigment concentration ([Phaeo], mg m⁻³) calculated as:

$$[Phaeo] = \frac{(F_a \tau - F_b)}{F_R} \cdot \frac{\tau}{\tau - 1} \cdot \frac{V_S}{V_F}$$
 (7)

where F_b and F_a (FSU) are fluorescence measured before and after acidification, respectively, F_R

(FSU (mg m³)⁻¹) is the slope of the fluorometer calibration, τ (unitless) is the acid-ratio of the

fluorometer calibration and V_S and V_F (mL) are the volume of solvent and filtered water sample,

226 respectively.

Both organic and inorganic SPM are quantified following the methodology of the American Public Health Association, American Water Works Association and Water Environment Federation (2005) and the recommendations of Boss et al. (2009) and Woźinak et al. (2011). Whatman GF/F filters (47mm, 0.7μm pore size) are prepared by washing with ~100mL ultrapure water to rinse away any loose glass fibers. They are then dried at 103-105°C for 1 hour and then muffled at 550°C for 4 hours and allowed to cool in a desiccator. Filters are pre-weighed on a high-precision balance (W_F, mg). Duplicate samples are filtered with a minimum of 3L of collected water and stored at -80°C until analysis. Samples are removed from the freezer, dried at 103-105°C for 1 hour and placed in a desiccator to cool. Dried filters are

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236 then weighed within 2 hours of removal from the oven (W_R, mg). Filters are muffled at 550°C

for 1 hour, returned to the desiccator to cool and re-weighed (W_I, mg). Total and inorganic SPM

are retrieved as:

$$SPM = (W_R - W_F)/V \tag{8}$$

$$SPM_{I} = (W_{I} - W_{F})/V \tag{9}$$

241 where V is the volume filtered (L). The organic component can be retrieved by difference

$$(SPM_O = SPM - SPM_I).$$

3. Results

Data are collected from 106 stations from May through October of 2013 to 2016. Collection is aimed at maximizing spatial and temporal variability and dynamic range. Thus, repeat stations are only found for 6 locations. Most of the observations are made in July (n=29) and August (n=28), but also show reasonable distribution in the early spring (May n=12; June n=8) and fall (September n=16; October n=13). Data are only collected in U.S. waters, thus the Canadian waters encompassing the northern reaches of the lake are not observed. To some extent, sampling locations are dependent on the home-port location of research vessels. The western basin (41%) and the central basin, near the Keweenaw Peninsula (41%) of the lake are well observed. The eastern basin is observed only during July of 2015 and a single site in July 2013, August 2014, and September 2013 (Figure 1).

The lake is dimictic (Assel, 1986) and develops a strong thermocline and deep chlorophyll maximum (DCM (Barbiero et al., 2004) in the summer months. The water column is completely mixed in May and June of all years (Figure 2 A, B). A thermocline is evident in

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Depending on location and year, the thermocline either deepened or shallowed in September and October, with evidence of a fully mixed water column in some locations in October (Figure 2 E, F). In the fluorometric [Chl] profiles shown in Figure 2, [Chl] is slightly lower in the near surface water in May and June due to fluorescence quenching when the water column is well mixed (Figure 2 G, H). When [Chl] is derived from absorption, the profiles are uniform at this time of the year (not shown). The formation of a DCM follows the development of the thermocline. As the thermocline sets up in July the DCM develops and becomes more pronounced and deeper in August as the thermocline deepens (Figure 2 I, J). The DCM is still evident in September, but becoming weaker in magnitude and beginning to shallow (Figure 2 K). By October, when the lake is beginning to become fully mixed, [Chl] remains low in the near surface waters and slightly elevated in mid-depths where the DCM was previously found (Figure 2 L). Discreet absorption samples partitioned into water, phytoplankton, non-algal particles and CDOM display contributions from riverine inputs, and broad lake processes near surface at mid-depth (Figure 3). The St. Louis River enters Lake Superior in its western arm. Absorption is fairly evenly distributed there between water, phytoplankton, and CDOM, with some contributions of NAP in May. By June and July, the surface waters are dominated by CDOM and a growing contribution from phytoplankton. This is presumably due the spring melt entering the lake. August sees declining overall absorption as this is a time of lower river flow, with a greater transition toward phytoplankton dominance in September and October when fall storms generate more run off and the river serves as a nutrient delivery conduit to the lake (Figure 3). The surface waters of the central region of the lake remain dominated by water year-round with a

nearly all locations by July (Figure 2 C), becoming stronger and deeper in August (Figure 2 D).

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significant CDOM contribution near the Ontonagon River outlet and within the Keweenaw Peninsula waterway that contains Portage Lake. Samples at mid-depth are not targeted when the water column is well mixed in May and June. The mid-depth becomes progressively dominated by phytoplankton between July and September as the DCM develops and declines. By October, surface and mid-depth absorption are similar (Figure 3). Considering the absorption budget, CDOM is by far the largest contributor (50-80%), followed by phytoplankton, while NAP makes less than a 15% contribution across all stations. Phytoplankton make up a greater percentage of mid-depth absorption, while CDOM is more variable in surface waters. NAP contributions are only slightly less in the mid-depth (Figure 4).

To display the spectral variability observed in the absorption budget, three stations are selected in different regions throughout the lake (Figure 5). As $a_{CDOM}(\lambda)$ absorbs most strongly in the ultraviolet and blue region of the spectrum, its contribution decreases exponentially with increasing wavelength. Phytoplankton also absorb strongly in the blue region of the spectrum, with a peak near 443 nm and a secondary peak at 687 nm. While phytoplankton account for a larger proportion of absorption at mid-depth compared to the surface, they comprise <50% of the total absorption budget in blue wavelengths at all depths (Figure 5). NAP is a larger contributor in the surface, particularly for the sites near river outlets (Figure 5 A, C).

The particulate portion of absorption (NAP and phytoplankton) can be further explored by SPM_i and SPM_o . $a_{NAP}(\lambda)$ includes all non-phytoplankton particles, while SPM_i includes only minerogenic particles. Similar to absorption, greatest SPM loads are found near the outlet of the St. Louis River with magnitude varying with season (Figure 6). The high volumes of spring melt also delivered or stimulated growth resulting in high SPM_o in June. By July, freshwater entering the lake from the river is warm enough to cause a density gradient, where the warm river water

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sat on top of the colder lake water; thus, SPM is significantly lower at the mid-depth. In August, SPM magnitude is relatively similar across the lake and between surface and mid-depth but SPM_i contributed a greater proportion of SPM at depth. By September and October, the depth distribution of SPM_i and SPM_o are similar, but the magnitude of SPM in the western arm is much greater in October due to runoff and water column mixing from fall storms. At this time of year, the riverine water is cooling faster than the lake, resulting in a submerged river plume; thus greatest SPM is observed in the mid-depth.

In addition to the discreet measurements, in situ profiles of $a_n(\lambda)$, $a_{CDOM}(\lambda)$ and $b_{hn}(\lambda)$ are collected. Figure 7 displays examples of this spectral data for the same three stations as in Figure 5. The St. Louis River inflow in the western basin is seen in $a_n(\lambda)$ in the surface of the water column (Figure 7A), and the DCM at mid-depth. The presence of the phytoplankton in $a_p(\lambda)$ is evident by a secondary spectral peak near 683 nm. Riverine impact is also seen where $a_{CDOM}(\lambda)$ is elevated in the near-surface waters but slightly deeper than $a_n(\lambda)$, due to CDOM photobleaching (Figure 7B). In the Ontonagon Outflow, the riverine signal is also seen in $a_n(\lambda)$ and $a_{p}(\lambda)$, with a large deep chlorophyll contribution to $a_{p}(\lambda)$ at mid-depth. $a_{CDOM}(\lambda)$ is elevated below the deep chlorophyll layer, possibly from bloom degradation and sinking (Figures 7 C, D). Along the South Shore, there is no significant riverine source, thus $a_p(\lambda)$ is low in the surface and high at depth associated with the DCM. $a_{CDOM}(\lambda)$ is uniform throughout the water column and significantly lower than the riverine impacted sites (Figures 7 E, F). Spectra are extracted from the profiles at 5 m and 25 m to show an alternative view of the spectral site comparison. Given the low contribution of $a_{NAP}(\lambda)$ to the overall absorption budget (Figure 5), variability in the spectral shape of in situ $a_p(\lambda)$ can be generally attributed to differences in the phytoplankton community between the three locations (Figures 7 G, H). The most distinct difference is the

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basin, which is not present at the other two sites. The variability in the slope of $a_{CDOM}(\lambda)$ is related to the molecular weight of the source carbon (Helms et al., 2008). The slopes of a_{CDOM}(λ) are slightly lower at 5 m than 25 m suggesting proportionately more terrestrial material in surface waters than water at depth (Figures 7 I, J). $b_{bp}(\lambda)$ magnitude indicates the amount of particulate matter in the water, while the slope is related to the size of the particles, with steeper slopes associated with smaller particles (Reynolds et al., 2001). The magnitude of $b_{bp}(\lambda)$ at 5 m is directly proportional to the particle delivery to that location, with the St. Louis River, followed by the Ontonagon River outflow, contributing significant particle loads. The $b_{bo}(\lambda)$ of the western basin and the South Shore are nearly identical at 25 m suggesting a similar size of phytoplankton at these locations. The $b_{bp}(\lambda)$ at the Ontonagon outflow at 25 m is higher and had a greater slope, suggesting a greater abundance of phytoplankton and also a different community composition (Figure 7 K, L). Absorption and scattering processes impact the underwater light field as well as light exiting the water column (i.e. $R_{rs}(\lambda)$). $R_{rs}(\lambda)$ is the parameter that a satellite radiometer observes. Figure 8 displays $R_{rs}(\lambda)$ from in-water measurements at the surface, across all observation months, and light field variability with depth for the three stations also highlighted in Figures 5 and 7. There are groups of similarly shaped $R_{rs}(\lambda)$, with peaks near 490 or 570 nm and varying magnitudes (Figure 8 A-F). The magnitude of $R_{rs}(\lambda)$ is related to particulate abundance. The shape of $R_{rs}(\lambda)$ is primarily related to the variations in the relative contributions and composition of phytoplankton, NAP and CDOM. The spectra with the highest $R_{rs}(\lambda)$ are found in the St. Louis River outflow. The underwater light field also varies spectrally depending on the

presence of a peak at green wavelengths (centered at 550 nm) for the location in the western

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abundance and type of optical constituents. The spectrally dependent depth of light penetration (i.e. $E_d(\lambda)$) as well as the spectral light at depth (ratio of $L_u(\lambda)$ to $E_d(\lambda)$), show significant variability across the three example sites (Figure 8 G-L). The least amount of light is available in the western basin with most available light at 510 nm, while light penetrates the deepest near the South Shore and the most available light is at 490 nm. The diffuse attenuation coefficient at 490 nm ($K_d(490)$) is mapped to show geographic variability (Figure 9). The shortest attenuation lengths (highest $K_d(490)$) are found in the western basin; the greatest lengths are found in the central basin away from land in August.

4. Conclusions

This dataset has increased the coincident optical observations in Lake Superior from 8 observed in July 2006 (Effler et al., 2010) to 106 stations sampled during the ice-free season (May through October) in 2013 to 2016. In addition to greatly expanding the temporal resolution of available data, the spatial (geographic and depth) distribution has grown to cover the western basin, central lake and the southern shore of the eastern basin. In a system that is challenging to observe from direct methods for half the year due to prolonged winter conditions, remote sensing is an important tool. The data presented here are essential for remote sensing algorithm development and validation and this dataset will go a long way toward improving and ensuring science quality satellite products are produced for the interpretation of lake processes. Additionally, this dataset is valuable for characterizing the variable light field, particle, phytoplankton, and colored dissolved organic matter distributions, and helpful in food web and carbon cycle investigations, among others.

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5. Data Availability

The data set contains coincident observations of total, particulate, phytoplankton and CDOM absorption, total scatting, total and particulate backscattering, attenuation, remote sensing reflectance, chlorophyll concentration and total suspended matter from 106 stations, collected between 2013 and 2016. The compiled data are available from the National Aeronautics and Space Administration's SeaWiFS Bio-optical Archive and Storage System (SeaBASS): https://seabass.gsfc.nasa.gov/archive/URI/Mouw/LakeSuperior/ (Mouw et al.,

379 2017).

Author Contributions. Mouw secured resources for the purchase of the optical instrumentation and funding supporting the effort and designed the sampling. Ciochetto maintained the optical instrumentation throughout the project period, quality controlled and achieved the dataset. While all authors contributed toward the fieldwork to collect the observations, Grunert and Ciochetto were responsible for the greatest amount of fieldwork. Ciochetto oversaw the laboratory sample analysis, processed all data and prepared figures. Mouw prepared the manuscript with contributions from all co-authors.

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- support from the Legislative-Citizen Commission on Minnesota Resources are acknowledged for providing the opportunity to collect data aboard the RV Blue Heron. Conversations with Gary Fahnenstiel were helpful to determine sampling strategy.
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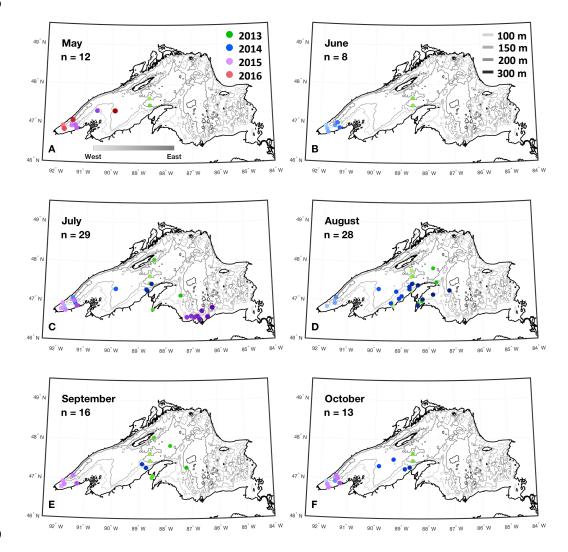
525	Table 1. Summary of parameters.		
526			
527	Found in Dataset		
528	$a_{nw}(\lambda)$	spectral non-water absorption (m ⁻¹)	
529	$a_p(\lambda)$	spectral particulate absorption (m ⁻¹)	
530	$a_{ph}(\lambda)$	spectral phytoplankton absorption (m ⁻¹)	
531	$a_{\mathrm{NAP}}(\lambda)$	spectral non-algal particle absorption (m ⁻¹)	
532	$a_{CDOM}(\lambda)$	spectral colored dissolved organic matter absorption (m ⁻¹)	
533	$b_b(\lambda)$	spectral total backscattering (m ⁻¹)	
534	$b_{bp}(\lambda)$	spectral particulate backscattering (m ⁻¹)	
535	$c_{nw}(\lambda)$	spectral non-water attenuation (m ⁻¹)	
536	$E_d(\lambda)$	downwelling irradiance (μW cm ⁻²)	
537	$E_s(\lambda)$	surface irradiance (μW cm ⁻²)	
538	$L_{u}(\lambda)$	upwelling radiance (μW cm ⁻² sr ⁻¹),	
539	[Chl]	chlorophyll a concentration (mg m ⁻³)	
540	[Pheo]	pheopigment concentration (mg m ⁻³)	
541	SPM	total suspended particulate matter (mg L ⁻¹)	
542	SPM_i	inorganic suspended particulate matter (mg L ⁻¹)	
543	SPM_o	organic suspended particulate matter (mg L ⁻¹)	
544			
545		used in Calculations	
546	$a_t(\lambda)$	spectral total absorption (m ⁻¹)	
547	$a_w(\lambda)$	spectral water absorption (m ⁻¹)	
548	$b_t(\lambda)$	spectral total scattering (m ⁻¹)	
549	$b_{\mathrm{w}}(\lambda)$	spectral water scattering (m ⁻¹)	
550	$b_{bw}(\lambda)$	spectral water backscattering (m ⁻¹)	
551	$c_t(\lambda)$	spectral total attenuation (m ⁻¹)	
552	$L_{\rm w}(\lambda)$	water-leaving radiance (μW cm ⁻² sr ⁻¹)	
553	$R_{rs}(\lambda)$	spectral remote sensing reflectance (sr ⁻¹)	
554	$R(\lambda)$	spectral reflectance (sr ⁻¹)	
555	$K_d(\lambda)$	spectral downwelling diffuse attenuation coefficient (m ⁻¹)	
556	$K_u(\lambda)$	spectral upwelling diffuse attenuation coefficient (m ⁻¹)	
557			
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Figure 1. Station locations for the Lake Superior dataset sampled between 2013-2016 and grouped by month. Locations are colored by year and shaded by longitude from dark to light running east to west as shown in the legend (A). Depth contours shown in grey.

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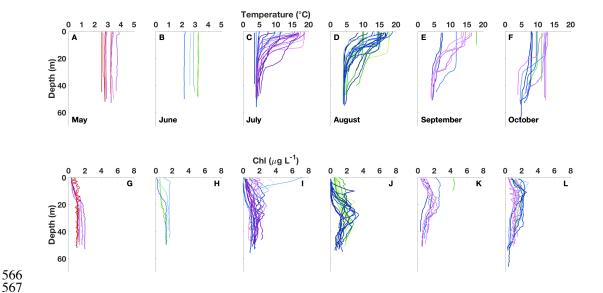


Figure 2. Profiles of temperature (A-F) and [Chl] retrieved from *in situ* fluorescence (G-L) for the Lake Superior dataset. Profiles are colored by year and shaded by longitude as in Figure 1. Data are binned by 1-m. Note compressed x-axis limits for temperature profiles in May and June (A-B). Throughout the region, water column stratification is not observed until July or even August (C-D) and is eroded in the fall (F). Absolute [Chl] levels are low throughout the lake, even during spring and summer months (G-L).

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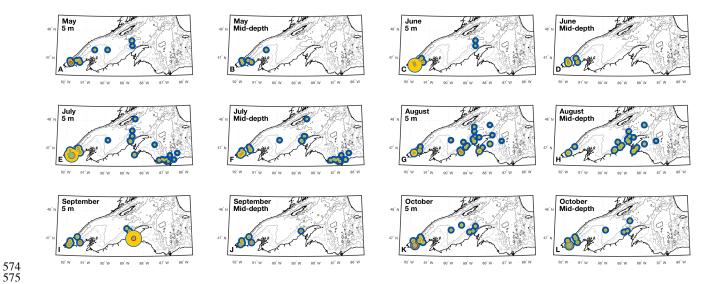


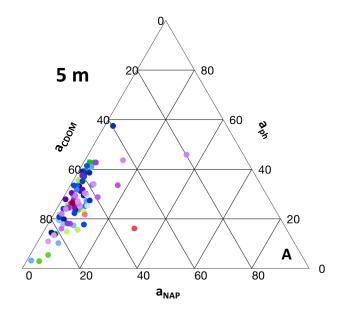
Figure 3. Absorption for discrete water samples taken at 5m and mid-depth throughout Lake Superior grouped by month. The outer diameter for each station marker is relative to total absorption with the contribution of water, CDOM, non-algal particles and phytoplankton indicated by the relative size of concentric circles.

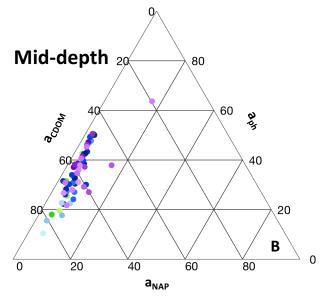
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Figure 4. Ternary plots of the contribution of CDOM, non-algal particles and phytoplankton to total non-water absorption at 443 nm from discrete water samples taken at 5m (A) and mid-depth (B). Markers are colored by year and shaded by longitude as in Figure 1. The system is CDOM dominated, especially near the surface; phytoplankton contributed more than 50% in only 2 cases for the entire dataset.

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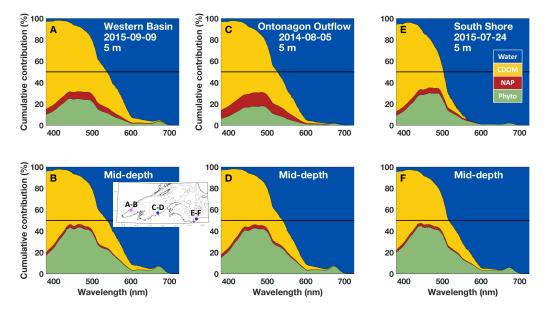


Figure 5. Spectrally resolved contribution of water, CDOM, non-algal particles and phytoplankton to total absorption for discrete water samples taken at 5m (A, C, E) and at middepth (B, D, F) from the Western Basin (A-B, light purple marker on map inset in B), Ontonagon River Outflow (C-D, blue marker on map) and South Shore (E-F, dark purple marker on map) of Lake Superior in the summer of 2014-2015. Black lines indicate 50% cumulative contribution for visual reference.

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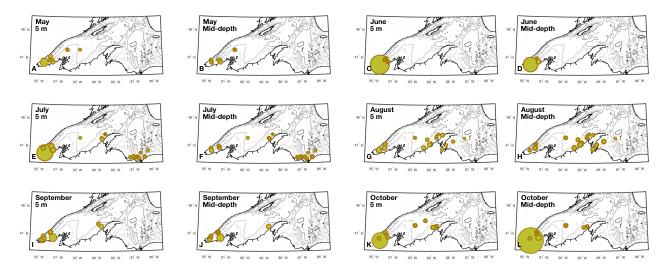


Figure 6. Suspended particulate matter for discrete water samples taken at 5m and mid-depth for the Lake Superior dataset grouped by month. The outer diameter of each station marker indicates total SPM with concentric circles representing the relative contributions of organic and inorganic components.

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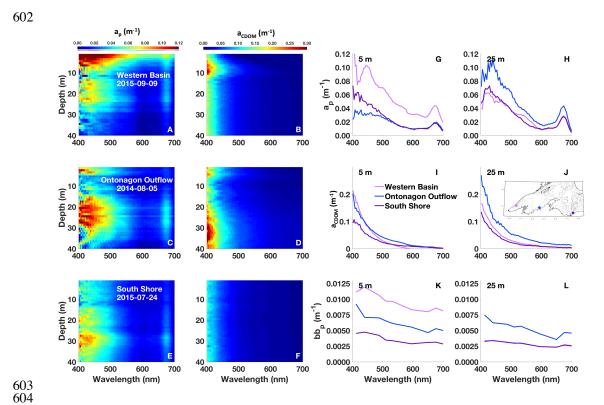


Figure 7. In situ IOP for select stations throughout Lake Superior in the summer of 2014 and 2015 (see inset map in J). Profiles of $a_p(\lambda)$ (A, C, E) and $a_{CDOM}(\lambda)$ (B, D, F) for the Western Basin (A-B), Ontonagon River Outflow (C-D) and South Shore (E-F). Note different color scales for $a_p(\lambda)$ and $a_{CDOM}(\lambda)$ profiles. Examples of individual spectra at 5m and 25m are shown in G-J along with *in situ* particulate backscattering (K-L). Data are binned to 1-m.

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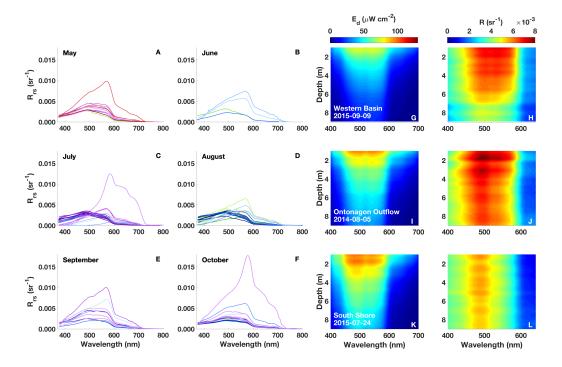


Figure 8. $R_{rs}(\lambda)$ (= $L_w(\lambda)/E_s(\lambda)$) retrieved from combined near-surface multi-cast AOP profiles (A-F). Spectra are colored by year and shaded by longitude as in Figure 1. Example spectral multi-cast AOP profiles for downwelling irradiance ($E_d(\lambda)$) and reflectance ($R(\lambda)=L_u(\lambda)/E_d(\lambda)$) for the three locations from Figures 5 and 7 (G-L).

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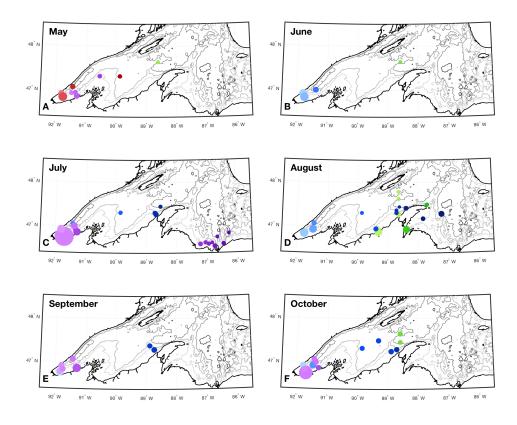


Figure 9. $K_d(490)$ (m⁻¹) for the Lake Superior dataset grouped by month. Stations are colored by year and shaded by longitude as in Figure 1.